

Bioavailability of sediment-associated Cu and Zn to *Daphnia magna*

P.L. Gillis^{a,*}, C.M. Wood^a, J.F. Ranville^b, P. Chow-Fraser^a

^a McMaster University, Hamilton, Ont., Canada L8S 4K1

^b Colorado School of Mines, Golden, CO 80401, USA

Received 27 October 2005; received in revised form 11 January 2006; accepted 13 January 2006

Abstract

Exposures to mining-impacted, field-collected sediment (Clear Creek, CO, USA) contaminated with Cu (2.4 mg/g) and Zn (5.2 mg/g) were acutely toxic to juvenile *Daphnia magna*. Dissolved Cu and Zn in the overlying water (sediment + reference water) were at levels that could cause acute toxicity. To reduce dissolved metals below toxic levels, the sediment was repeatedly rinsed to remove any easily mobilized metals. Washing the sediment reduced dissolved Cu by 60% and Zn by 80%. *D. magna* exposed to washed sediment experienced higher survival (95%) compared to those exposed to the original sediment (<50%). Cu and Zn that remained associated with suspended sediment after washing were not bioavailable, since survival and tissue metal concentrations in *D. magna* exposed to both filtered (>0.45 μm) and unfiltered overlying water were statistically similar. Multiple regression analysis indicated that only dissolved Cu significantly contributed to mortality of *D. magna* whereas particulate Cu, particulate Zn, and dissolved Zn did not. Regression analysis on a combined dataset from all Clear Creek exposures (washed and unwashed), revealed a significant ($p < 0.0001$, $r^2 = 0.76$) relationship between the concentration of dissolved copper in the overlying water and the mortality of exposed *Daphnia*, yielding an estimated LC50 of 26 μg/L dissolved copper (hardness approximately 140 mg/L). The results of this study indicate that if the sediment of Clear Creek was subjected to a resuspension event that there would be a significant efflux of metals from the sediment into the water column, resulting in potentially toxic levels in the water column.

© 2006 Elsevier B.V. All rights reserved.

Keywords: Metal bioavailability; *D. magna*; Sediment associated metals; Labile metals; Copper; Zinc

1. Introduction

Aquatic organisms may be exposed to metals through the dissolved phase and/or through contact with metal-contaminated particles. Previously the dissolved phase was thought to be the main source of metal exposure for planktonic organisms, but recently a number of studies have demonstrated that dietary exposure is also an important route for metal accumulation (Taylor et al., 1998; Wang and Fisher, 1998; Hooke and Fisher, 2001; Adam et al., 2002; Barata et al., 2002; Fisher and Hooke, 2002; Yu and Wang, 2002a,b). These dietary studies have focused on the uptake of metals from organic particles, in particular from metal-contaminated algae, but the uptake of metals from suspended inorganic particles and sediments remains relatively unexplored. A better understanding of the fate of

sediment-associated metals is needed because if dissolved metal concentrations alone are used to predict toxicity, then environmental risk could be underestimated if there is bioavailable metal associated with inorganic particles.

Filter-feeding planktonic organisms such as *Daphnia* may be exposed to sediment-associated metals in two ways. Metals can be released from the sediment into the pore water by changes in geochemical speciation, and then by simple diffusion and convection can move into the overlying water. Metals released in this way will be incorporated into the dissolved phase and depending on the ionic composition of the water are typically considered bioavailable. In addition, metal-contaminated sediment particles may become resuspended into the water column by convection associated with bioturbation and currents. Metals associated with suspended sediment particles have the potential to be ingested by filter-feeding organisms but whether or not they are bioavailable will depend on the nature of the association. Filter-feeding organisms are often exposed to both dissolved and particle-associated metals simultaneously.

As filter-feeders, the preferred food of *Daphnia magna* is algae but any suspended particles over 0.45 μm will be retained

* Corresponding author at: Department of Biology, McMaster University, 1280 Main Street West, Hamilton, Ont., Canada L8S 4K1. Tel.: +1 905 525 9140x23237; fax: +1 905 522 6066.

E-mail addresses: gillisp@mcmaster.ca, gillispatria@hotmail.com (P.L. Gillis).

on the filtering appendages and may be ingested (Brendelberger, 1985). *D. magna* will periodically browse at the sediment–water interface if the availability of suspended food (i.e. algae) falls below a threshold (Horton et al., 1979). In order to overcome periods of food shortage, *D. magna* will actively stir up sediments and detritus by scraping the bottom of shallow waters with their thoracic appendages and then will produce water currents to remove particles from the water (Lampert, 1987). By grazing at the sediment–water interface *Daphnia* combines epibenthic with suspension feeding (Martinez-Madrid et al., 1999). Recently we demonstrated that *D. magna* does ingest sediment, by showing the presence of sediment particles in the gut after exposure in whole sediment bioassays (Gillis et al., 2005).

Based on the limited data available to date, the bioavailability of sediment-associated metals appears to depend on the metal, the composition of the particle and possibly whether the particles were naturally or artificially contaminated. Weltens et al. (2000) reported enhanced accumulation of Cd and higher mortality in *D. magna* exposed to Cd- and Zn-spiked particles than in *D. magna* exposed to only the equivalent dissolved concentration. Weltens et al. (2000) suggested that the physiochemical condition of the gut would favor desorption of metals during the digestion process; therefore, sediment bound particles could be a source of bioavailable metals. In contrast, Erickson et al. (1995) reported that Cu bound to (spiked) particles did not significantly contribute to toxicity in *Ceriodaphnia dubia* any more than exposures to the dissolved phase alone. Ma et al. (2002) demonstrated that toxicity in *C. dubia* decreased with the binding of free Cu (Cu^{2+}) since there was a linear relationship between survival and Cu^{2+} . Weltens et al. (2001) found that suspended solids collected from rivers polluted with a range of metals and organics, released contaminants into the surrounding water (laboratory exposures) creating an acutely toxic environment but there was no further toxicity associated with contaminants that remained bound to the particles.

The overall goal of this study was to determine if the metals associated with field-collected sediment are bioavailable to *D. magna*. This was accomplished by determining the relative contribution of dissolved and particle-associated metals to levels of accumulation in *D. magna* and any observed acute toxicity. Sediment collected from a mining-impacted stream was employed, where preliminary investigations determined that copper and zinc, and not others, were of primary concern for toxicity. Our initial screening of the overlying water revealed that substantial Cu and Zn desorbed from the sediment into the overlying water, resulting in acutely toxic concentrations of Cu and Zn in the overlying water. Therefore, it was necessary to significantly reduce the amount of easily mobilized metals associated with the sediment through a series of rinsings. After exposure to the ‘washed’ sediment demonstrated that dissolved metals had been reduced below acutely toxic levels, the final aim was to separate the dissolved fraction from the particulate fraction to determine which phase was the main source of bioavailable metal. Survival and tissue Cu and Zn concentrations were determined for the exposed organisms and compared with the amounts of dissolved and particle-associated metal in the overlying water.

2. Materials and methods

2.1. Algae cultures

A pure culture of *Pseudokircheriella subcapita* purchased from University of Toronto Culture Collection (Toronto, Ont., Canada) in November 2002, was cultured with Bristol’s medium according to USEPA protocol 16.60049002F (1993). Algae were cultured in 3 L volumetric flasks and held under 24 h light in climate-controlled chambers at $22 \pm 2^\circ\text{C}$.

