A GENERALIZED, STEADY-STATE SIMULATION

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OF

WASTEWATER TREATMENT PLANTS

A GENERALIZED, STEADY-STATE SIMULATION

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WASTEWATER TREATMENT PLANTS

by

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ii

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SCOPE AND CONTENTS:

Laboratory and field measurements were combined with theory in developing models for the various wastewater treatment processes. Some assumptions were made, both in an attempt to simplify the complex mechanisms encountered and to explain known experiences and observations.

Models for a primary settling tank, a secondary settling tank, a generalized aerobic biological sludge treatment scheme, an anaerobic digestion system and several other unit processes have been developed. The generalized aerobic biological sludge treatment scheme is an attempt to describe in one physical package, all the various activated sludge process modifications.

The executive system used is GEMCS, together with its library of subroutines, such as CONTLl, SEPAOl, JUNCOl and

iii

and SETSPI, some of which have been modified slightly.

The models have been tested on the Drury Lane and the Skyway, Burlington, Sewage Treatment Plants, and have been found to agree very well with plant data. The former has a conventional activated sludge process, whilst the latter employs the extended aeration modification.

An inter-active version has been developed, which requires the minimum knowledge of GEMCS for its use. This can be applied as a training and teaching tool for plant operators and students in the field.

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(v)

TABLE OF CONTENTS

PAGE NUMBER

 $\ddot{}$

(vii)

PAGE NUMBER

LIST OF FIGURES

 $\overline{}$

(x)

PAGE NUMBER

 $\ddot{}$

FIGURE NUMBER

 $\ddot{}$

 $\ddot{}$

 \overline{a}

PAGE NUMBER

FIGURE NUMBER

LIST OF TABLES

k,

(xiii)

PAGE NUMBER

TABLE NUMBER

CHAPTER 1

1. INTRODUCTION

Although waste treatment practices have been known ever since the turn of the century, it was not until about the late 1940's that an effort was made to provide firm theoretical bases for design. Significant advances have been made in recent years, resulting in a better understanding of the complex variables affecting the operation of the various unit processes. It is now possible, where it would be beneficial, to bring together the models of the various waste treatment processes, to simulate the operation of full-scale plants, on a computer.

A successful simulation can have great potential applications, a few of which are:

- (1) The performance of the individual units and of the plant as a whole can be predicted over a wide range of feed flows and operating conditions,
- (2) The computer simulation can help in the selection of an optimal design for a plant to treat a given waste to a satisfactory level, at the least cost,
- {3) It can be.used effectively in the training of plant operators and as a valuable educational tool for students in the field, and
- (4) It can be used for on-line process control purposes.

Several computer simulations have been attempted, both in the United States and in Canada, examples of which include those by Smith (46, 47), Silveston (40, 41), and by Fan and Erickson (10, 11). However, there is a severe limitation in their models for the activated sludge system. They have based their kinetics either on the entire sludge mass or on the volatile suspended matter in the sludge mass, when it is known that microorganisms are the only agents capable of stabilizing the waste. It is for this reason that their models break down when they are applied to the various modification of the conventional activated sludge process, as the composition of the sludge then changes.

One of the major objectives of this work is to develop a means by which the microbial mass may be estimated. The kinetic expressions can then be formulated around the physiological activities of the-microorganisms, leading to a more meaningful model for the activated sludge process. This generalized model can then be applied to the various modifications of the activated sludge process.

As a great variety of raw wastes are encountered in wastewater treatment practice, it would be useful to develop generalized models that cap handle all of them. However, the parameters pertaining to each type of waste will be different. Consequently, another objective of this work is to compile a series of experiments that can be used to

characterize a given wastewater and its treatability parameters. Although the work in this thesis is based primarily on domestic sewage, the same experiments used here may be performed on other wastewaters.

It is also realized that the complexity of a simulation system is discouraging its use by most plant operators or engineers. Furthermore, there is a tendency for interested parties to create their own simulation system rather than to try to understand and use other systems. It is for this reasons that we feel it necessary to develop an interactive version of the computer simulation which would require the minimum knowledge of the simulation system, for its use.

CHAPTER 2

2. LITERATURE REVIEW

The model for the primary settler proposed by R. Smith (46), was derived by plotting the data for suspended solids removal versus overflow rate for full-sized plants, as reported in "Sewage Treatment Plant Design", (38). He obtained the relationship

FRPS = $0.82 \exp(-GPS/2780)$ (2.1)

where FRPS = fraction of solids removed

GPS = overflow rate usqpd/ft.²

The fractional removal was applied to all classes of suspended solids, which may not be a valid assumption.

P. Silveston (41), tested the model on two Ontario plants and found that the effluent suspended solids predicted were about 25% higher than that actually observed. This led him to suggest that actual plant data have to be used to achieve a reasonable simulation. He proposed the relationship

FRPS = $1 - \exp(-k/GPS)$ (2.2)

where $k = constant$ fitted from plant data.

Both models, however, do not account for the effect of turbulence on the settling process. The constant k in Silveston's model could be interpreted to include both the effects of turbulence and the settling characteristics of the feed solids.

Camp (4) has developed a mechanistic model for discrete particle sedimentation in a continuous-flow tank. He first introduced the concept that a real settling tank may be divided into an inlet zone, a settling zone and an outlet zone. , The actual sedimentation occurs in the middle zone, whilst turbulence at both the inlet and outlet zones prevent any further sedimentation from occuring. He also introduced the use of batch settling curves to predict the performance of a continuous-flow sedimentation tank.

Most of the models for the activated sludge process have evolved from the complete mixing model first presented by McKinney (25). The original paper was a little vague, and in a later discussion, Washington (47) corrected and clarified several points. However, his model can be interpreted as follows, noting that all the various forms of organic carbon are in their oxygen equivalents:

In the presence of unlimited food the rate of synthesis of the active solids is limited only by the concentration of the active solids, i.e.,

$$
\frac{d(MLVSS)}{dt} = k_{S}^{\prime (MLVSS)}
$$
 (2.3)

But in the case of food-limiting situation, the rate becomes

$$
\frac{d(MLVSS)}{dt} = k_{S}C
$$
 (2.4)

where $MLVSS$ = oxygen equivalent of active solids $c =$ oxygen equivalent of substrate $k_{\rm e}$, $k_{\rm e}$ = rate constants.

He assumed that the volatile solids in the sludge is entirely active. He also proposed that the rate of substrate removal is proportional to the rate of synthesis of active solids, i.e ,

$$
\frac{dC}{dt} = -(1 + k_a) \frac{d(MLVSS)}{dt}
$$
 (2.5)

The basal metabolism rate was also assumed to be proportional to the concentration of active solids, i.e.,

$$
\frac{d(MLVSS)}{dt} = -k_d(MLVSS)
$$
 (2.6)

By performing mass balances around the stirred tank reactor, he obtained, assuming a food-limiting situation:

$$
C_0 = \frac{C_i}{1 + (1 + k_a) k_s \bar{t}}
$$
 (2.7)

and $M_o = {K_S C_O \over S_O}$ (2.8)

$$
\vec{c}_{\text{in}} = \text{influent substrate concentration}
$$
\n
$$
c_{\text{o}} = \text{effuent substrate concentration}
$$
\n
$$
M_{\text{o}} = \text{effuent active solids concentration}
$$
\n
$$
\vec{t} = \text{detection time}
$$
\n
$$
x = \text{fraction of sludge mass wasted.}
$$

 k_{d} + \underline{x}

Estimated values of his constants were given as: $k_a = 0.5$ $k_d = 0.006$ hr.⁻¹ k_e = 15 hr.⁻¹ (minimum) and k_{s} = 7 hr.⁻¹ (minimum)

R. Smith (46) in his model, assumed that the rate of substrate removal is proportional to both the concentration of active solids and the substrate concentration, i.e.,

$$
\frac{dC}{dt} = - k_{S}^{\text{''}} C \text{ (MLSS)}
$$
 (2.9)

as contrasted to equations (2.3) and (2.4). k_{S} " is a rate constant given by:

$$
k_{S}^{\text{''}} = 0.0001 (1.047)^{T-20} (hr)^{-1} (mg/l)^{-1}
$$
 (2.10)

Erickson, Ho and Fan (11) assumed Michaelis-Menten kinetics with the expression

$$
U = \frac{1}{\text{MLSS}} \frac{dC}{dt} = -\frac{U_{\text{max}} C}{Y (C^* + C)}
$$
\nwhere $U_{\text{max}} = \text{maximum unit growth rate}$

\n
$$
C^* = \text{value of C where } U = \frac{1}{2} U_{\text{max}}
$$
\n
$$
Y = \text{yield factor.}
$$
\n(2.11)

It may be noted that equation (2.11) reverts to the form of equation (2.3) when $C \gg C^*$, and to the form of equation (2.9) when $C \ll C^*$.

Little quantitative information exists on the operation of the secondary settler. The models proposed by Ott and Bogan (29), Erickson and Fan (10, 11), and Silveston (41), have assumed that a constant fraction β , of the input solids escapes over the wier and that the underflow solids are compacted by a ratio c. R. Smith (46) proposed a model, based on the data of Villiers (51) where

$$
\beta = \frac{556 \text{ (GPS)}^{0.494}}{\text{MLSS}^{1.82} \text{ (TA)}^{0.439}}
$$
 (2.12)

GPS is the settler overflow rate (USqpd/ft.²), MLSS is the inlet mixed liquor suspended solids (mg/1) , and TA the aeration time (hours). For some reason he later reverted back to the simple splitter model.

Anaerobic digestion is a complex operation involving several reaction stages and different microorganisms. An accurate description for the process is difficult to obtain. McCarty (24) using Michaelis-Menten kinetics and a completely stirred tank model, arrived at the relationship:

$$
\frac{\kappa_{\max} \ \mathbf{F}}{\mathbf{F}^* \ + \ \mathbf{F}} = \frac{1}{\mathbf{E}}
$$
 (2.13)

where k_{max} maximum unit growth rate constant F^* = value of F at which unit growth rate is $\frac{1}{2}$ k_{max}

 $F =$ concentration of biodegradable carbon in effluent

and \bar{t} = digester residence time.

The reported values of k_{max} and F^* are:

 k_{max} = 0.28 exp $[- 0.036 (35 - T)]$ (2.14) and F^* = 700 exp $[0.10 (35 - T)]$ (2.15) where $T =$ temperature of digestion, ^oc.

His relationship implies that the effluent degradable carbon is independent of the influent value, but depends only on the detention time. He also suggested an expression to estimate the organic carbon of the volatile acids (VAC) in the effluent as:

$$
VAC = \frac{200 \exp [0.12 (35 - T)]}{1 + k_{max} \bar{t}}
$$
 (2.16)

Extensive cost information is available in the literature. R. Smith (45, 46} presented a thorough investigation of the capital and operating costs of domestic waste treatment unit processes. The capital and operating costs of a unit are calculated as a function of its most significant capacity variable. Michel et al. (26) reported on the total operating and maintenance costs of a treatment plant as a function of the waste flow and also of the population equivalent. Barnard and Eckenfelder (2) included industrial waste treatment costs in their report. A more complete list of references may be obtained from the above reports.