2.2. *D. magna* cultures

A *D. magna* clone (lot #090600 DM) purchased from Aquatic Research Organisms (Hampton, NH, USA) was held ($20\text{--}22^\circ\text{C}$) in continuous culture according to USEPA protocol 16.60049002F (1993). *Daphnia* were fed a combined diet of yeast, cerophyll and trout chow (YCT) and unicellular algae (*P. subcapita*) daily. Culture media was changed three times per week. Neonates were used to initiate new cultures once a week. Dechlorinated Hamilton city tap water (Lake Ontario) was used as culture water, the overlying water in exposures and the water for gut clearing. This water (herein referred to as reference water) was dechlorinated on site and routinely monitored for chlorine, metals and major ions. Ionic composition in mM: $[\text{Na}^+] = 0.86$, $[\text{Cl}^-] = 1.0$, $[\text{Ca}^{2+}] = 1.0$, $[\text{K}^+] = 0.05$, $[\text{Mg}^{2+}] = 0.2$. Hardness was approximately 140 mg/L (as CaCO_3), pH was 7.8–8 and dissolved organic carbon was approximately 3.0 mg C/L. Background Cu was 2–4.0 $\mu\text{g/L}$ and Zn was $<50 \mu\text{g/L}$.

Rather than *D. magna* neonates, we used 5-day-old juveniles because their larger size provided more tissue for metal analysis. We did not use adults to avoid confounding effects of changes in size and ionic status that accompany brood production and release.

2.3. Sediments

The metal-contaminated sediment used in this study was collected from Clear Creek, CO, USA ($39^\circ44'54''\text{N}$, $105^\circ23'55''\text{W}$). The reference sediment used to dilute the Clear Creek sediment was collected from Long Point, Lake Erie, Ont., Canada ($42^\circ33'54''\text{N}$, $80^\circ02'28''\text{W}$). Bulk sediments were digested using a 1:1 HNO_3 :HCl mixture using trace element grade acid (Fisher Scientific) and then analyzed by inductively coupled mass spectrometry. Sediment metal concentrations are given in Table 1.

Clear Creek is a high-gradient stream which receives metal-rich effluent from a number of mining sites, and has elevated metal levels in both the water and sediments (Table 1). Clear Creek has been designated as a USEPA ‘Superfund site’, indicating that the creek is significantly contaminated and requires study and remediation. Clear Creek streambed sediments are dominated by amorphous iron oxyhydroxide precipitates (i.e. schwertmannite, ferrihydrite, and goethite). These phases coat other detrital silicate minerals and form within the water column as a result of ferrous iron oxidation and precipitation due to upstream inputs of acid mine drainage. The composition of

Table 1
Metal concentrations ($\mu\text{g/g}$) of sediments used in the initial and phase separation exposures

	Sediment											
	Al	Ba	Ca	Cd	Co	Cu	Fe	K	Mg	Mn	Pb	Zn
Initial exposure	10050.7	81.7	4085.0	31.2	22.1	2424.2	141523.7	274.0	494.2	2098.6	225.4	5150.2
Separation experiment ^a	7379.1	100.5	49311.1	9.2	21.4	398.0	44402.1	2101.1	11056.9	1987.3	122.1	1824.6

Both Ag ($<0.24 \mu\text{g/g}$) and As ($<1.8 \mu\text{g/g}$) were below detection.

^a Sediment used in the phase separation experiment was washed and diluted with reference sediment, metal concentrations shown are for the resulting diluted exposure sediment.

the streambed sediments vary seasonally with silicates dominating after high flow periods and iron precipitates during low flow periods (Harvey et al., 2003). The sediment used in this study contains an intermediate amount of the iron-rich precipitate.

Although the concentrations of Cu, Zn, Pb and Cd in Clear Creek sediment were all elevated above what Persaud et al. (1992) designated to be the severe effects level, our initial investigations indicated that only Cu and Zn were released from the sediment in such quantities that the concentrations in the overlying water were within the acutely toxic range for *D. magna* (De Schampelaere et al., 2002; De Schampelaere and Janssen, 2002; Muysen and Janssen, 2001). The concentration of cadmium in the overlying water of the initial exposures (unwashed sediment) was less than $10 \mu\text{g/L}$, whereas Stuhlbacher et al. (1993) reported that in water with similar hardness (170 mg/L) to the water used in this study (140 mg/L), the Cd LC50s for juvenile (3–6-day old) *D. magna* ranged from 49 to $250 \mu\text{g/L}$ depending on the clone used, and similarly, Barata et al. (1998) reported that Cd LC50s for neonates (hardness of 179 mg/L) ranged from 23 to $233 \mu\text{g/L}$, depending on the clone. The concentration of Pb in the overlying water of the unwashed sediment exposures was deemed non-toxic since all concentrations were below $5 \mu\text{g/L}$ and the LC50s for neonate *Daphnia* are reported to be in the range of hundreds of micrograms to milligrams per liter (Biesinger and Christensen, 1972; Chapman, 1980; Fargašová, 1994; Arambašić et al., 1995; Carvalho et al., 1998). Therefore, based on our understanding of the stream, the concentrations of metals in the overlying water of our sediment exposures along with published *Daphnia* metal toxicity data, we chose to focus on the bioavailability and toxicity of Cu and Zn from the Clear Creek sediments.

2.4. *D. magna* exposures: initial exposure

Fig. 1 provides an overview of the study design, and illustrates the sequence of experiments conducted in this study. *Daphnia* exposures were conducted in 250 mL glass beakers at room temperature ($18\text{--}21^\circ\text{C}$) for 48 h without aeration, addition of supplemental food or renewal of overlying water. Reference water was used as overlying water except in treatments where site water was required (initial exposure only). In all cases the water-to-sediment ratio was 4:1. Dissolved oxygen, pH, and dissolved (filtered through an Acrodisc $0.45 \mu\text{m}$ in-line syringe-tip filter) and total (unfiltered) Cu and Zn in the overlying water were measured at initiation and upon completion of the exposure. To determine if the metals associated with the field-collected sedi-

ment were bioavailable, survival and tissue Cu and Zn concentrations were determined for *D. magna* exposed to: (1) reference water alone; (2) site water alone; (3) reference water + sediment; (4) site water + sediment. Ten juvenile *D. magna* were added to each of four replicate beakers. Mortality was recorded at the end of the exposure (48 h). Surviving *D. magna* were transferred to reference water for one hour to purge their gut before being analyzed for tissue metals.

2.5. Sediment ‘washing’

The dissolved concentrations of Cu and Zn in the overlying water in the Initial Exposure were within the range of reported EC50s (Muysen and Janssen, 2001; De Schampelaere et al., 2002; De Schampelaere and Janssen, 2002). Therefore, in order to differentiate toxicity associated with the dissolved fraction from toxicity associated with the particle-bound metals, sediments were subjected to a series of ‘washings’ (using reference water) to remove any easily mobilized metals from the sediment particles. Two methods of washing were compared. In the first (Method A), the sediments that were used in the initial exposure underwent a series of resuspensions by being stirred for 10 min and then allowed to settle for 1 week, after which the over-

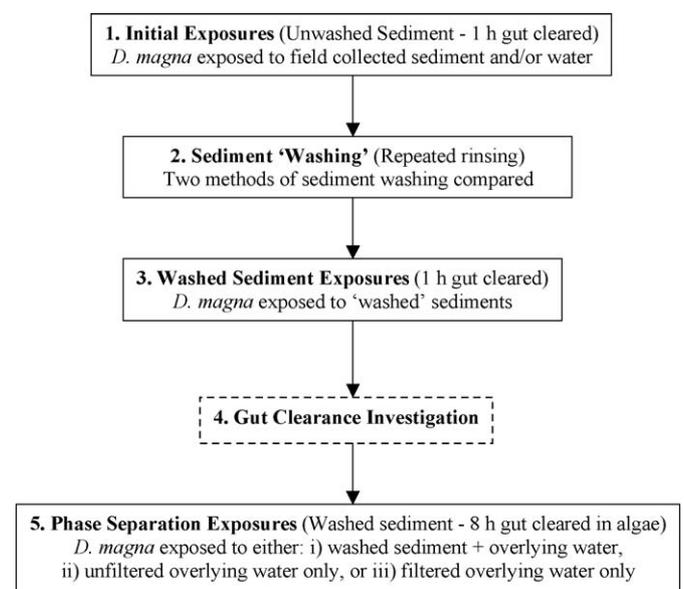


Fig. 1. Flowchart of study design and experiments conducted. Items 1–3, and 5 are part of the current study, item 4 was the subject of a separate investigation with results presented in Gillis et al. (2005).

lying water was replaced. This was repeated three times over a month. In the second (Method B), fresh sediments were subjected to a series of mixing by a Vortex mixer for 5 min followed by centrifugation (3000 rpm for 10 min) and finally replacement of overlying water. This process was repeated six times in the course of a day. In both methods, the overlying water was sampled for dissolved metals after each ‘wash’ before it was gently decanted and replaced with new reference water for the next ‘wash’. The water-to-sediment ratio in the washing vessels was approximately 3:1.

2.6. Washed sediment exposure

Following the removal of the easily mobilized metals from the sediment, *D. magna* were exposed to the washed sediments with the same experimental conditions described for the initial exposure. Survival (48 h) and tissue metal concentrations (1 h gut clearance) were determined for control *D. magna* and those exposed to sediment washed using each of the two methods (A and B).

2.7. Separation of exposure phases

Sediment was washed using Method B (vortex-and-centrifuge) rather than Method A (stir-and-settle), simply because of the reduced time investment required (hours versus weeks). The sediment used in this experiment was collected at a different date than sediment used in the previous experiments. Results from preliminary tests with this new batch of sediment indicated that even after the sediment was washed it was still moderately toxic (30% mortality) to *D. magna*; therefore, to ensure survival in the exposures, the Clear Creek sediment was diluted by 50% with reference sediment before it was used in the exposures. Metal concentrations in the diluted sediment are given in Table 1.

After confirming that dissolved metals in overlying water of exposures with washed and diluted sediment had been reduced below toxic levels, we proceeded to separate any remaining dissolved metals from the particle-associated metals. The term ‘particle-associated metal’ refers to the metals associated with suspended particles $>0.45 \mu\text{m}$, whereas ‘dissolved metal’ refers to any metal that will pass through a $0.45 \mu\text{m}$ filter. In this paper, the concentration of ‘particle-associated metal’ is determined from the difference between total metal (unfiltered) and dissolved metal (filtered).

To each of 15 exposure beakers, 200 mL of reference water and 50 mL of washed sediment were added and left to settle for 1 week (18–21 °C). To create the first treatment (referred to as ‘unfiltered’), approximately 150 mL of the overlying water was removed from a beaker by syringe and transferred to a clean beaker. For the second treatment, overlying water was removed from another beaker as above but was filtered through an in-line syringe-tip filter ($0.45 \mu\text{m}$) before it was transferred to a new beaker (referred to as ‘filtered’). In a third treatment, the beakers (containing sediment and reference water) were used without any manipulation; therefore, the overlying water in this final treatment was also unfiltered. Five replicate beakers were prepared for each treatment.

Ten *D. magna* were added to each exposure vessel and held for 48 h under the same experimental conditions (temperature, without aeration, etc.) described above for the other exposures. Following exposure, *D. magna* were transferred to 200 mL of reference water containing 5×10^5 cells of *P. subcapita* and allowed to purge their guts for 8 h prior to metal analysis. This extended period of purging in the presence of algae was necessitated by closer examination of the *D. magna* from the initial exposure and the washed sediment exposures which revealed that there were still sediment particles in their gut after 1 h of clearance in water. After further investigation into the gut clearance patterns of *Daphnia*, we concluded that only when *D. magna* were held in the presence of algae for 8 h were they able to clear their gut of metal-contaminated sediment particles (Gillis et al., 2005).

2.8. Metal analysis

After gut clearing, *D. magna* from one replicate beaker were combined into a single sample for tissue metal analysis due to minimum requirements for analysis (approximately 0.3 mg). All tissue metal concentrations presented here are whole-body concentrations. Tissues were dried at 60 °C for 24 h, weighed and then digested with 50 μL of concentrated metals grade nitric acid (in 2 mL micro-centrifuge tubes) for 24 h at 60 °C. Samples were brought up to a final volume of 1.5 mL with 1% nitric acid prior to metal analysis. Tissue and water concentrations of Cu were measured with graphite furnace atomic absorption spectroscopy (220, Varian) and Zn concentrations were determined with flame atomic absorption spectroscopy (220 FS, Varian). Method blanks (5) and Fisher Scientific calibration standards (every 20 samples) were included in every run. A maximum of 5% difference between duplicates was accepted. The detection limit for Zn (flame) was 50 $\mu\text{g/L}$ and Cu (furnace) was 2 $\mu\text{g/L}$.

2.9. Statistical analysis

Means are given \pm S.E. Statistical analyses were conducted with the software SPSS version 10.0 and Sigma Stat version 3.0. Comparisons between treatments were made using analysis of variance followed by Tukey’s multiple comparison test to determine differences between treatments ($p < 0.05$). Simple linear regressions and stepwise linear regression were used to determine the relationships between variables (dissolved, particulate, and tissue metal) in the phase separation experiment. Nonlinear regression analyses were used to determine which variables (particulate and dissolved Cu and Zn) contributed significantly to the prediction of *D. magna* survival in a composite dataset from all exposures (initial, washed, and phase separation).

3. Results

3.1. Initial exposure

Dissolved oxygen remained relatively constant throughout the exposure (range 7.0–9.5 mg/L). The pH in the treatments shifted slightly downwards during the exposure from a median

of 7.42 at the beginning of the exposure, to a median of 7.03 at the end of the exposure. The concentrations of Cu (2–3 $\mu\text{g/L}$) and Zn (<50 $\mu\text{g/L}$) in the reference water were significantly lower than in any of the other exposures (e.g. sediment + reference water) (Fig. 2A and B). The concentrations of Cu and Zn in the overlying water (dissolved and total) were significantly ($p < 0.05$) higher in the ‘site water + sediment’ treatment than in either the ‘site water only’ or the ‘reference water + sediment’ treatments, and this suggested that both the sediment and the overlying site water could serve as a source of metals (Fig. 2A and B). In all treatments, dissolved Cu (range 28–50 $\mu\text{g/L}$) accounted for over 60% of the total Cu while dissolved Zn (range 423–1143 $\mu\text{g/L}$) accounted for more than 85% of the total Zn in the overlying water at the beginning of the exposure (Fig. 2A and B, Table 2). Survival was significantly lower (<50%) in treatments containing sediment compared to the site-water-only treatment (>80%) (Fig. 2C). Tissue concentrations of Cu and Zn were about 2–10-fold higher in treatments containing sediment compared to reference water controls (Fig. 2D), suggesting that there was substantial bioavailable metal associated with these sediments. For Zn, tissue levels were similar for the three experimental exposures, but for Cu, tissue levels were significantly higher in the ‘site-water + sediment’ treatment than in the ‘site-water only’ treatment.