Several papers have been published on the optimization studies of wastewater treatment processes. Erickson and Fan (10) studied the optimization of the hydraulic regime of activated sludge systems. Erickson, Ho and Fan (11) later investigated the modelling and optimization of the step aeration processes, which consists of a series of completely mixed tanks. Shih and Krishnan (39) reported on their studies on the dynamic optimization of industrial waste treatment plant designs. Realizing that many design parameters are not accurate, Berthouex and Polkowski (3) studied the effect of such uncertainties on the optimal design of a treatment plant.

Further references from the literature will be made, as the thesis is developed.

CHAPTER 3

3. THE SIMULATION SYSTEM

3.1 The GEMCS Executive

Any processing or manufacturing plant can be broken down into its component unit processes. The unit process is a basic operation within the framework of the plant, and can usually be identified with a distinct physical piece of equipment, examples of which are primary sedimentation, chlorination, drying, etc. The mathematical model of a unit process is known as a unit computation.

In a simulation system, it is necessary to have a . master-mind to execute each unit computation in its logical sequence. Such is the purpose of the executive program. It should also supervise the flow of information between storage locations and the unit computations.

The Chemical Engineering Department of McMaster University, in co-operation with the Canadian General Electric Company, has developed a simple but yet very flexible executive program known as GEMCS (the General Electric-McMaster Simulator) for the simulation of steady-state processes. It has been used successfully in the simulation of several chemical process industries. The extensive work done is well summarized . in the book "Chemical Plant Simulation" (5}. Only a very brief description of the executive program will be attempted here.

Figure 3.1.1. Information Flow Within the GEMCS Executive

The aid of Figure 3.1.1 is invaluable. The executive program first calls an assistant subroutine DLOADl which reads in input streams, equipment parameters and an order of calculation in which to execute the unit computations. The information is then stored for subsequent use. The main program then prepares to execute the first unit computation in the plant. It retrieves the equipment parameters for the unit from the storage location, the EEN vector, via a linking subroutine DISKIO. It then places the equipment parameters (this is done uirectly by DISKIO) into a working equipment vector, the EN vector. The main program then retrieves from the storage location for the streams, the SN matrix, the input streams flowing into the unit computation. Subroutine STREAM(S) acts as the link between the SN matrix and the main program. The input streams are then placed in the working matrix, the SI matrix. The unit computation is then called and executed using information provided in the SI and EN via the subroutine MODULE(NT) locations. Any output streams generated by the unit computation are stored in the working matrix, the SO matrix, and which are then transferred by the main program into the SN storage matrix.

The other unit computations in the calculation order are similarly executed. In the case where there is a recycle of streams, a control subroutine must be placed in the calculation order to direct the main program to recalculate all the

unit computations within the recycle loop, until convergence is reached, within a specified tolerance.

It should be noted that the unit computations deal directly only with the working equipment vector and input and output stream matrices. This creates a uniform pattern whereby the unit computations may be constructed.

3.2 The Stream List

In every process simulation a list of components that will adequately describe any stream within the process has to be selected. Crowe et al {5) have listed several criteria for the selection of stream variables. They include

- (1) The variables should be useful to satisfy the objectives of the simulation,
- (2) All variables in recycle streams are usually important, and
- (3) The unit computations must be able to manipulate the variables to an accuracy consistent throughout the whole system.

These criteria were used as an aid in the selection of the stream variables for the present generalized wastewater treatment plant simulation. In addition, the best features of the stream lists used by Smith (46) and Hoffman (20) were selected and in some cases modified. The stream list used is shown in Table 3.2.1.

Table 3.2.1 Stream List for Wastewater Simulation

It may be noticed that the stream list is divided into three parts. The first part is a list of traditional variables commonly used in wastewater treatment practice. The second part covers the water-insoluble components normally found in wastewaters and the last category includes the water-soluble components. The second and third parts may be further sub-divided into an inorganic and an organic fraction.

The traditional variables in elements $8-12$ are not fundamental components in the sense that they may be derived from the finer classification provided in elements 13-22. However, these are the variables that most operators and environmental engineers understand and hence are invaluable in our communication with them. Another reason for retaining these variables is that several of the correlations used in our simulation are expressed in terms of them. A simple simulation can be effected by the use of only these first twelve variables.

Suspended solids can either be organic or inorganic in nature. The inorganic portion is inert and does not take part in any biological processes, although chemically, it may be altered by a changing pH. However, we have to distinguish between a colloidal inorganic solid, which remain in suspension and a settleable inorganic solid which is removed in physical sedimentation.

For the organic suspended solids, we have to distinguish between the non-living organic objects from the living biomass which is active in bringing about the stabilization of the waste-load. The major portion of the non-living organic solids is biodegradable, while the remainder may be relatively non-biodegradable. The inertness may be due to several factors, one of which is a high degree of polymeric crosslinking. It is impossible to distinguish between the biodegradable portion from the non-biodegradable portion, and the distinction, although provided in the stream list is not used.

Physically, the non-living organic solids can be differentiated by their settling characteristics. We can distinguish a colloidal fraction, which remains in suspension, a settleable fraction and a scum (or floating) fraction.

Organics in solution can also be classified as biodegradable and non-biodegradable. Physically, certain organics in solution may be volatilized by purging with air. These are the solvents and other low molecular weight compounds. The non-biodegradable fraction, being largely composed of high molecular weight compounds, is relatively non-purgeable.

Soluble inorganics of interest include the nutrients phosphorus and nitrogen, and the alkalinity causing ion. Inorganic nitrogen is further subdivided into ammonia and the sum of nitrite and nitrate ions. Several blank spaces

are left in the soluble inorganics section to facilitate the addition of variables, should the simulation be extended to include tertiary treatment.

The units used are also shown in Table 3.2.1. It should be noted that concentration units facilitate the splitting of streams, while mass flow units facilitate the mixing of streams. A disadvantage of concentration units is that while concentrations based on unit volume are usually measured, the simulation system invariably has to use concentrations based on unit mass. Volumetric and mass concentrations are approximately equivalent under 1%, but become increasingly inaccurate above that. To avoid this complication, mass flow units are chosen.

As in Hoffman et al {20), the organic components are all measured in terms of the carbon content. This greatly facilitates the arithmetic of the biological processes.

The interplay being the variables is schematically represented in Figures 3.2.1 and 3.2.2, together with the analytical procedure followed. This is essentially the outline given by Hoffman et al with slight modifications. Further analytical details are given in "Standard Methods" {48).

The conversion of yariables within our "finer" classification to the traditional variables *is* described in Section 9.1.

wastewater

Components

filtrate

Figure 3.2.2, Inter~relationship Between Dissolved

Components

CHAPTER 4

4. PRIMARY SEDIMENTATION

The solid .particles carried in suspension by wastewater flows can be partially removed by gravity settling in a relatively quiescent tank. The process, often called primary sedimentation, is used to reduce the sludge load downstream of the units. In the case where no further treatment is provided, primary sedimentation reduces the formation of sludge banks in the receiving waters. In most cases, it is used as a preliminary step ahead of biological treatment, to reduce the amount of solids being carried around in the latter system.

4.1 Theory

During sedimentation, the solid particles in a dilute suspension, may retain their individual shapes and sizes, and settle out as discrete particles. However, many other solids tend to flocculate (coalesce into a larger particle upon contact with each other), as they settle. This increases their settling velocities and hence the flocculation of the particles is a desirable property from this view-point.

The simpler case of discrete particle sedimentation will be dealt with first, and then it will be expanded to include the case of flocculent sedimentation.
A discrete particle, settling through a fluid, quickly reaches a terminal velocity. At this point, the gravitational force is exactly balanced by the bouyant and drag forces acting on the particle. This leads to the classical equation of discrete particle sedimentation

$$
U_{t} = \begin{bmatrix} \frac{4}{\pi} g & \frac{P_{s} - P}{\pi} \\ 3 C_{D} & P \end{bmatrix}^{1/2} \tag{4.1.1}
$$
\nwhere $U_{t} = \text{terminal velocity (ft/sec)}$

\n
$$
g = \text{acceleration due to gravity (ft/sec2)}
$$
\n
$$
P_{s} = \text{specific gravity of the solid particle}
$$
\n
$$
P = \text{specific gravity of the fluid}
$$
\n
$$
C_{D} = \text{drag coefficient}
$$
\nand $D_{p} = \text{diameter of the particle (ft)}$

The drag coefficient C_D is a function of the particle Reynolds Number and of the sphericity of the particle. This can be obtained from most standard texts on Fluid Mechanics.

Using the fact that a discrete particle quickly reaches a terminal velocity, during sedimentation, we can calculate the removal obtained in a batch settling test.

Consider the sketch of a batch settling column in Figure 4.1.1. If a dilute suspension of discrete particles occupies the entire volume uniformly, at the start of the test, then if a sample is drawn off at a depth h at time t, it will not contain any particles with terminal velocities greater

Figure 4.1.1. Settling Column

than $U = h/t$. The fraction of the solids concentration in the sample to the original solids concentration will be the weight fraction of particles having terminal velocities $U~\leqslant~h/t$. Let this weight fraction be represented by x , and $x = f(u)$ - may be used to represent the settling characteristics of the particles. Figure 4.1.2 shows a typical settling velocity analysis curve for a suspension of non-flocculating particles.

If the liquid above a depth N_{α} were to be decanted, at time t, it will only contain particles with terminal velocities $\langle h_{\alpha}/t = U_{\alpha}$. But particles with terminal velocities U $\langle U \rangle$ will have fallen through a distance h = Ut, and hence only a fraction $(h_o - h)/h_o = (U_o - U)/U_o$ of such particles will remain in the decanted liquid.

Hence the weight fraction of the solids in the decanted (or clarified) liquid will be

$$
W_{C} (U_{O}) = \int_{O}^{X_{O}} (U_{O} - U)/U_{O} dx
$$

= $x_{O} - \int_{O}^{X_{O}} U/U_{O} dx$ (4.1.2)

where W_{α} = weight fractions of solids in decanted liquid x_0 = weight fraction of particles having terminal velocities $U \leq U_0$.

and U_{α} = sometimes referred to as overflow velocity.

The weight fraction of solids not in the decanted liquid (or the weight fraction removal) is then given by

$$
W_r
$$
 (U_o) = 1 - W_c (U_o) = (1 - x_o) + $\int_o^{x_o} U/U_o dx$ (4.1.3)

The integral may be approximated by a summation term and hence equations 4.1.2 and 4.1.3 may be rewritten as

$$
W_{C} (U_{O}) = X_{O} - \sum_{x=0}^{x=x_{O}} U/U_{O} \Delta x
$$
 (4.1.4)

and
$$
W_r
$$
 (U_o) = (1 - x_o) + $\sum_{x=0}^{x=x_o}$ U/U_o Δx (4.1.5)

The integral or the summation term can be evaluated with the use of Figure 4.1.2.

If, during the batch settling test, samples are withdrawn at various depths and elapsed times, Figure 4.1.3 will result. The lines shown join points of equal weightfraction. The slopes of the lines will give the associated terminal velocities.

The weight fraction of solids removed can also be evaluated from Figure 4.1.3. Equation 4.1.5 may be rewritten, for that purpose, as

$$
W_{r} (U_{0}) = (1 - x_{0}) + \sum_{x=0}^{x=x_{0}} h/h_{0} \Delta x
$$
 (4.1.6)

From Figure 4.1.3, if h_A is the average of h_O and h_1 , h_{β} the average of h_{1} and h_{2} , and so on, then the summation term can be evaluated as

Figure 4.1.4. Settling Curves for Flocculating Solids

$$
\sum_{x=x_0}^{x=x_0} h/h_0 \Delta x = \frac{1}{h_0} \left\{ h_A(x_0 - x_1) + h_\beta(x_1 - x_2) + H_C(x_2 - x_3) + h_D(x_3 - 0) \right\}
$$
 (4.1.7)

In the case where the particles tend to flocculate upon contact with each other, the settling velocities of the particles are increased, as sedimentation proceeds. Hence there is an added effect of the settling time.