3.2. Sediment washing

Cu and Zn levels in the overlying water decreased with repeated washing of the sediment (Fig. 3). After washing, Cu concentrations were reduced to approximately 40% (Method A: stir-and-settle: 38%, Method B: vortex-and-centrifuge 42%) and Zn concentrations reduced to approximately 20% (Method A 20%, Method B 23%) of initial concentrations in the overlying water of unwashed sediments. The final dissolved Cu concentrations were $16(\pm 2.5)$ $\mu\text{g/L}$ and $12(\pm 0.3)$ $\mu\text{g/L}$ after washing with Methods A and B, respectively. The final dissolved concentration of Zn in the overlying water was $89(\pm 21.0)$ $\mu\text{g/L}$ after washing with Method A (stir-and-settle) and $194(\pm 5.6)$ $\mu\text{g/L}$ using Method B (vortex-and-centrifuge).

3.3. Washed sediment exposures

Dissolved oxygen remained constant throughout the exposure (range 8.0–10.0 mg/L). The median pH shifted downward during the exposure, from 7.18 at the beginning to 6.44 at the end of the exposure. Dissolved Cu (10–14 $\mu\text{g/L}$) and Zn (63–80 $\mu\text{g/L}$) in the overlying water of washed sediment were significantly lower than in exposures with unwashed sediment (Fig. 4A and B, Table 3). The concentration of total (unfiltered) Zn and Cu in the overlying water of sediment washed by Method A (stir-and-settle) was significantly reduced compared to total metal levels in the overlying water of unwashed sediment. In contrast, the concentration of total Zn in the overlying water of Method B washed sediment (vortex-and-centrifuge) was statistically similar to the amount of total Zn for unwashed sediment, and the concentration of total Cu was significantly higher in the overlying water of Method B sediment compared to unwashed

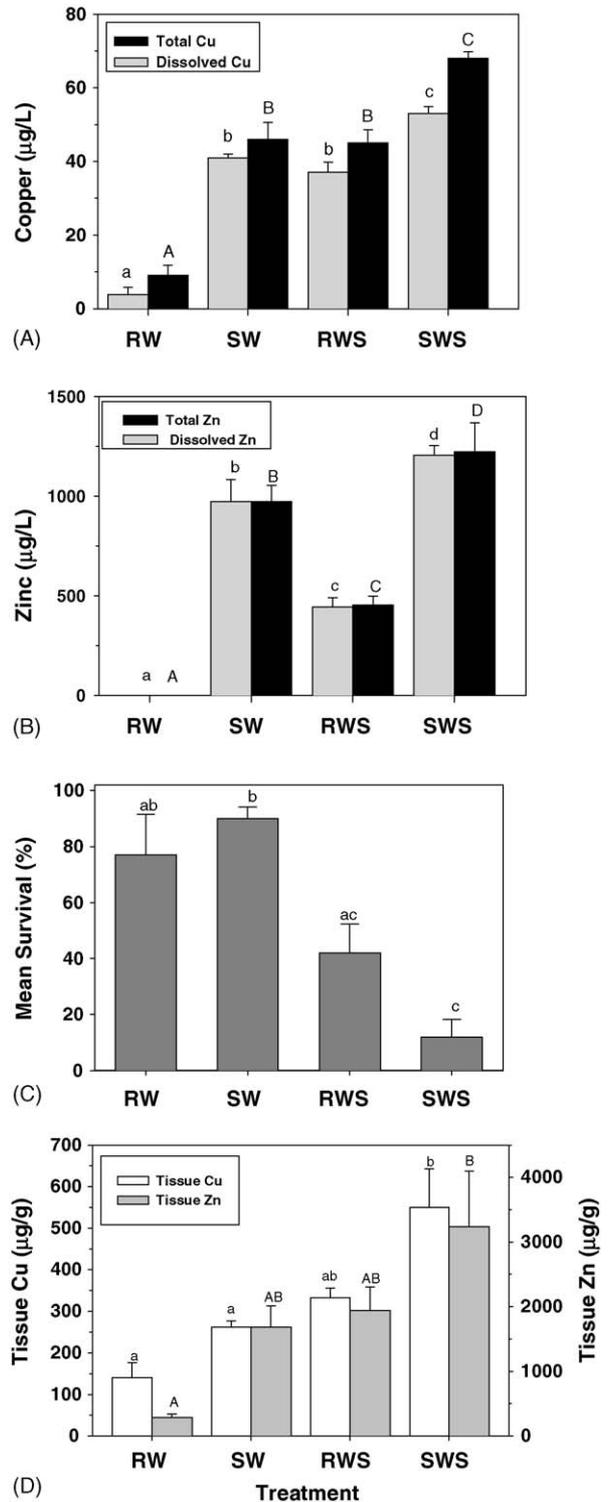


Fig. 2. Mean (\pm S.E., $n=4$), concentrations of total and dissolved (A) Cu, and (B) Zn in the overlying water, (C) survival, and (D) whole-body tissue concentrations of Cu and Zn in *D. magna* exposed to unwashed sediments. RW = reference water, SW = site water, RWS = reference water + sediment, and SWS = site water + sediment. Note Zn was below detection limit in the reference water. Bars labelled with different letters are significantly ($p < 0.05$) different from each other. In panels (A) and (B) statistical comparisons using lower case letters are for dissolved metal and upper case letters are for total metals. In panel (D), statistical comparisons using lower case letters are for Cu (open bars) and upper case letters (shaded bars) are for Zn.

Table 2

Mean (\pm S.E., $n = 4$) total and dissolved concentrations ($\mu\text{g/L}$) of Zn and Cu in the overlying water at the beginning and the end of the initial exposure with unwashed sediment

Metal	Treatment	T = 0 h			T = 48 h		
		Total metal	Dissolved	% dissolved	Total metal	Dissolved	% dissolved
Zn	Reference water (control)	ND	ND	–	ND	ND	–
	Site water only	759 (76.5)	643 (69.5)	84.7	973 (81)	972 (115.5)	99.9
	Sediment + site water	1300 (14.5)	1143 (22.5)	87.9	1222 (73)	1205 (48)	98.6
	Sediment + reference water	482 (53.5)	423 (50.0)	87.8	454 (45)	443 (47)	97.6
Cu	Reference water (control)	13.9 (5.2)	6.0 (2.6)	43.2	13.4 (6.0)	12.8 (5.7)	95.5
	Site water only	50.2 (5.5)	29.9 (1.0)	59.6	45.8 (4.5)	41.4 (1.0)	90.4
	Sediment + site water	78.0 (3.9)	49.5 (0.8)	63.5	67.9 (1.8)	53.0 (1.9)	78.1
	Sediment + reference water	45.7 (4.0)	28.4 (1.3)	62.1	45.4 (3.6)	37.3 (2.8)	82.2

Note: ND = not detected ($<50 \mu\text{g/L}$).

sediment (Fig. 4A and B). Survival of *D. magna* exposed to washed sediment was significantly higher ($>95\%$) compared to those exposed to unwashed sediment ($<50\%$) (Fig. 4C). Washing by both methods reduced tissue Zn concentrations to a similar extent but tissue Cu concentrations were significantly ($p = 0.002$) higher in *D. magna* exposed to sediments washed with Method B compared to sediments washed with Method A (Fig. 4D).

3.4. Separation of phases

Dissolved Cu was $2 \mu\text{g/L}$ in the reference water control and $24\text{--}27 \mu\text{g/L}$ in the exposures. Particle-associated ($>0.45 \mu\text{m}$) Cu was below the detection limit in the reference water and the filtered treatments but was $30 \mu\text{g/L}$ in the overlying water of unfiltered treatments (unfiltered and unfiltered with sediment). Dissolved Zn concentrations in all treatments and the particle-associated Zn in the reference water control were all near the detection limit. Particle-associated Zn in the overlying water of the unfiltered treatments ranged from 150 to $480 \mu\text{g/L}$. The concentrations of Cu and Zn in the exposure sediment were 398 and $1825 \mu\text{g/g}$, respectively.

Survival of *D. magna* exposed to filtered overlying water ($74\% \pm 10.7$) was not significantly different ($p > 0.05$) than survival in the treatments with suspended particles (unfiltered, $54\% \pm 8.7$; unfiltered + sediment $44\% \pm 6.8$) (Fig. 5A). Similarly, tissue concentrations of Cu and Zn were statistically similar across all treatments (Fig. 5B).

Linear regression analysis revealed a significant relationship ($p = 0.0003$, $r^2 = 0.61$) between the concentration of dissolved Cu in the overlying water and the concentration of Cu in the tissue of *D. magna*. The relationship between particle-associated Cu and tissue Cu was also significant ($p = 0.04$, $r^2 = 0.27$) but explained far less of the variation in the data. No significant relationships were found between the concentration of Zn (dissolved or particulate) in the overlying water and the concentration of Zn in *D. magna* tissues. Similarly, stepwise linear regression using the concentrations of dissolved Cu and Zn, and the concentrations of particulate Cu and Zn, revealed that, only the concentration of dissolved Cu significantly contributed to *D. magna* mortality.

3.5. Combined dataset

In order to obtain metal concentrations that spanned a wide range of exposure concentrations we combined all the data from the unwashed and washed sediment exposures along with the data from the phase separation experiment into a composite dataset for the study. Regression analysis of the combined dataset showed a significant relationship ($r^2 = 0.76$, $p < 0.0001$) between the concentration of dissolved Cu and the survival of *D. magna* (Fig. 6A) but not between the concentration of particle-associated Cu and survival (Fig. 6B). There were no significant relationships between dissolved or particulate Zn and survival. Because of incomplete gut clearance in the first two experiments,

Table 3

Mean (\pm S.E.) total and dissolved concentrations ($\mu\text{g/L}$) of Zn and Cu in the overlying water at the beginning and end of the 48 h exposure with washed sediment

Metal	Treatment	T = 0 h			T = 48 h		
		Total metal	Dissolved	% dissolved	Total metal	Dissolved	% dissolved
Zn	Reference water	ND	ND	–	ND	ND	–
	Method A	159 (20)	63 (12.5)	39	69 (23)	ND	–
	Method B	345 (17.0)	80 (1.4)	23	175 (11.3)	ND	–
Cu	Reference water	32.8 (12.8)	4.0 (1.6)	12	4.4 (0.5)	3.9 (0.6)	89
	Method A	31.8 (2.1)	9.5 (1.0)	30	18.3 (3.5)	12.0 (1.6)	66
	Method B	91.8 (5.5)	13.7 (0.4)	15	57.2 (1.1)	15.8 (0.4)	28

Reference water was used a control. Method A is 'stir-and-settle'. Method B is 'vortex-and-centrifuge'. Note: ND = not detected ($<50 \mu\text{g/L}$).

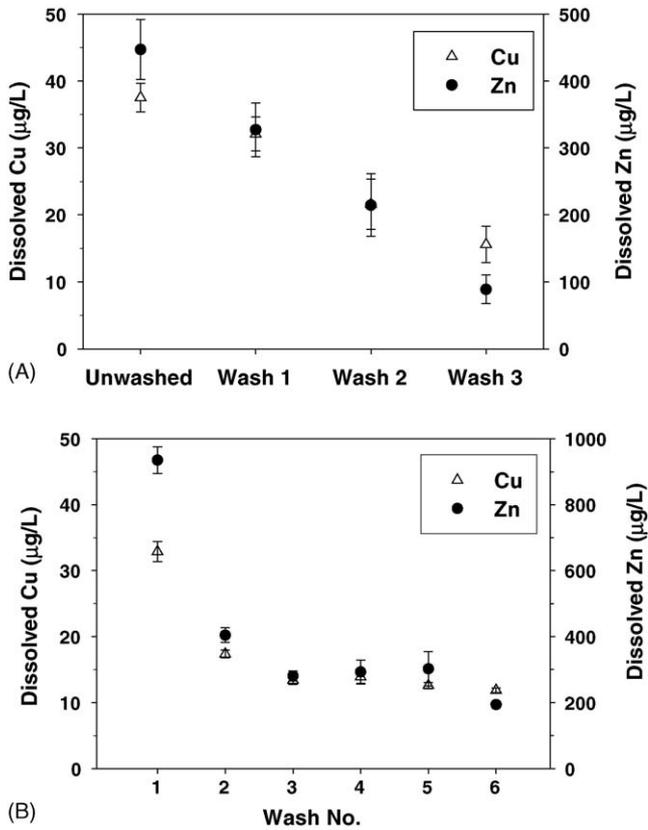


Fig. 3. Mean (\pm S.E.) dissolved Cu and Zn concentrations in the overlying water of sediments washed using (A) Method A (stir-and-settle) $n=4$, and (B) Method B (vortex-and-centrifuge) $n=8$.

tissue metal concentrations from those exposures could not be combined with the tissue metal data from the phase separation experiment for similar analysis.

4. Discussion

4.1. Sediment toxicity

The low survival (<50%) of *D. magna* exposed to unwashed sediment demonstrated that there was substantial bioavailable metal associated with the Clear Creek sediment. The concentrations of Cu and Zn in the overlying water of unwashed sediment fall within the range of published EC50s for neonate (<24 h old) *D. magna* (De Schampelaere et al., 2002; De Schampelaere and Janssen, 2002; Muysen and Janssen, 2001). Although 5-day-old *D. magna* used in this study may be less sensitive to metal exposure than neonates (Stuhlbacher et al., 1993) some toxicity would be expected at these levels of dissolved Cu and Zn. Therefore, in order to reduce dissolved Cu and Zn below the toxic range, it was necessary to remove as much as possible of the easily mobilized metals from the sediment. Our preliminary analyses (see Section 2) demonstrated that Cd and Pb were not of concern in these exposures. The significant decrease in water-borne Cu and Zn, along with the corresponding increase in survival in washed sediment exposures (compared to unwashed) confirmed that Zn and/or Cu in the overlying water