The effect of flocculation is best seen in Figure 4.1.4, which is obtained in the same manner as Figure 4.1.3. The lines joining points of equal weight-fraction are no . longer straight, as in the case of discrete particle sedimentation, but curved. The curvature reflects the increasing settling velocity, as the sedimentation proceeds.

Equation 4.1.2 to 4.1.6 still holds and may be used . to evaluate the solids removal. Also, a similar procedure, as the one used in obtaining Equation 4.1.7, may be used as a graphical solution.

The batch settling test can be directly applied to a continuous flow primary settling tank, assuming plug flow conditions exist in the tank. The depth $h_{\rm o}$ refers directly to the depth of the tank and the time t is the residence time of the fluid in the tank. Deviations from plug flow conditions are treated in section 4.3.

4.2 The Correlation of Batch Settling Data

Raw sewage from the Drury Lane, Burlington and Dundas Sewage Treatment Plants were used in the settling experiments. The settling column is 8 in. in diameter and 8 ft. in height. The column has five sampling points, equally spaced 1 ft. apart.

The raw sewage was introduced into the column and allowed to equilibrate to room temperature for at least four hours. This was done to minimize thermal convection currents. Then the suspension was stirred up by blowing air into the sewage for one minute.

Samples were withdrawn initially, just after the mixing, and at regular time intervals from the five sampling ports. Prior to sampling, the sampling lines were flushed to wash out any accumulated solids. The column.was also left for 24 hours, and the suspended solids determined. This represented the non-settleable or colloidal portion of the waste. The suspended solids of the samples were determined by vacuum filtration on 0.45 micron membrane filter papers. The papers were dried in an oven set at 103° C for at least half an hour. A Mettler balance with an accuracy of 0.1 mg was used for the weighing. All determinations were done in duplicate, and averaged.

The suspended solids data are given in Appendix A. Two previous runs, performed in 1969 and 1970, as part of a

wastewater course, using raw sewqge from the Dundas Sewage Treatment Plant are also included. Figure 4.2.1 to 4.2.4 show the weight fraction of settleable solids *in* the samples withdrawn as a function of time and depth. Equation 4.1.6 was used to calculate the weight fraction of settleable solids removal as a function of depth and time, and the results are tabulated *in* Appendix A.

In the absence of theoretically derived expressions, empirical correlations had to be derived. However, we note that at constant h_{α} ,

if $t \rightarrow \infty$, then $W_C \rightarrow 0$ or $W_T \rightarrow 1$ and if $t \rightarrow 0$, then $W_c \rightarrow 1$ or $W_r \rightarrow 0$. (4.2.1)

Also that at constant t,

if $h_0 \rightarrow \infty$, (or $U_0 \rightarrow \infty$), then $W_c \rightarrow 1$ or $W_c \rightarrow 0$ and if $h_0 \rightarrow 0$ (or $U_0 \rightarrow 0$), then $W_c \rightarrow 0$ or $W_r \rightarrow 1$ (4.2.2)

This suggests a relationship of the form

$$
W_{\rm C} = e^{-kt^{\alpha}/h_{\rm O} \beta} \qquad (4.2.3)
$$

or
$$
W_r = 1 - W_c = 1 - e^{-kt^{\alpha}/h_0^{\beta}}
$$
 (4.2.4)

By comparing Equations 4.2.4. a .d 4.2.2 we note the similiarty between our model and Silveston's model. However, Si1veston has assumed that $\alpha = \beta = 1$ (note: GPS α α). t

Figure 4.2.1. Fraction of Settleable Solids Versus Depth and

 $\frac{21}{1}$

Figure 4.2.2. Fraction of Settleable Solids Versus Depth and

Time (minutes)

 $\frac{2}{5}$

Figure 4.2.3. Fraction of Settleable Solids Versus Depth and

Time

RUN A.3

Time (minutes)

ပ္ပ

Figure $4.2.4.$ Fraction of Settleable Solids Versus Depth and

Time

RUN A.4

Time (minutes)

 $\frac{2}{4}$

Figure 4.2.5. Comparison of Removals Obtained from Batch Settling

and from Correlation

Correlation

from

Calculated

imoval

Removal Obtained from Batch Settling

Comparisons of Removals Obtained from Batch Settling Figure 4.2.6. and from Correlation

from Correlation

Calculated

Removal

Removal Obtained from Batch Settling

Comparison of Removals Obtained from Batch Settling Figure 4.2.7.

and Correlation

Removal Calculated from Correlation

Removal Obtained from Batch Settling

and Correlation

Removal Obtained from Batch Settling

A three parameter search, using a SIMPLEX routine was then used to determine k, α and β . The objective function uses a least sum of squares criterion, the sum of squares being \sum_{r} (W_r experimental - W_r correlation)². The values of k , α and β are summarized in Table 4.2.1 and the correlated data are shown in Figures 4.2.5 to 4.2.8.

TABLE 4.2.1. Parameters for Quiescent Settling Model (Note: t is in minutes and h_0 is in feet.)

The correlations obtained are good and the closeness in the values of the parameters indicate that the raw waste from the two sources are not too different. The average values of k, α and β obtained are 0.226, 0.494, and 0.239.

4.3 Deviations from Ideal "Plug Flow" Settling

Conditions in a primary settling tank are seldom of the ideal "plug flow" nature. Turbulence and shortcircuiting in the flow can be caused by several factors, and their effects on sedimentation have to be accounted for.

Short-circuiting in the flow results mainly from density currents, which are bulk movements of fluids caused by density variations. Temperature differences between the feed and the water in the tank, or the presence of high suspended solids in the feed, will cause these density differences to occur.

Due to poor inlet design, the kinetic energy of the incoming flow can be quite considerable. The resulting energy dissipation can cause turbulent conditions to exist in the inlet zone. Inlet baffles, if properly designed, can reduce the kinetic energy to a large extent.

Several investigators have examined the effects of turbulence on settling. A very good summary of the various approaches used can be found in Fitz (15). Ensign Dobbins (6), starting from the Continuity Equation, made several assumptions and arrived at a simplified expression, for the case of discrete particle sedimentation

$$
\frac{dC}{dt} = e \frac{d^2C}{dy^2} + U \frac{dC}{dy}
$$
 (4.3.1)

where $C =$ concentration of solids having settling velocity U

- $U =$ settling velocity
- $y =$ depth
- $t = time$

and $e = eddy diffusivity$.

Dobbins has obtained a solution for Equation 4.3.1, but it is too complex for use in a simulation model. A graphical solution of Equation 4.3.1 is available (4), and could be used in conjunction with the batch settling test as outlined in Section 4.2.

Another approach to the problem would be to obtain information on the flow patterns in the tank from a residence time distribution analysis. Fitz (15) used mixed models of combinations of stirred tanks, plug flow volumes and dead spaces to fit the residence time distributions thus obtained, to predict solids removal. His results were not too encouraging.

A dispersion model also does not appear to be promising as it rarely fits the observed exit age distributions. This is to be expected as the dispersion model assumes only relatively small deviations from plug flow with no gross short-circuiting or density currents.

In an attempt to estimate the degree of turbulence in the primary tanks of the Dundas and Drury Lane Water Pollution Control Plants, tracer studies were carried out, using Rhodamine B dye and a Fluorometer, a Turner and Associates Model III. The experimental technique *is* well described by Wilson (49). The exit age distributions were followed for at least two residence times. Samples of the influent and effluent were collected during the tracer studies and analyzed for suspended solids.

Eight runs were made, four at Dundas and four at Drury Lane. Due to pump and recorder difficulties, two runs had to be discarded, one from each of the two plants. The exit age distributions are given *in* Appendix B. The tracer recoveries were poor and were approximately 80-85%. The calculated residence times from the exit age distributions were much lower than the actual fluid residence times. The tracer results are too unreliable for any use.

However, there seemed tobe a consistent efficiency factor for each of the two plants, where the efficiency factor is defined as

observed % of settleable solids removed ¢ ⁼ theoretical (batch) % of settleable solids removed

(4.3.2)

A high efficiency factor will indicate a low degree of turbulence or short-circuiting, while a low effiency factor indicates a highe level of turbulence or short-circuiting.

The theoretical or batch removal is calculated from the equation developed in Section 4.2. The efficiency factors obtained are summarized in the table below:

TABLE 4.3.1. Efficiency Factors for the Primary Sedimentation Tanks at the Dundas and Drury Lane Treatment Plants.

The average efficiency factor for the primary settlers at Dundas is 0.57 whilst that at Drury Lane is 0.85. The high degree of turbulence in the primary settling tank at Dundas is reflected in the low efficiency factor and is quite evident visually from the effluent turbidity.

Our overall correlation·for the primary settler is then described by

$$
W_{r} = \emptyset \Big\{ \left| \exp \left(-k \frac{t^{\alpha}}{H} \right) \right\} \tag{4.3.3}
$$

4.4 The Primary Settling Tank Model - PRISETl

A listing of the program *is* given *in* Appendix K. The equipment vector for the model *is* reproduced below.

The computations carried out by the program can be explained *in* the following steps:

- (1) The pH of the output streams is initially calculated.
- (2) Input streams 2 and 3 are sludge streams carrying solids which have much higher settling velocities than the solids *in* the raw waste. Hence, the removal of the sludge solids *is* essentially complete. EN{22) provides

 $\check{\epsilon}$

for the small fraction of the sludge solids which do escape over the wiers due to short-circuiting or turbulence.

- (3) The removal of the solids in the raw waste is calculated from Equation 4.3.3.
- (4) The underflow is calculated from pump capacities. The concentration of the underflow solids is then calculated, for which a maximum value of 6% is assumed. If the pump capacity is insufficient, excess solids over 6% is carried over to the effluent.
- (5) The scum flow is then calculated, assuming 90% removal of scum solids. The solids concentration of the scum flow is specified by EN(20).
- (6) Soluble components are then split according to the split in water flow among the three output streams.

CHAPTER 5

5. AEROBIC BIOLOGICAL SLUDGE PROCESSES

The theory here represents an attempt to model all aerobic biological sludge processes in one general package. Currently, all the various modifications of the activated sludge process, such as extended aeration, step aeration, contact stabilization and aerobic digestion, have been considered as separate processes. However, a bacterial cell does not know what hydraulic regime it is in, nor does it know what process it is supposed to work under. The kinetics describing the activity of the bacteria should be similar in all cases. The differences in rates observed with the various processes can only be due to differences in composition of the activated sludge mass.

5,1 The Bacteria and its Activities

The morphology of the bacteria is well described in any standard textbook on microbiology. A very brief description will be given here, while the physiological functions of the bacteria and their corresponding kinetic expressions will be described in more detail.

Bacterial cells are small and have sizes of the order of one micron. The cell shape may be spherical (coccus), rod-like (bacillus), or spirally-shaped (spirillum), and is defined to a large part by a relatively tough outer cell wall.