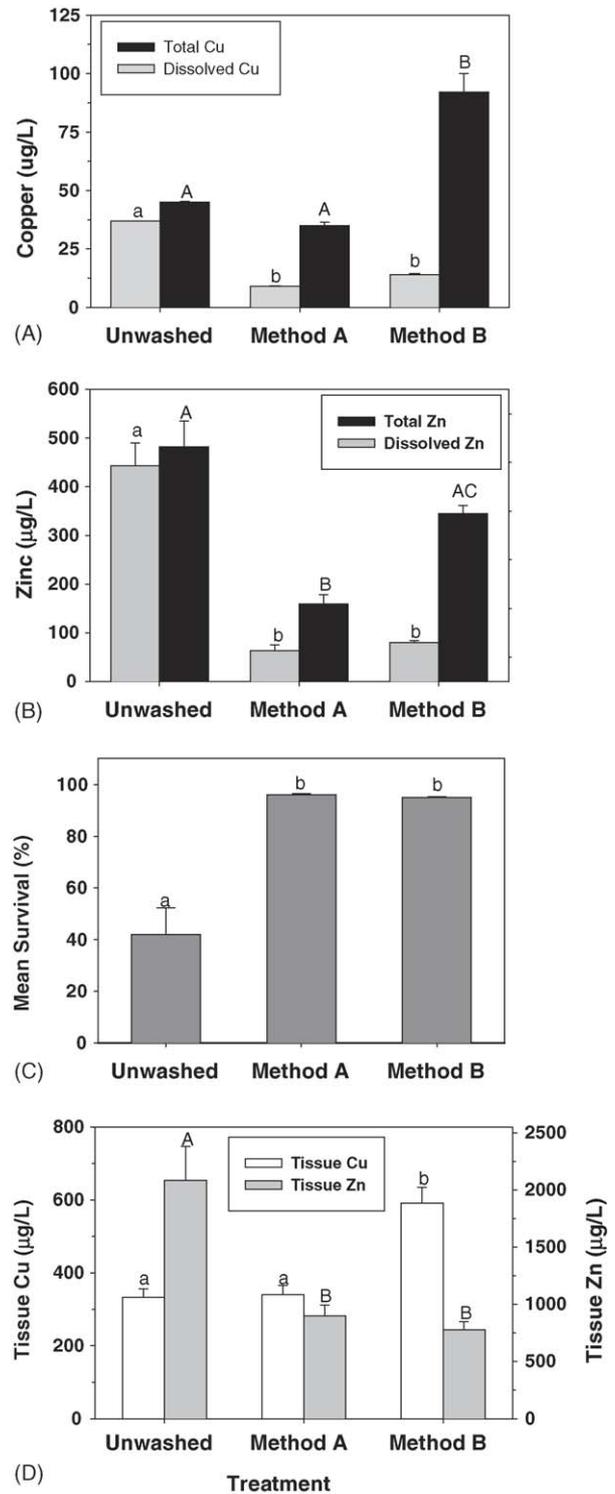


Fig. 4. Mean (\pm S.E., $n=4$), concentrations of total and dissolved (A) Cu, and (B) Zn in the overlying water, (C) survival, and (D) whole-body tissue concentrations of Cu and Zn in *D. magna* exposed to washed (Methods A and B) and unwashed sediments. Note Zn was below detection limit in the reference water. Bars labelled with different letters are significantly ($p < 0.05$) different from each other. In panels (A) and (B) statistical comparisons using lower case letters are for dissolved metal and upper case letters are for total metals. In panel (D), statistical comparisons using lower case letters are for Cu (open bars) and upper case letters (shaded bars) are for Zn.

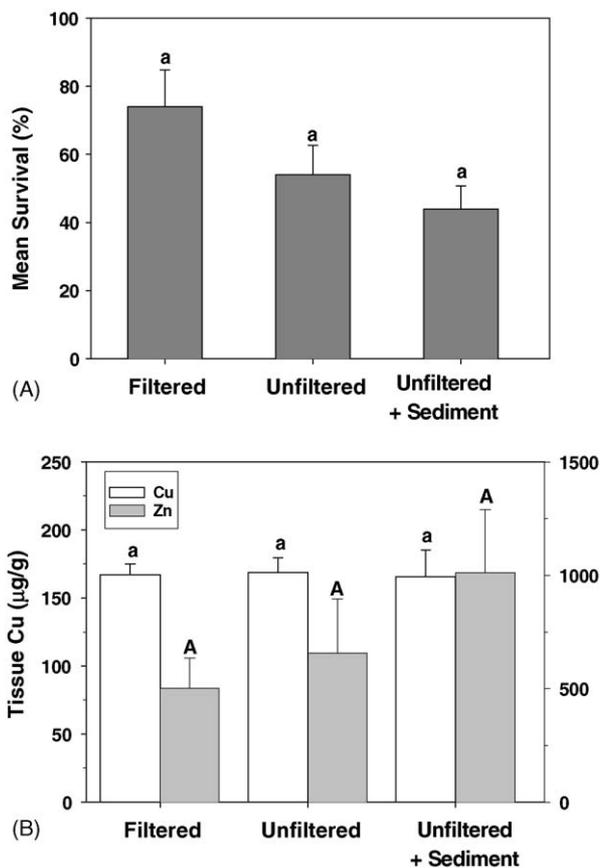


Fig. 5. Mean (\pm S.E., $n = 5$) (A) survival and (B) whole-body tissue concentrations of Cu and Zn of *D. magna* exposed to filtered and unfiltered overlying water, and unfiltered overlying water with sediment. In panel (B), statistical comparisons using lower case letters are for Cu, and upper case letters are for Zn.

of unwashed sediments were likely responsible for *D. magna* mortality.

4.2. Sediment washing

Repeated rinsing of the sediment was successful at removing most of the labile metals from the sediment. Both methods of washing were adequate for this task but the quicker, vortex-and-centrifuge, Method B, was favored in the interest of time. It should be noted that the level of suspended particle-associated metal in the overlying water was enhanced when sediments were washed this way (Fig. 4A and B). We suspect that ‘vortex’ mixing may increase the bioavailability of particle-bound metals to *Daphnia*. Tissue Cu concentrations in *D. magna* exposed to sediments washed with Method B were significantly higher than those in animals exposed to sediment washed by the stir-and-settle Method A or to unwashed sediment (Fig. 4D). After undertaking a more detailed investigation into the gut clearing patterns of *D. magna* (Gillis et al., 2005), we strongly suspect that at least some of this ‘tissue Cu’ reported for the unwashed (initial) and washed sediment exposures after only 1 h of purging in reference water alone was due to Cu-bearing particles that remained in the gut because of incomplete gut clearance. Based on the calculations of Gillis et al. (2005) we estimate that