Inside the cell wall is a semi-permeable cytoplasmic membrane, which is responsible for a selective transfer of nutrients and waste products into and out of the cell. The region within the cytoplasmic membrane includes the cytoplasm and the nuclear region. The cytoplasm is characterized by a relatively high concentration of ribonucleic acid (RNA), which is associated with protein synthesis. The nuclear region contains a high concentration of deoxyribonucleic acid (DNA). The genetic code is stored in the DNA. Some bacteria secrete slimy materials on their surfaces, creating a slimy layer outside the cell wall. A rough sketch of a bacterial cell is shown in Figure 5.1.1.

The activitiy of the bacteria will be described in the following four sections.

(a) Metabolism:

Bacterial cells, like all living cells are capable of feeding and growing. Soluble organic matter (substrate) diffuses into the bacterial cell where they are metabolized and converted partly into new cell matter. The remainder of the organic substrate is respired to provide energy for the cell. The respiration products are mainly carbondioxide and water.

Schematically, this may be represented as:

New cell matter
Superintion products soluble degradable $+$ microorganisms $+$ O₂

The yield factor, Y , is defined as the fraction of soluble carbon metabolized which goes into new cell growth. The fraction (1-Y) which is respired will require an equivalent amount of oxygen.

The rate of soluble carbon degradation can be \'lritten as

rate =
$$
\frac{dC}{dt}
$$
 = - UM (5.1.1)

where $C =$ concentration of soluble degradable carbon

$$
U = f(c) = \text{unit growth rate}
$$

and $M =$ concentration of microorganisms

(as equivalent carbon)

For the case of a single substrate and a pure culture, the functional dependence of u is described by Michaelis-Menton kinetics as

> $U = \frac{U_{\text{max}} C}{(5.1.2)}$ $C^* + C$

where $U_{\text{max}} = \text{maximum growth rate}$

and C^* = constant = value of C at which $U = \frac{1}{2}U_{\text{max}}$

However, in waste treatment, mixed substrates are generally encountered. Pure cultures are also difficult to maintain and is often less efficient than mixed cultures. Consequently, there is m need to restrict the description of U by Michaelis-Menton kinetics. The measurements of

U and Y as described in Section 5.2 show that a linear correlation

$$
U = k_{4}C \qquad (5.1.3)
$$

is a better fit. It should also be noted that the Michaelis-Menton equation becomes linear when C << C*.

(b) Endogenous Respiration

Bacterial cells can also utilize stored food products (bacterial fat) to provide energy for its various physiological activities, especially in perids of low external food supplies. This is known as endogenous respiration or sometimes as basal metabolism.

Schematically, this may be represented as

stored "bacterial fat" + $0₂$ \longrightarrow Respiration products (5.1.4)

The rate of endogenous respiration should be proportional to the number of bacterial cells present, or is first order with respect to the bacterial concentration, namely,

rate =
$$
\frac{dM}{dt}
$$
 = - k₁M (5.1.5)

where k_1 = endogenous respiration rate constant.

(c) Death and Cell Lysis

In cell lysis, the inner protoplasm of the cell is released to the surroundings through a breach in the cell wall. Part of the protoplasm is available as food for other microorganisms, while the remainder consists of relatively

non-biodegradable high molecular weight matter, schematically, this may be shown as

[~]is the fraction of soluble degradable carbon, and β is the fraction of soluble non-degradable carbon released during cell lysis. The remaining fraction $(1-\frac{\alpha-\beta}{\beta})$ is particulate matter consisting of cell walls, slime layers, various membranes and so on, with varying degrees of·biodegradability.

This process explains the build-up of soluble nondegradable carbon during the prolonged aeration of activated sludge.

The rate of death and subsequent cell lysis should also be proportional to the number of bacterial cells present:

rate =
$$
\frac{dM}{dt}
$$
 = - k₂M (5.1.7)

where k_2 = lysis rate constant.

(d) Solubilization of Particulate Organics

In general, solid organic matters have to be solubilized . before they can be utilized by the bacteria. This process is speeded up by the secretion of exo-enzymes by the bacterial cells. The solubilized organic matter then diffuses back into the bacterial cell as substrate.

The rate of solubilization should be proportional to the concentration of the organic solids. It should also be proportional to the rate of release of exo-enzymes or equivalently, to the number of bacterial cells present:

$$
\text{rate} = \frac{dP}{dt} = -k_3PM \qquad (5.1.8)
$$

where $P = solid$ organics concentration

(as equivalent carbon),

 k_2 = solubilization rate constant. and

The four reactions proposed above involves principally five components, namely the microorganisms (M), the particulate organics other than microorganisms (P), soluble degradable organic carbon (SDOC), soluble non-degradable organic carbon (SNDOC) and carbon dioxide. However, carbon dioxide is partially purged from the system by the air and hence is difficult to monitor. But, it can always be obtained from a mass balance on carbon.

The total soluble organic carbon, SDOC+SNDOC, can be determined by means of the carbon analyzer. The soluble degradable organic carbon can be obtained from the Biochemical Oxygen Demand of the filtrate, using the relationship

 $SDOC = BOD (filterate)/1.9$ (5.1.9) This is discussed in Section 9.1. The SNDOC is then obtained by difference.

The total concentration of microorganisms and particulate solids, $M + P$, can be obtained from the volatile suspended solids using the relationship (see also Section (9.1)

$$
M + P = VSS/2.1 \t\t(5.1.10)
$$

The concentration of microorganisms alone can be indirectly measured from its oxygen uptake rate. In addition to the oxygen required for the carbon respired, oxygen is also consumed in the nitrification of ammonia-nitrogen associated with the sludge. A chemical analysis of activated sludge reports an average composition of $C_5H_7O_2N$ (54). The oxidation reaction can then be written as:

 $C_5H_7O_2N + 7O_2 \longrightarrow 5CO_2 + 3H_2O + HNO_3$ (5.1.11) Hence each gram of carbon lost through respiration requires 3.73 grams of oxygen or

$$
OUR = 3.73 [k_1 + (1-Y)U]M
$$
 (5.1.12)

therefore,

$$
M = \frac{OUR}{3.73 [k_1 + (1-Y)U]}
$$

Hence, our aerobic biological system involves four simultaneous, kinetically independent reactions, five components and seven constants. The constants involved are k_1 , k_2 , k_3 , k_4 , Y , α and β .

5.2 Chemostat Experiments

The purpose of this series of experiments is to evaluate the functional dependence of the unit growth rate on the soluble degradable carbon concentration. A chemostat is essentially a biological reactor that acts as a continuous-flow, completely stirred tank reactor. The defining equations are easily derived from a mass balance around the reactor

In Figure 5.2.1, Q is the flow rate through the reactor and V the volume of the reactor. The concentration of soluble degradable organic carbon in the feed is C_{α} and that in the effluent is C. The concentration of microorganisms in the feed is assumed to be zero while that in the vessel or effluent is M (as equivalent carbon).

Performing a mass balance on the soluble carbon, we have at steady-state,

FIGURE 5.2.1. A Schematic Representation of a Chemostat, and Operating Variables.

where $U =$ unit growth rate (see Section 5.1)

Hence
$$
U = f(c) = \frac{(C_0 - C)}{M\bar{t}}
$$
 (5.2.2)

where $\bar{t} = V/Q$ = residence time. Also from definition, the yield factor

 $= Y = M/(C_0 - C)$ (5.2.3)

In our experiments, settled raw sewage was filtered through a two-inch thick layer of glass wool and used as the feed. This was stored in a constant head bottle and allowed to drip into an aerated 6 litre vessel (see Figure 5.2.2). Seeding was accomplished by the addition of about 100 ml of activated sludge to the reaction vessel. After two detention times, the effluent was sampled and analyzed regularly, until a steady-state was reached.

The results from seven runs are summarized in Appendix c. The average value of the yield factor Y obtained ·is 0.57 with a standard deviation of 0.04. the functional dependence of U is found to be linear with respect to the soluble degradable carbon concentration, in the range studies $(25 - 50 \text{ mg}/1)$. This is plotted in Figure 5.2.3. The correlation obtained is

 $U = 0.0010 C = k_AC$ (5.2.4) with a correlation coefficient of 0.85.

Concentration

Soluble Degradable Carbon (mg/1)

 $\overline{57}$

We have assumed that the solids in the feed is not significantly solublized in the reactor.

To compare the results obtained we have to convert Equation S.l.2 to traditional units:

We have
$$
\frac{dC}{dt} = \frac{d \text{ BOD}_5}{dt} \times \frac{1}{1.9}
$$

It is estimated that microorganisms account for approximately 30% of the Mixed Liquor Suspended solids. Hence $M \times 2.1 = 0.3$ (MLSS)

Therefore, Equation S.1.2 becomes

$$
\frac{1}{1.9} \times \frac{dBOD_5}{dt} = -k_4 \times \frac{BOD_5}{1.9} \times \frac{0.3}{2.1}
$$
 (MLSS)

or $\frac{\text{dBOD}_5}{\text{dBOD}_5}$ = - k₄ $\frac{0.3}{\text{dBOD}_5}$ (BOD₅) (MLSS) dt 2.1

$$
= - K_{s}^{"}
$$
 (BOD₅) (MLSS) (5.2.5)
or $K_{s} = k_{4} \frac{0.3}{2.1}$

 $= 0.00014 \text{ (hr)}^{-1} \text{ (mg/1)}^{-1}$

Eckenfelder (2) reports a value of

 $K_{\rm c}$ = 0.00010 - 0.00013 (hr)⁻¹ (mg/1)⁻¹
Keshayan et al (22) reports a value of

 $K_a = 0.00016$ (hr)⁻¹ (mg/1)⁻¹

Hence, our results are approximately in agreement with the literature values.

5.3 Aeration Studies of Activated Sludge

As some of the rate proposed in Section 5.1 are expected to be slow, batch studies of aerated activated sludge have to be carried out over an extended period. Return activated sludge is a convenient "reaction mixture" to use, as it contains all the components in our reaction set.

Return activated sludge from the Drury Lane plant was used, and the aeration was carried out in a 6-litre vessel, for twenty five days. Samples were taken periodically, and the following analyses performed:

(1) Biochemical Oxygen Demand of filtrate,

(2) Dissolved Organic Carbon,

(3) Suspended Solids, and

(4) Oxygen Uptake rate.

and at the end. Inorganic solids were determined both initially

Two runs were made with different samples of return activated sludge from the Skyway Plant, Burlington. The results obtained are tabulated in Appendix D. The plots of (P + M), OUR, SDOC and SNDOC versus time are shown

in Figures $5.3.1 - 5.3.6$. The volatile suspended solids and the oxygen uptake rate were observed to drop with length of aeration. The soluble degradable organic carbon dropped slightly and then remained fairly constant. There was a marked increase in soluble non-degradable organic carbon with aeration in both cases.

To obtain an estimate of the variances in the measurements, a third run was started and allowed to run for a week. Then five samples were withdrawn simultaneously from the reaction vessel and analyzed immediately. The results are also tabulated in Appendix D, together with the variancecovariance matrix and its inverse matrix.

Before the parameter search was performed, an eight constant $\Omega_{_{\mathbf{O}}}$ was introduced and it is defined as the ratio of the initial concentration of microorganisms, M_0 , to that of the total volatile solids, namely,

$$
Q_{\text{O}} = \frac{M_{\text{O}}}{(P + M)_{\text{O}}} \tag{5.3.1}
$$

where the subscript $_0$ refers to initial values. Otherwise, we have to calculate $M_{\overline{O}}$ from the intial value of the oxygen uptake rate, and as M_{\odot} is a critical value, an error in the oxygen uptake rate can have a marked effect on the search.

A fourth order Runge Kutta was used to calculate the values of the four components, P, M SNDOC, and SDOC. The

Solids (mgC/1)

Suspended

Volatile

Aeration Time (days)

L9

 \mathbf{I}

Aeration Time (days)

 $\frac{8}{3}$

Aeration Time (days)

င္ပ

Soluble Degradable Carbon (mg/1)

Aeration Time (days)

თ \cdot

-~

Aeration Time (days)

Soluble Degrad $\frac{1}{2}$ w

ტ

set of differential equations used were

$$
\frac{dP}{dt} = \begin{bmatrix} -k_3 P + (1-\alpha-\beta)k_2 \end{bmatrix} M
$$

with an initial value of $(1-Q_0) (P+M)_0$ (5.3.2)

$$
\frac{dM}{dt} = \begin{bmatrix} k_4 & (\text{SDOC})Y - (k_1 + k_2) \\ \text{with an initial value of } Q_0 \text{ (P+M)}_0 \end{bmatrix} M \tag{5.3.3}
$$

$$
\frac{\text{dsmDC}}{\text{dt}} = \beta k_2 M
$$
\ndt with an initial value of (SMDC), (5.3.4)

and
$$
\frac{dSDOC}{dt}
$$
 = $\begin{bmatrix} k_3 P - k_4 (SDOC) + k_2 \alpha \end{bmatrix} M$.
with an initial value of (SDOC)_O (5.3.5)

A SIMPLEX optimization routine was used and the objective function to be minimized *is* (18)

$$
\phi = \sum_{i=1}^{n} \sum_{r=1}^{4} \sum_{s=1}^{4} \sigma^{rs} (y_{ri} - \eta_{ri}) (y_{si} - \eta_{si})
$$
 (5.3.6)

where ϕ = objective function to be minimized $n =$ number of sets of observations made, σ^{rs} = the (r,s) th element of the inverse of the variance-covariance matrix,

 y_{ri} = the ith observation of the rth response and η_{ri} = the ith prediction of the rth response.

(

The four responses used are:

- (1} Volatile suspended solids (mgC/1)., with the predicted value calculated from P + M
- (2) Oxygen uptake rate (mgO₂/l/hr), with the predicted value calculated from $3.7M k_1+(L-Y)k_4$ (SDOC)
- {3} Soluble non-degradable organic carbon, SNDOC, (mgC/1), and
- (4} Soluble degradable organic carbon, SDOC, (mgC/1)

A schematic representation of the parameter search *is* shown *in* Figure 5.3.7.

FIGURE 5.3.7. Schematic Representation of

Parameter Search

The values of the constants obtained are shown in Table 5.3.1. The fitted curves are shown in Figures 5.3.1 to 5.3.6, together with the observed points.

Table 5.3.1. Parameters for the Aerobic Biological Sludge Process Model

The agreement in the values of the parameters in both runs are very good, considering the analytical errors involved. From the Chemostat experiments, the values of k_A and Y obtained are 1.0 x 10^{-3} and 0.57. The agreement in the yield factor is good, but the metabolism rate factor is much lower in the "endogenous phase" of the extended aeration studies. The results imply that the metabolism rate is retarded under conditions of prolonged starvation.

Sedivy (37) in his thesis on residual organics found β to be about 1-5%, using glucose as substrate. Our value is a bit high. The value of θ obtained suggests that only about half of the volatile suspended solids *in* the return sludge from the Skyway Plant, is actually "active mass".

No equivalent values for the other constants exist *in* the literature. Note that the basal metabolism rate constant kd proposed by other workers *ia* actually measured from the rate of decrease of volatile suspended solids and which *in* our case is a combination of k_1 , k_2 and k_3 . To illustrate that our reaction scheme fits the rate of decrease of volatile suspended solids better than the simple exponentialdecay, semi-log plots are shown *in* Figure 5.3.8 and 5.3.9, of the volatile solids versus time. In both runs, our model agrees with the exponential decay model up to about an aeration time of ten days. Beyond that the exponential decay model predicts much lower solids than *is* actually observed. Our model gives a much better fit beyond ten days, although it still predicts lower solids than is actually observed. This *is* probably because our model still lacks the ability to predict the concentration of particulate non-degradable organic carbon, which would build up with extended aeration.

As a matter of interest, the k_d value from our data, which is 0.055 day^{-1} (base e) agrees with Pipes and Meade (31) very well. Their data showed a range of k_d between 0.025 to 0.078 day⁻¹ with an average value of 0.053 day⁻¹. Eckenfelder (2) reports a value of 0.048 day $^{\textnormal{\texttt{-1}}}.$

As the statistical calculations are quite difficult, the estimation of confidence limits for the various constants

Figure 5.3.8. Log (Volatile Suspended Solids) Versus Time

Aeration Time (days) $\overline{2}$

 \tilde{c}

will not be attempted. Instead, the average sum of squares deviation from the regression line will be compared to the variance obtained in the replicate samples. This is presented in Table 5.3.2.

Table 5.3.2. Comparison of Average Sum of Squares Deviation from Regression Line, with the Variance from Replicate Samples

The average sum of squares from the regression line compares quite well with the variance from the replicate analysis. This indicates that the model proposed *is* significant.

No measurements were made on the effect of temperature on the various rates. The above two runs were made at 23° C + 2° C. A temperature correction factor of 1.047 will be used, as proposed by several authors (2, 47). Hence,

> $k_1(T) = 0.0026 (1.047)^{T-23}$ $k_2(T) = 0.0015 (1.047)^{T-23}$

 $\frac{1}{4}$

 $k_3(T) = 2.0 \times 10^{-6} (1.047)^{T-23}$ $k_A(T) = 0.0010 (1.047)^{T-23}$ or $k_4(T) = 0.0006(1.047)^{T-23}$ in food limiting conditions.

(5.3.7)

5.4 The Stripping Rate of Purgeable* Soluble Organics

Purgeable soluble organics tend to be stripped off. *in* the aeration process. Hence their removal can occur both by biological action and by physical stripping due to the passage of air (16, 17).

According to mass transfer principles, the mass transfer will depend on the first order of the driving force, or concentration difference and also on the surface area of the air bubbles, namely,

$$
\frac{\text{dspoc}}{\text{dt}} = -k_{\text{L}} \frac{\text{A}}{\text{V}} \quad (\text{SPoc} - \text{SPoc}_{\text{b}}) \tag{5.4.1}
$$

where SPOC = soluble purgeable organic carbon *in* solution SPOC_k= soluble purgeable organic carbon in bubble k_{r} = a mass transfer coefficient $A = surface area of bubbles$ and $V = volume of reactor$

Waste treatment terminology reserves the use of the word "volatile" for volatiles at 600°C. The term "purgeable" *is* then used here to define the soluble organics which can be physically removed by purging with a gas, at normal temperatures.

But
$$
A/V = a_{V}q
$$
 (5.4.2)

where a_{17} = surface area to volume ratio of air bubbles and α = volumetric rate of air per unit volume of reactor.

Hence,

$$
\frac{d(SPOC)}{dt} = -k_L a_V q. (SPOC)
$$
 (5.4.3)

$$
= - k5 q. (SPOC)
$$
 (5.4.4)

Solving,

$$
1n \quad \frac{\text{(SPOC)}}{\text{(SPOC)}} = -k_5 q t \quad (5.4.5)
$$

where $(SPOC)$ = initial value of SPOC. $(SPOC)$

To estimate the rate of stripping, settled raw sewage was filtered through a two-inch layer of glass wool and aerated at a fixed rate in a 2 litre vessel (no activated sludge was added). The soluble organic carbon was measured at regular intervals, until no further drop was observed. The remaining soluble organics *is* non-purgeable. The difference between the soluble organic carbon and the ultimate value (or the non-purgeable organic carbon) is the soluble purgeable organic carbon (SPOC).

Four air rates of 0.5, 1.0, 1.5 and 2.1 cu. ft. air/ hr/cu. ft. reactor were used and the results are tabulated in

Figure 5.4.1. Semi-Log Plot of SPOC Versus Time

Time (minutes)

Time (minutes)

Appendix E. Semi-log plots of SPOC versus time are shown in Figures 5.4.1 to 5.4.4. The slopes of the lines will be $-k_5q$. Table 5,4.1. shows the values of k_5 obtained from the four runs. The average value of k_5 is 2.06 with a standard deviation of 0.27.

5.5 Oxygen Transfer Efficiencies

The rate of oxygen transfer can be found by using the same equation as that for the stripping of purgeable organics, namely

$$
\frac{d [o_2]}{dt} = k_L' \frac{A}{v} ([o_2]_{sat} - [o_2])
$$
 (5.5.1)

where $\begin{bmatrix} 0 \\ 2 \end{bmatrix}$ = concentration of oxygen in liquid $[0_2]_{sat}$ = saturation concentration of oxygen in liquid $k_{r}^{'}$ = liquid film mass transfer coefficient $A = surface area of air bubbles$

and $V =$ volume of aeration tank

The saturation concentration of oxygen in water at 1 atm can be calculated from the equation (9)

$$
O_{2 \text{ sat.}, 1 \text{ atm.}} = 14.16 - 0.3943T + 0.007714T^{2}
$$

$$
- 0.0000646T^{3}
$$
(5.5.2)

where $T =$ temperature in ^{0}C

 $k_{\rm T}^{\rm T}$ A is a function of the aeration device, the air flow rate and of the depth of the ^aeration tank. The functional relationship is

$$
k_A^{\dagger} = N \cdot G^{(1-n)} H^{(1-m)}
$$
 (5.5.3)

where $N =$ sometimes referred to as absorption number

 $G =$ total air rate, scfm.

 $H =$ depth, ft.

and $n,m = constant$

Table 5.5.1 shows the various values of the constants, for several aeration devices (9).

5.6 Nutrient Balance

The assumption made is that the nitrogen to carbon (N/C) and phosphorus to carbon (P/C) ratios in the raw waste is the same for both the particulate and soluble organic

Unit			N (1-n) (1-m)
Aloxite tubes	270	0.85	0.45
Sparjer	210	0.86	0.78
Seran tubes	275	0.8	(0.60)
Carborundum plates	65	0.8	(0.75)

Table 5.5.1. Oxygen Transfer Characteristics of Some Common Diffused Aeration Equipment.

components. This is supported by analyses made by the Ontario Water Resources Commission for the Drury Lane W.P.C.P. This is shown in the table below.

Table 5.6.1. Average Nitrogen, Phosphorus and Carbon Concentrations in Raw Sewage from Drury Lane W.P.C.P. The Nitrogen and Phosphorus Values are Obtained from 29 Analyses by the o.w.R.C. The carbon concentrations are

estimated from our analyses.

The composition of the bacterial cells is assumed to be $C_gH_7O_2N$ (54). This has a (N/C) ratio of 0.23. The {P/N) ratio in activated sludge is reported to be about 0.25 (9), hence (P/C) is approximately 0.06 .

After the various effluent organic carbon components have been calculated, a mass balance over nitrogen and phosphorus is then performed. All excess nitrogen is assumed to be converted to ammonia and all excess phosphorus to inorganic phosphates.