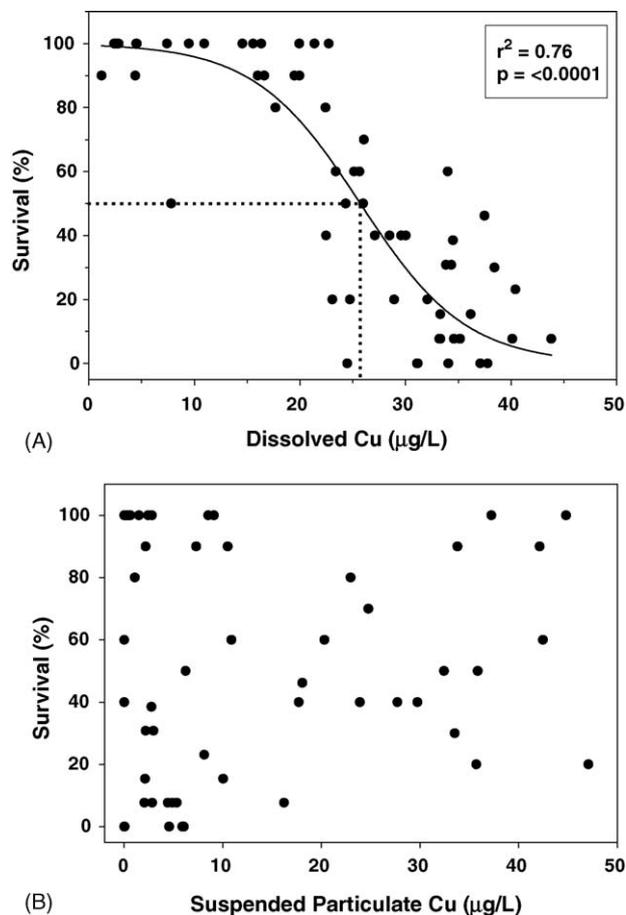


Fig. 6. Relationship between (A) dissolved and (B) suspended particulate Cu and *D. magna* survival. Data were compiled from the unwashed, washed, and phase separation exposures. Dotted line indicates the observed (26 µg/L) LC50 for dissolved Cu in these exposures.

over 60% of the whole-body Cu concentration (400 µg/g) for the unwashed and washed sediment exposures could be attributed to Cu-laden particles in the gut. Therefore, we suggest that the significant increase in tissue Cu in *D. magna* exposed to sediment washed using Method B (Fig. 4D), was due to an increase in sediment-associated Cu in the gut. The vigorous action of ‘vortex’ mixing likely broke up the larger aggregates into smaller particles. Smaller particles would not only have stayed in suspension longer (as was reflected in the significant increase in total Cu in the overlying water of Method B washed sediment) but they may also now be within the size range that *D. magna* can retain on their filtering appendages, thereby contributing to the increase in tissue Cu. For Zn, the concentration of total Zn in the overlying water of sediment washed by Method B was only slightly (not significantly) reduced compared to levels in unwashed sediment, with tissue Zn concentration in washed sediment significantly lower than in unwashed sediment exposures. The reason for these differences are unknown but may be related to differences in the association between the metals and the sediment particles. Nevertheless the lack of a corresponding negative effect on the survival of *D. magna* exposed to ‘vortex-washed’ sediment further supports the conclusions that this ‘tissue Cu’ had not actually bioaccumulated.

4.3. Separation of phases

Once the amount of metal dissociating from the Clear Creek sediment had been reduced through sediment washing and dilution with clean sediment, we could begin to investigate whether the particle-associated metals were bioavailable to *D. magna*. By removing the suspended particles (>0.45 µm) through filtration, the amount of metal accumulated from the dissolved and the particulate phase could be quantified. Although there was no significant difference in survival between the treatments, there was a trend of decreasing survival from the filtered, to unfiltered, to the unfiltered + sediment exposure (Fig. 5A). Because the concentration of dissolved copper was similar across all treatments (24–27 µg/L) we suggest that this trend is not the result of differential copper exposure across the treatments. Since *D. magna* exposed to filtered and unfiltered overlying water, even in the presence of sediment, accumulated similar amounts of Cu and Zn (Fig. 5B), we concluded that only the remaining labile metal was bioavailable and that any Cu or Zn that remained associated with the sediment after washing (i.e. the particle-associated metal) was not available to *Daphnia*. The significant relationship between dissolved Cu and survival (Fig. 6A) and the lack of a significant relationship between the particle-associated Cu and survival in the combined dataset (Fig. 6B) supports an earlier study by Erickson et al. (1995) who reported that Cu associated with sediment particles was not bioavailable to *C. dubia*. However, our findings contrast with Weltens et al. (2000), who found that particle-associated Cd was bioavailable and thus toxic to *C. dubia*. The difference in toxicity between the Weltens et al. (2000) study and this study could be attributed to differences in desorption behavior of sediment-associated Cd versus Cu.

4.4. Combined dataset

Using our combined dataset (all exposures in this study) we observed 50% mortality (LC50) when the dissolved Cu concentration reached 26 µg/L (Fig. 6A). The Biotic Ligand Model (BLM) (HydroQual, 2005) was used to predict copper toxicity for *Daphnia* over the pH range observed in our exposures (6.44–7.42). According to the BLM, the predicted LC50 for Cu for *D. magna* would be between 9 and 35 µg/L. This appears to be a reasonable prediction, considering that the sediment used in these exposures was mining-impacted, field-collected sediment which is contaminated by numerous metals, and that BLM predictions are based upon single metal (Cu) exposures.

4.5. Environmental relevance

There has been considerable interest in the fate of metal-contaminated sediment during natural resuspension events and dredging. Bonnet et al. (2000) simulated a resuspension event by stirring a number of moderately contaminated sediments (Cu 40–70 µg/g, Zn 140–200 µg/g) and found that 24 h after mixing that only a small fraction of the sediment-associated metals were released into the overlying water and that no toxicity to *Hydra attenuata* and *D. magna* was observed. By contrast in the present study, there was a higher potential for desorption of

metals into the overlying water due to the much higher metal load in the Clear Creek sediments (unwashed: Cu 2424 µg/g, Zn 5150 µg/g; Table 1). Carvalho et al. (1998) also investigated the effect of sediment resuspension and thus oxidation of metal-contaminated anoxic sediment, and reported that Cu and Zn were mobilized from the sediments resulting in concentrations well in excess of acutely toxic levels, which was reflected in the toxicity to *D. similis*. The watershed of Clear Creek has been identified as an area of high priority because of metal pollution from historic mining activities in the area. Recently, USEPA (2005) concluded that dissolved metals in the surface waters of Clear Creek were not likely to have a significant acute or chronic impact to aquatic biota but that sediment leachate tests and sediment toxicity tests both indicated that portions of Clear Creek are toxic for survival and growth. Based on the results of our laboratory study, we would expect that if the sediment of Clear Creek was subjected to a resuspension event, there could be a large efflux of metals from the sediment into the water column resulting in an acutely toxic environment for sensitive organisms such as *D. magna*.

5. Conclusions

Exposure to field-collected mining-impacted sediment was acutely toxic to *D. magna*, at least in part due to high levels of Cu in the dissolved phase of the overlying water. Once the labile metals had been exhausted, the Cu and Zn that remained associated with the particles were not bioavailable *D. magna*.

Acknowledgements

This study was funded by the Center for the study of Metal in the Environment (USEPA). CMW is supported by the Canada Research Chair Program. We would like to thank Sonia Sharma and Andrée McCracken for assistance in the laboratory and Dr. Chris Glover for input into this study.