Downing's data for the rate of nitrification will be used (7);

$$
\frac{dNH_3}{dt} = - \frac{G. NH_3 \cdot X}{Y (K + NH_3)}
$$
 (5.6.1)

and $\frac{dX}{dx} = -y \frac{dS}{dx}$ (5.6.2) dt dt

where NH_3 = ammonia concentration (mgN/1) $X =$ Nitrosomonas concentration (mg/l) $G = 0.014 hr^{-1}$ $K = 1.0$ mg/1

and $Y = 0.05$ gms Nitrosomonas/gm NH_{3}^{-N}

A slight modification will be made to Downing's equation. This is to account for a decrease in the rate due to an oxygen concentration of less than 1.0 mg/1. The retarding factor f is defined·where

$$
f = 1 \text{ for } [0_2] \ge 1.0 \text{ mg/l}
$$

and $f = [0_2] \text{ for } 0.0 \le [0_2] \le 1.0 \text{ mg/l}$ (5.6.3)

i.e.,

$$
\frac{dNH_3}{dt} = - \frac{f.G. NH_3.X}{Y (K + NH_3)}
$$
 (5.6.4)

The mass of Nitrosomonas is found iteratively and stored in the EN vector as a fraction of the mixed liquor suspended solids mass.

The oxygen requirement during nitrification can be obtained from the equation

 NH_3 + 20₂ \rightarrow HNO₃ + H₂O (5.6.5)

Hence, for each gram of NH_{3}^{-N} converted to NO_{3}^{-} , 4.57 grams of oxygen are required.

· 5.7 The Aerobic Biological Sludge Process Model- ACTSLl and CSTR2

CSTR 2 is a continuous-flow, completely-stirred tank reactor model of the aerobic biological sludge process. ACTSLl is a stirred tanks in series model which calls the basic CSTR2 program. ACTSLl is the program that is actually used as the unit computation. Listings of the two programs are given in Appendix K. The equipment vector for ACTSLl is reproduced below, of which $EN(16) - EN(32)$ are actually used by the CSTR2 routine.

EN VECTOR *********
1.-15. ST C (1.-15. STANDARD FORMAT)
C (4. NUMBER OF REACTORS IN PARALLEL
C (5. TOTAL HEAD LOSS THROUGH ACT, SL. TANKS (FT OF WATER) $\overline{4}$. $5.$ ζ 6. ζ 7. $C \qquad 8 \cdot$ ζ 11. ζ 12. $\zeta = \frac{16}{17}$ ζ 18. ζ 19. ζ 20. ζ ζ ζ \bar{z} . ζ 23. ζ 24. ζ ζ ζ 26. ζ 27. ζ \bar{z} \bar{z} ζ $\frac{29}{20}$. $\overline{30}$. ζ $\frac{31}{2}$. ζ 32. c 33. $34.$ ζ . 35. ζ 36. $C = 37.$
 $C = 44.$ ⁴⁴⁵. NUMBER
FIRST **SECOND** NUMBER OUTPUT OF INPUT STREAMS (= 2.0) INPUT STREAM (= RETURN SLUDGE 'INPUT STREAM' (= FEED 'STREAM)
OF OUTPUT STREAMS (= 1.0)
STREAM NUMBER NOTE ENC16e-32.> ARE USED BY CSTR2 LENGTH OF REACTOR (FEET)
WIDTH OF REACTOR (FEET)
DEPTH OF REACTOR (FEET) LEPTH OF NOT USING A RESIDENCE TIME $=$ $ENC34$) IF USING A RESIDENC LYSIS RATE (KŽ /HR)
BASAL METAB. RATE (K1 /HR] RATE OF SOLUBILIZATION OF DEGRADABLE SOLIDS (K3 STRIPPING RATE OF VOLATILE ORGANICS (K5 /HR) MICROORGANISMS YIELD FACTOR (Y DIMENSIONLESS)
FRACTION OF LYSIS PRODUCTS TO SOLUBLE DEGRADAE
FRACTION OF LYSIS PRODUCTS TO SOLUBLE NONDEGRA RATE OF AIR UNIT GROWTH FRACTION OF N/C RATIO IN P/C RATIO IN RAW FEED LYSIS PRODUCTS TO SOL0BLE LYSIS PRODUCTS TO SOLUBLE ADDITION TO EACH REACTOR RATE MLSS ATE CONSTANT (K4 7HR)
LSS THAT IS NITRIFIERS
RAW FEED THAT IS NITRIFIERS .
FEED FLOW> DISTRIBUTION DEGRADABLE NONDEGRADABLE CARBON <SCF/HRl ACTUAL NUMBER OF TANKS MAXIMUM OF TEN TANKS IN FRACTION OF FEED STREAM FRACTION OF FEED STREAM **ETC** FRACTION OF FEED STREAM NUMBER OF BLOWERS USED
CAPACITY OF EACH BLOWER IN A STEP SERIES TO FIRST TO SECOND TO TENTH SCF/HR AERATION PROCESS TANK TANK TANK (IF ANY) MODEL /HRl CARBON ABSORPTION NUMBER (FOR OXYGEN TRNSFERl NOT PRESENTLY USED NUMBER OF CSTR,S IN SERIES - EITHER AN RTD MODEL OR THE

ACTSLl may be used as the step aeration process model or as a model for the activated sludge process and its various modifications. For the contact stabilization process, ACTSLl will have to be used twice.

The computations carried out by the CSTR2 program may be explained by the following steps:

 $\frac{c}{c}$.

c

- (1) Stream variables are first converted to concentration units,
- (2) The iteration then begins on the effluent concentrations of the reactive components. All inert components are passed through the model. The reaction rates used are:

(a)
\n
$$
\frac{dM}{dt} = (Yk_{4}C - k_{1} - k_{2})M
$$
\nwhere $M = SO(1, 15)$
\n
$$
C = SO(1, 20) + SO(1, 21) = C' + C''
$$
\n
$$
k_{4} = EN(28)
$$
\n
$$
k_{1} = EN(21)
$$
\nand $k_{2} = EN(20)$
\n(b)
\n
$$
\frac{dS'}{dt} = (-k_{3}S' + k_{2} (1-\alpha-\beta))M
$$
\nwhere $S' = SO(1, 16)$
\n
$$
\alpha = EN(25)
$$
\n
$$
\beta = EN(26)
$$
\nand $k_{3} = EN(22)$
\n(c)
\n
$$
\frac{dS''}{dt} = -k_{3}S''M
$$
\ndt
\nwhere $S'' = SO(1, 18)$ as well as $SO(1, 19)$

ر'.

(d)
$$
\frac{dC'}{dt} = - (k_4 M + k_5 q)C'
$$

dt
where $C' = SO(1, 20)$
and $q =$ volume air used/hr/volume of reactor.

(e)
$$
\frac{dC''}{dt} = (k_3S + k_2d - k_4C'')M
$$

dt
where $S = S' + S''$
and $C'' = SO(1, 21)$

 (f) dSNDC $=$ $k_2 \beta M$ dt

where $SNDC = SO(1, 22)$

The concentration of ammonia nitrogen released and the subsequent nitrification to nitrates are also calculated, within the iteration loop. The rate of change of dissolved oxygen is next given by

(g)
$$
\frac{dO_2}{dt} = k_{\text{L}}a (O_{\text{S}} - O_2) - \frac{32}{12} \left\{ (1-Y) k_4 C + k_1 \right\} M
$$

\n
$$
- 4.57 \frac{dN}{dt}
$$

\nwhere $O_2 = SO(1, 7)$
\n
$$
O_{\text{S}} = saturation oxygen concentration
$$

 $k_{\text{L}}a$ = oxygen mass tranfer coefficient and $\frac{dN}{dx}$ = nitrification rate. dt

- (3) A balance is then made on phsophorus,
- (4) The exit pH is next calculated assuming that the carbondioxide concentration is in equilibrium with air.

CHAPTER 6

6. SECONDARY SEDIMENTATION

In the activated sludge process the waste to be treated is mixed with a flocculent sludge containing microorganisms and other organic and inorganic solids, and aerated. The microorganisms are used to feed on the soluble portion of the waste and to convert them into more cell mass which may then be settled out. The level of suspended solids used *in* the activated sludge process range from 2000-4000 mg/1. As the treated water will constitute the plant effluent *in* most cases, it *is* important that the activated sludge *is* removed and the suspended solids concentration *in* the effluentbe reduced to the level of about 20 mg/1. Secondary sedimentation performs such a purpose.

As the settled sludge will be recycled to the activated sludge tanks, and partially "wasted", it *is* equally important that we keep the solids content of the sludge high. The benefits resulting from a "thick" sludge are:

- (a) It maintains a high level of mixed-liquor suspended solids in the activated sludge tanks,
- {b) It reduces pumping costs,
- (c) It improves digester operation on the waste sludge and hence requires smaller digester volumes,

- (d) It increases the economy of sludge dewatering systems such as centrifuges, vacuum filters, etc., and
- (e) It reduces the sludge volume for land or sea disposal.

6.1. Activated Sludge Characteristics

The clarification and thickening propoerties of an activated sludge have been related to its composition and character. A sludge with a relatively high content of inorganic solids (such as clay) will tend to be denser and hence more easily thickened. However, the character of the microorganisms in the sludge can determine whether a sludge will be flocculent and hence have good clarifying properties or whether the sludge will be compact and have good thickening properties.

The microbial population in activated sludge include the bacteria, fungi, protozoa and some rotifiers. The bacteria are the most predominant group, and as they are responsible for stabilizing the organic matter in the waste and in floc formation, they are the most important group as well. Numerous types of bacteria may be found in any sample of an activated sludge, but sometimes one or several genera may predominate, depending on the particular waste being treated.

A brief description of new operating conditions may affect the character of the microorganisms in the sludge

will be presented here. Assuming that oxygen is supplied in sufficient quantities, a high £ood to microorganism ratio will lead to a rapid rate of bacterial metabolism and reproduction. The bacterial cells are highly energetic and will tend to stay as discrete entities. Another reason for the bacterial cells to stay apart, is the high surface area that each cell will enjoy, aiding in the transfer of food and oxygen. This leads to the formation of a dispersed or diffused sludge with poor clarifying and thickening properties.

At a lower food to microorganism ratio, the bacterial cells have less energy and floes being to form when the cells are brought into contact with each other. The sludge also becomes more compact. The clarifying and thickening properties of the sludge improve.

At a very low food to microorganism ratio the bacterial cells are starved and many will die off. With less viable bacterial cells, the sludge will have a poorer flocculating ability, resulting in a poorer effluent. However, the sludge is still compact and will have good thickening characteristics. The above observations are illustrated in Figure 6.1.1.

In the case where the dissolved oxygen concentration in the activated sludge tanks is low, (0.5 mg/1) there is little oxygen penetration into the biological floc, resulting

in lowered bacterial growth. This encourages the growth of filamentous bacteria such as the Sphaerotilus, as they have a high surface area/volume ratio, and are more able to absorb the small amount of oxygen present. These filamentous growths render the sludge less compact and hence displays poor thickening properties. The effect of these filamentous growths on the flocculating and hence clarifying properties of the sludge is not too well know or reported.

Most fungi also have a filamentous structure and their growth tends to be stimulated by carbohydrate wastes, or conditions of low pH and nutrient deficiencies.

Prolonged periods of anaerobiasis in the secondary clarifier could lead to the production of gas, which when entrapped could cause the activated sludge flocs to rise, and escape over the effluent wiers. Denitrification could also contribute to the problem of a rising sludge.

6.2 Correlations for Activated Sludge Settling

From the above discussion, we realize that the twin functions of the secondary settling tanks, namely those of clarification and thickening, are very much related to the operating conditions in the activated sludge tanks. The level of clarification obtained can be described by the fraction of suspended solids escaping in the effluent, and the degree of compaction obtained, by the Sludge Volume Index.
The Sludge Volume Index is the volume in ml. occupied by 1 gm of the sludge after settling for half an hour.

Rex Chainbelt, Incorporated, has developed two correlations for this purpose (47):

$$
XRSS = \frac{382 \text{ (GSS)}^{0.12} \text{ (FM)}^{0.27}}{\text{(MLSS)}^{1.35} \text{ (TA)}^{1.03}}
$$
 (6.2.1)

and SVI = 56.1 + 113 $(1.05)^{\text{T}-20}$ (FM) (6.2.2) where XRSS = fraction of solids escaping in the effluent $GSS = overflow rate (usqpd/sq.fit.)$ $MLSS$ = mixed liquor suspended solids $(mq/1)$

$$
FM = food to microorganism ratio (lbs BOD5)
$$

\n`/lb MLVSS/day)`

MLVSS = mixed liquor volatile suspended solids TA = aeration time (hrs.)

$$
SVI = sludge volume index (ml/gm)
$$

and $T =$ temperature (^oC)

The above correlations are based on 41 observations and the correlation coefficient for the first equation is 0.63 whilst that of the second is 0.78.

The maximum concentration of the underflow solids can be estimated by the value ($10^{\verb|6/SVI|}$ mg/l. Although this represents the compaction obtained after half an hour of settling, it is a good estimate of the final compaction, as further changes in syi are small, especially with mixed

liquor suspended solids of less than 3000 mg/l. It should be noted that there is a wide misconception that the SVI represents the settlins rate *ot* an activated sludge. The half an hour of settling required by the test is not meant to be a measure of the rate, but merely provides a reasonable time for which the sludge to achieve close to the ultimate compaction.

R. v. Villiers (51) performed laboratory scale settling column studies and arrived at the correlation

XRSS =
$$
\frac{556 \text{ (GSS)}^{0.494}}{\text{(MLSS)}^{1.82} \text{(TA)}^{0.439}}
$$
 (6.2.3)

This correlation does not include the effect of the food to microorganism ratio, as does the previous correlation by Rex Chainbelt Inc. However, there is quite a significant difference in the exponents on the variables, between the two studies.

In a preliminary study, P. Leung from the Chemical Engineering Department, McMaster University, using laboratory scale experiments, arrived at the correlation

$$
XRSS = \frac{2560}{(MLSS)^{1.57} (TA)^{0.20}}
$$
 (6.2.4)

This correlation does not include either the food to microorganism ratio or the overflow rate. However, the exponent that he obtained for the effect of the mixed liquor suspended

solids is intermediate between those of the earlier studies, but he found a very much smaller effect of aeration time than did the other two investigators. His results are tabulated in Appendix F.

The disparity between the three correlations could be explained if the nature of the raw wastes entering the plants from which the sludges were taken, were somehow significantly different. It would seem necessary to obtain a correlation based on a sludge actually produced by the plant.

In the absence of a good correlation for the sludge produced in the Burlington plants we have to choose either Equations 6.2.1 or 6.2.3. It should be noted that the above equations are developed from quiescent batch column studies and to apply them to real settling tanks, a correction factor of 1.5 is suggested by most authors. We will use the operating data for the Drury Lane Plant, in 1969, as a comparison:

 $OR = 960$ usgpd/sq.ft.

 $MLSS = 2200 \text{ mg/l}$

 $TA = 9.6$ hrs.

 $F/M = 0.3$ lbs BOD_{F/T} lb MLVSS/day and the obtained XRSS $= 0.090$

Using Equation 6.2.1:

 $XRSS (calculated) = 0.019$ $XRSS$ (corrected) = $0.019 \times 1.5 = 0.029$

Using Equation 6.2.3;

 $XRSS (calculated) = 0.054$ $XRSS$ (corrected) = $0.054 \times 1.5 = 0.081$

The Rex Chainbelt correlation gives too good an effluent, whilst the Villiers' equation gives a result which is closer to the actual operating condition. Hence, Equation 6.2.3. will be selected for use in our program.

6.3 The Secondary Clarifier Model - SECLAR 1

The listing for the program is given in Appendix K. The equipment vector for the model is reproduced below.

The computations carried out by the program can be explained in the following steps:

- (1) The Sludge Volume Index of the activated sludge is first calculated, based on operating conditions in the aeration tanks. Equation 6.2.2. is used.
- (2) The concentration of solids in the effluent, is then calculated from Equation 6.2.3.
- {3) Secondary clarifiers can be operated with either a constant fraction of recycle of the underflow or with a constant depth of sludge blanket. In the second case, the underflow-solids is at its maximum concentration, which can be estimated from the Sludge Volume Index. In the first case, the sludge blanket

č

depth is not constant but could drop when the underflow pumps are set too high or could rise and finally escape over the wiers when the underflow pumps are set too low. When there is no sludge blanket, the concentration of solids in the underflow is below the maximum value, as calculated from the SVI. EN(l6) determines the policy used in the plant, and the underflow is calculated accordingly.

- (4) The scum flow is then determined assuming 100% removal of all incoming scum. The scum solids concentration is determined by EN(l9).
- (5) Dissolved components are split according to the water flows.

CHAPTER 7

7. ANAEROBIC DIGESTION

The "Sewage Treatment Plant Design Manual" (38) adequately describes the purpose of anaerobic digestion as follows:

" ••••• A primary purpose of sludge digestion is to reduce the complex organic matter present in the raw sludge removed by sedimentation processes to a simpler, non-objectionable state. Digestion produces a sludge more amenable to dewatering without nuisance, and it renders the sludge fit for easy disposal by lagooning, dilution or similar means......Digestors also reduce the volmne of sludge and in so doing produce gas which can be utilized for heating purposes."

The anaerobic digestion process has several disadvantages which may discourage their use in a wastewater treatment plant. .Firstly, a high capital outlay is required, amounting to about 25-35% of the total capital cost. Secondly, digester upsets are frequent, thus requiring constant attention, and thirdly, there is always the hazard of a gas explosion.

7.1 Theory

There are two distinct processes occuring in an anaerobic digester - liquifaction and gasification. Liquefaction of the sludge solids is brought about by the extracellular

enzymes secreted by the bacteria, which hydrolyses the complex solids. Cellulose and carbohydrates are converted to simple sugars, alcohols and fatty acids. Proteins are converted to amino acids, while fats and grease are converted to glycerol and fatty acids. The formation of organic acids could result in a depression of the pH, if the alkalinity in the digesting sludge is not sufficient for neutralization. This liquefaction stage of digestion, is also called the acid pnase, for obvious reasons, and the bacteria responsible are usually referred to as acid-formers.

In the gasification process, the end products of liquefaction are further broken down to gaseous end products. This could involve the activity of the same bacteria in the first phase, but with the exception that they are not capable of utilizing their OWn acid end products. This requires the work of a second group of bacteria which metabolize the fatty acids to give methane and carbon dioxide, and convert .amino acids to ammonia. The ammonia released tends to neutralize a portion of the acids remaining and raise the pH. The gasification phase is also sometimes referred to as the methane phase, and the bacteria involved as methane-formers.

The digestion process can be schematically represented as:

Figure 7.1.1. Schematic Representation of The Digestion Process

Since the digester is operated under anaerobic conditions, the bacterial population has to be composed of either the facultative bacteria or the strict anaerobes. The acid-formers are predominantly facultative and many of them originate from the activated sludge fed to the digester. The methane formers are strict anaerobes, and are a small specialized group of bacteria. They are highly sensitive to pH changes and have an optimum pH range of 6.4 to 7.2. When a digester goes "sour" (or acidic) the methane-formers die off, and the gasification phase is seriously retarded.

A properly operating digester should have the liquefaction and gasification processes proceeding at about the same rate. An upset can be produced by an increase in the liquefaction rate, which could be prompted by a sudden increase in the solids fed to the digester.

To provide a reasonable description of the digestion process we need an expression to account for the rate of liquefaction of the organic solids. If a steady-state operation

. 103

is reached, the rate of gasification should equal the rate of liquefaction. We would also need an expression to determine the level of volatile organic acids maintained in the digester. This is necessary since the pH and the BOD of the digesting sludge would depend on the amount of volatile acids present.

^Amodel for the anaerobic digester was first proposed by Fair and Moore (13) in 1932. They followed the digestion process in batch reactors and concluded that the rate of gas production is proportional to the amount of "gas" remaining in the digesting sludge:

$$
\frac{dy}{dt} = k (G-y) \qquad (7.1.1)
$$

where $y =$ amount of gas produced up to time t G = total amount of gas produced

and $k =$ rate constant.

They found k to be 0.168 day⁻¹, at 95[°]F. Since then, other workers have also obtained first order rates. Schulze (36) determined k to be 0.14 day⁻¹ at 92^oF., and Simpson (42) arrived at a k value of 0.3 day^{-1} .

Since it would be useful to our modelling studies to know the rate of liquefaction of· the digesting solids, we have to rearrange Equation 7.1.1. If gas production is a measure of volatile solids destruction, then (G-y) is proportional to the amount of organic solids remaining.

'i.e.,

$$
(G-y) = a S \t\t (7.1.2)
$$

where $S =$ amount of organic solids remaining

and $a =$ proportionality constant.

The proportionality constant is the volume of gas produced per unit mass of organic solids. This has been reported to be approximately 16-18 cu. ft./1b. solids destroyed, for digesting sewage sludge.

Equation 7.1.1 then becomes

$$
\frac{dS}{dt} = -KS \qquad (7.1.3)
$$

Since most reactors are operated continuously or semi-continuously, rather than batchwise, we will assume a stirred tank model for the digester. The defining equation for the digester then is:

$$
S = \frac{SO}{1 + K\bar{t}} \tag{7.1.4}
$$

where $S =$ volatile solids remaining

so = feed volatile solids

and \bar{t} = detention time

or

$$
R = \frac{S}{SO} = \frac{1}{1 + K_{E}^{2}}
$$
 (7.1.5)

where $R =$ fraction of volatile solids remaining. Or

$$
(1 - R)/R = K\bar{t}
$$
 (7.1.6)

The relation here implies that the effluent volatile solids depend on both the influent volatile solids and the detention time, as contrasted to the theory of McCarty (see Chapter 2). This seems to be a more reasonable expression, and the importance of solids loading is supported by the observations of other workers (13, 19, 50).

Rankine (32) has presented data for the fraction of volatile solids destroyed as a function of the detention time *in* a conventional digester. His data is tabulated *in* Appendix G. A plot of $(1 - R)/R$ versus \bar{t} is shown in Figure 7.1.2. $\,$ K is found to be 0.025 day $\,$, and the correlation coefficient is 0.65. The range of solids loading employed as $0.033 - 0.135$ lbs VM/cu.ft./day.

For the high-rate digestion process, the data of Torpey (50), Roy and Sawyer (34) and Estrada (12) are used. Their data *is* also tabulated in Appendix G, and the plot of $(1 - R)/R$ versus t is shown in Figure 7.1.3. The reaction rate constant K is found to be 0.085 $\mathtt{day}^{\mathtt{-1}}$, and the correlation coefficient is 0.58. The range• of solids loading employed here is $0.101 - 0.575$ lbs/cu.ft./day.