References

- Adam, C., Garnier-Laplace, J., Baudin, J.P., 2002. Bioaccumulation of ^{110m}Ag, ⁶⁰Co, ¹³⁷Cs and ⁵⁴Mn by the freshwater crustacean *Daphnia magna* from dietary sources (*Scenedesmus obliquus* and *Cyclotella meneghiniana*). Water Air Soil Pollut. 136, 125–146.
- Arambašić, M.B., Bjelić, S., Subakov, G., 1995. Acute toxicity of heavy metals (copper, lead, zinc), phenol and sodium on *Allium cepa* L., *Lepidium sativum* L. and *Daphnia magna* St.: comparative investigations and the practical applications. Water Res. 29, 497–503.
- Barata, C., Baird, D.J., Markich, S.J., 1998. Influence of genetic and environmental factors on the tolerance of *Daphnia magna* Straus to essential and non-essential metals. Aquat. Toxicol. 42, 115–137.
- Barata, C., Markich, S.J., Baird, D.J., Soares, A.M.V.M., 2002. The relative importance of food as cadmium sources to *Daphnia magna* Straus. Aquat. Toxicol. 61, 143–154.
- Biesinger, K.E., Christensen, G.M., 1972. Effects of various metals on survival, growth, reproduction and metabolism of *Daphnia magna*. J. Fish. Res. Bd. Canada 29, 1691–1700.
- Bonnet, C., Babut, M., Ferard, J.F., Martel, L., Garric, J., 2000. Assessing the potential toxicity of resuspended sediment. Environ. Toxicol. Chem. 19, 1290–1296.
- Brendelberger, H., 1985. Filter mesh-size and retention efficiency for small particles: comparative studies with Cladocera. Arch. Hydrobiol. Beih. Ergebn. Limnol. 21, 135–146.

- Carvalho, P.S.M., Zanardi, E., Burantini, S.V., Lamparelli, M.C., Martins, M.C., 1998. Oxidizing effect on metal remobilization and *Daphnia similis* toxicity from a Brazilian reservoir sediment suspension. *Water Res.* 32, 193–199.
- Chapman, G.A., 1980. Effects of Water Hardness on the Toxicity of Metals to *Daphnia magna*. USEPA Environmental Research Laboratory, Corvallis, OR (USEPA, 1980. Ambient water quality criteria for lead. EPA 440/5-80-057, Washington, DC).
- De Schamphelaere, K.A.C., Janssen, C.R., 2002. A biotic ligand model predicting acute copper toxicity to *Daphnia magna*: the effects of calcium, magnesium, sodium, potassium and pH. *Environ. Sci. Technol.* 36, 48–54.
- De Schamphelaere, K.A.C., Heijerick, D.G., Janssen, C.R., 2002. Refinement and field validation of a biotic ligand model predicting acute copper toxicity to *Daphnia magna*. *Comp. Biochem. Physiol. C* 133, 243–258.
- Erickson, R.J., Kleiner, C., Fiant, J., Highland, T., 1995. Effects of suspended solids on metals toxicity to aquatic organisms. Internal Report. Mid-continent Ecology Division, U.S. Environmental Protection Agency, Duluth, MN, USA.
- Fargašová, A., 1994. Toxicity of metals on *Daphnia magna* and *Tubifex tubifex*. *Ecotoxicol. Environ. Saf.* 27, 210–213.
- Fisher, N.S., Hooke, S.E., 2002. Toxicology tests with aquatic animals need to consider the trophic transfer of metals. *Toxicology* 181/182, 531–536.
- Gillis, P.L., Chow-Fraser, P., Ranville, J.F., Ross, P.E., Wood, C.M., 2005. *Daphnia* need to be gut cleared too: the effect of exposure to and ingestion of metal-contaminated sediment on whole-body metal concentrations and the gut clearance patterns in *D. magna*. *Aquat. Toxicol.* 71, 143–154.
- Harvey, B.B., Ranville, J.F., Ross, P.E., 2003. Seasonal changes in the characteristics of suspended sediment metal transport in a mining-impacted stream. In: Proceedings of the 20th National Conference of the American Society of Mine Reclamation/Ninth Billings Land Reclamation Symposium, June 3–6, pp. 353–366.
- Hooke, S.E., Fisher, N.S., 2001. Sublethal effects of silver exposure in zooplankton: importance of exposure pathways and implications for toxicity testing. *Environ. Toxicol. Chem.* 20, 568–574.
- Horton, P.A., Rowan, M., Webster, K.E., Peters, R.H., 1979. Browsing and grazing by cladoceran filter feeders. *Can. J. Zool.* 57, 206–212.
- Lampert, W., 1987. Feeding and Nutrition in *Daphnia*. In: Peters, R., de Bernadi (Eds.), *Memorie dell'Institutito Italiano di Idrobiologia Dott. Marco de Marchi*, vol. 45, 'Daphnia' R.H. pp. 143–192.
- Ma, H., Kim, D.K., Allen, H.E., Cha, D.K., 2002. Effect of copper binding by suspended particulate matter on toxicity. *Environ. Toxicol. Chem.* 21, 710–714.
- Martinez-Madrid, M., Rodriguez, P., Perez-Iglesias, J.I., 1999. Sediment toxicity bioassays for assessment of contaminated sites in the Nervion River (Northern Spain). 1. Three-brood sediment chronic assay of *Daphnia magna* Straus. *Ecotoxicology* 8, 97–109.
- Muysen, B.T.A., Janssen, C.R., 2001. Multigenerational Zn acclimation and tolerance in *Daphnia magna*: implications for water-quality guidelines. *Environ. Toxicol. Chem.* 20, 2053–2060.
- Persaud, D., Jaagumagi, R., Hayton, A., 1992. Guidelines for the Protection and Management of Aquatic Sediment Quality in Ontario. Water Resources Branch, Ontario Ministry of the Environment, pp. 1–23.
- Stuhlbacher, A., Bradley, M.C., Naylor, C., Calow, P., 1993. Variation in the development of cadmium resistance in *Daphnia magna* Straus: effect of temperature, nutrition, age and genotype. *Environ. Pollut.* 80, 153–158.
- Taylor, G., Baird, D.J., Soares, A.M.V.M., 1998. Surface binding of contaminants by algae: consequences for lethal toxicity and feeding to *Daphnia magna* Straus. *Environ. Toxicol. Chem.* 17, 412–419.
- USEPA, 1993. Methods for Measuring the Acute Toxicity of Effluents and Receiving Water to Freshwater and Marine Organisms, 4th ed. USEPA (Appendix A, 16.60049002F).
- USEPA, 2005. Ecotoxicological and water quality evaluation of Clear Creek and the North Fork Clear Creek, Colorado, Clear Creek/Central City Superfund Site Operable unit 4. USEPA Region 8. USEPA, Denver, CO.
- Wang, W.-X., Fisher, N.S., 1998. Accumulation of trace elements in a marine copepod. *Limnol. Oceanogr.* 43, 273–283.
- Weltens, R., Goossens, R., Van Puymbroeck, S., 2000. Ecotoxicology of contaminated suspended solids for filter feeders (*D. magna*). *Arch. Environ. Contam. Toxicol.* 39, 315–323.
- Weltens, et al., Weltens, R., Witters, H., Corbisier, P., 2001. Ecotoxic impact of suspended solids collected from polluted surface waters. *J. Soil Sedim.* 1, 223–233.
- Yu, R.-Q., Wang, W.-X., 2002a. Trace metal assimilation and release budget in *Daphnia magna*. *Limnol. Oceanogr.* 47, 495–504.
- Yu, R.-Q., Wang, W.-X., 2002b. Kinetic uptake of bioavailable cadmium, selenium, and zinc by *Daphnia magna*. *Environ. Toxicol. Chem.* 21, 2348–2355.