Data on the dissolved biodegradable organic carbon in the supernatant is quite sparse. Hence, the equation

Figure 7.1.2. Conventional Di9ester Correlation

,·

 $\overline{}$ '-..1 0

Figure 7.1.3. High-Rate Digester Correlation

proposed by McCarty {see Chapter 2). will be used. He also estimated that the dissolved organic carbon is mainly composed of volatile acids.

The acidity due to the volatile acids is estimated by assuming that acetic acid is the major component, i.e.,

> acidity $(meq/1) =$ soluble organic carbon $(mg/1)$ 24.0

> > (7.1.7)

The ammonia evolved tends to neutralize the volatile acids and raise the pH of the digester. Assuming that the nitrogen to carbon ratio in the feed sludge is approximately 0.23 (based on a sludge composition of $C_5H_7O_2N$), the ammonia nitrogen evolved is:

$$
\Delta N = 0.23 \Delta C \qquad (7.1.8)
$$

where ΔN = ammonia nitrogen evolved

and ΔC = organic carbon destroyed

Hence, the alkalinity due to the ammonia is given by

alkalinity = $\frac{\Delta N}{N}$ = 0.23\AA C $\text{meq}/1$ (7.1.9) 14 14

The effluent pH is then calculated by the module described in Section 9.2.

7.2 The Anaerobic Digestor Model - ANDIG1

A listing of the program is given in Appendix K. The equipment vector for the model is reproduced below:

The computations carried out by the program can be explained briefly by the following steps:

- (1) The fluid detention time is first calculated. EN(24) corrects for the fact that a fraction of the digester volume may be filled up with grit, etc., and hence is not available for digestion.
- (2) The fraction of volatile solids destroyed is calculated from Equation 7.1.5.
- (3) Based on a C:N:P ratio of $1.0:0.23:0.05$, the amounts of nitrogen (as ammonia) and phosphorus released are then estimated.
- (4) The soluble organic carbon of effluent is then obtained from Equation 2.
- (5) The effluent pH is then calculated as discussed in Section 7.1.

(6) If the digester is used as the first stage of a twostage digester, the contents are normally sent to the second stage, with little or no phase separation. This *is* maintained by constant stirring. Phase separation is used in the second stage to separate a a clearer supernate from a concentrated solids underflow. There is no known correlation for the solids concentration of the supernate and hence the phase separations is specified by EN(20), EN(21), and EN(22). Soluble components are split according to the water flows.

CHAPTER 8

8. OTHER UNIT PROCESSES

Other unit processes commonly used in·wastewater treatment include pretreatment, the use of biological filters (trickling filters} and chlorination. The primary purpose of pretreatment is to remove large and abrasive materials from the wastewater, to protect downstream equipment and pumps. It includes screening to remove the larger solid objects and grit removal.

Biological filters have been used as an alternative to the activated sludge process. In this process, the wastewater is passed over a biological growth, supported on a solid medium. The waste is absorbed by the biological growth and excess solids is sloughed off the "filter" and removed by sedimentation. The contact time is short, being of the order of a few minutes and the effluent is generally unsatisfactory.

Chlorination is practiced to prevent the transmission of pathogenic microorganisms to the receiving waters. It is generally applied to the secondary effluent and in cases where no secondary treatment is supplied, to the primary effluent. In the few cases where no treatment is practiced at all, the raw wastewater is chlorinated, prior to discharge.

The thickening of waste sludges is also practiced. This reduces the yolume of sludge to be handled, either for disposal or for further solids processing within the plant. Other methods of dewatering include vacuum filtration, centrifugation and air flotation.

8.1 Pretreatment

Screening devices are usually classified as fine or coarse screens. In the category of fine screens there are perforated plates, wire-meshes and closely spaced bars. The openings are generally 3/16 in. or less. Coarse screens include comminuting devices, bar screens and coarse wiremeshes. The openings may be as large as 3 in.

The choice for the size of the openings is dictated by the size of the largest object which may be allowed to pass. In the majority of cases, a 1 in. opening is quite satisfactory.

The volume of screenings removed is difficult to estimate and depends not only on the screen size but also on the nature of the waste received, and the velocity of flow through the screens. The Rex Chainbelt Company (33) has developed a graph from which the average and maximum amount of screenings may be expected as a function of screen opening. This is reproduced in Figure 8.1.1. The average curve *is* approximately described by:

$$
V = 19 \times 10^{-0.8d}
$$
 (8.1.1)

115

where $V =$ volume of screenings cu.ft./musq and $d =$ screen opening, inches

About 30 lbs. of dry solids may be expected per cu. ft. of screenings.

In grit removal, the object *is* to separate the nonputrescible solids from the waste flow-with the minimum amount of entrained organic matter. This *is* achieved by differential sedimentation, which *is* made possible by the fact that the grit particles have subsiding velocities substantially greater than those of the organic solids. The flow-through velocities are also controlled to maintain the organic solids in suspension, by scouring the settled solids.

Velocity control can be maintained by the use of specially designed wiers, such as the Sutro Wier or the proportional wier. More recent developments in velocitycontrol devices include the use of compressed air to create a spiral current within the grit chamber, More details of the various types of grit chambers in use may be obtained from the "Sewage Treatment Plant Design Manual" (38}.

As is the case with the quantity of screenings, the volume of grit to be expected at a plant cannot be confidently predicted. The quantities of grit received will depend on the area served by the sewers, the type of street and land

surfaces prevalent *in* the district and on the percentage of storm sewers feeding into the plant.

Due to the great variation in quantities of grit received, the model written for the grit chamber will require that we specify the volume of grit to be expected in cu.ft./mig. As a rough guide to use in Ontario, a survey of 28 plants having separate sewers gave an average grit volume of 2.4 cu.ft./mig, while 20 plants having partially combined sewers received on the average 3.3 cu.ft./mig.

8.2 Trickling Filters

 $\mathbf{r}_\mathbf{a}$

The theory proposed for the rapid rate of organic removal in a trickling filter *is* that the waste *is* biosorped and incorporated into the biological growth, supported on the solid media. As the wastewater passes down through the filter, progressive removal *is* achieved until the clear effluent emerges. Howland {21) and Schulze {35) have shown that at low concentrations, the rate of removal is of the first order with respect to the concentration of soluble organics remaining:

i.e.,
$$
\frac{C}{C_o} = e^{-k}1^t
$$
 (8.2.1)

where $C = effluent concentration of soluble organics$ = influent concentration of soluble organics c_{α}

 k_1 = reaction rate constant and $t =$ mean contact time.

Schulze (35) and Howland (21) have also shown that the mean contact time in a trickling filter is directly proportional to the depth and inversely proportional to the hydraulic loading to a power, n, which depends on the solid media used.

i.e.,
$$
t = k_2 D/Q^n
$$
 (8.2.2)
\nwhere $t = \text{mean contact time (min.)}$
\n $D = \text{depth of filter (ft.)}$
\n $Q = \text{hydraulic loading (usgpm/sq.fit.)}$
\n $k_2 = \text{proportionality constant}$
\nand $n = \text{constant.}$

Values of n range from 0.55 for spheres (55) to 0.66 for a screen filter (35). The n value for an increasingly popular media, the synthetic Dowpac Plastic Hedia, is 0.5.

It has also been shown that the reaction rate could decrease with depth. This could arise if the more easily removable portions of the waste are removed at the top of the filter, leaving a less readily degradable matter for the lower portion of the trickling filter. Another possibility exists whereby the microbial population in the filter is stratified with the more efficint bacteria and fungi at the top and progressing downwards to the predator microorganisms.

A general relationship can be developed by combining Equations 8.2.1 and 8.2.2, and with a modification for the effect of depth on the reaction rate:

$$
\frac{C}{C_{\rm o}} = e^{-kD^{\rm m}/Q^{\rm n}}
$$
 (8.2.3)

where $k =$ rate coefficient or treatability factor $m = constant$.

In the case where there is no effect of the depth on the reaction rate, $m = 1$. This has been found to be the case for domestic sewage by several investigators. Schulze (35) determined k to be 0.020 m to be 1.0 and n to be 0.66 for a screen treating settled sewage. McDermott (55) concurs similarly, but his rate coefficient is slightly lower, at 0.018. Germain (56) found m to be 1.0, n to be 0.48 and k to be much higher at 0.088, for the case of the Dowpac Filter Media. However, his data is based on only a few points.

8.3 Chlorination

The purpose of the model will be merely to calculate the chlorine requirements *in* a wastewater treatment plant. Since the effluent will not be directly used as a drinking water supply, but is discharged to a water-course, the criteria for chlorination is mt so strict.

The chlorine dosage required depends on the degree of treatment the effluent has gone through. Raw sewage will require a higher chlorine dosage than the eftluent from a secondary clarifier. The "Sewage Treatment Plant Design Manual" (38) gives the range of chlorine dosages commonly used, and this is reproduced in Table 8.3.1.

Table 8.3.1. Range of Chlorine Dosages Required for Disinfection.

8.4 Sludge Thickening and Dewatering

Sludge thickening performance is not very well correlated. Hence, only a simple mass balance model will

be used. This is a slight modification of the SEPAOl model available from the GEMCS library of routines, The fractions of water (and water soluble components) and solids to the supernatant *is* supplied as inputs to the program.

Other dewatering devices are also approximately described by the same model.

CHAPTER 9

9. MODULES FOR HANDLING STREAM VARIABLES

There are certain calculations which occur frequently in some or most of the simulation models used in this study. As mentioned in Section 3, the stream elements 13-22 form the basic components of a waste flow. The simulation models directly use these variables in their internal calculations. However, *in* wastewater treatment practice, the more commonly used variable are those of stream elements 8-12, which include the BOD, ss, VSS, TOC and DOC. Hence it *is* necessary to convert from the "working" variables to the commonly reported variables.

Another area where calculations are frequently needed is in the pH - alkalinity relationship. Also in the activated sludge model, it *is* more convenient to use concentration variables (i.e., mg/1) than *it* is to use flow variables such as lbs./hr. Hence a module *is* also available to convert component flows into component concentrations.

9.1 Inter-relationship of Stream Variables

A series of experiments were performed to determine the relationships between the stream elements 8-12 and those of 13-22. Section 3.2. gives the relevant defining equations.

The Total Organic Carbon (TOC), which includes both the particulate and dissolyed organic carbon forms, was first determined. To ensure that the injection needle would not "filter" out any solids, the sample was subjected to ultra-frequency sonication for five minutes.

The Total Five-day Biochemical Demand (TBOD₅), which includes the BOD₅ of both the particulate and soluble components, was then determined using the original sample.

The sample was then filtered on two Whatman's #40 filter papers (ashless). One filter paper was then ashed in a muffle furnace at 600°C and the other was dried at 103° C. This will yield the volatile suspended solids (VSS) of the sample. The filtrate was then used to determine the Dissolved Organic Carbon (SOC) and the Dissolved Five-day Biochemical Oxygen Demand (DBOD₅). All the above mentioned analyses were performed according to "Standard Methods" (48).

The results are summarized in Appendix G. The average values obtained for the conversion factors are:

(a) FAC1 = $^{DBOD}5 = 1.91$ soc

with a standard deviation of 0.17 (b) FAC2 = BOD_5 (particulate) = $TBOD_5 - DBOD_5 = 1.24$ OC (particulate) TOC - SOC

with a standard deviation of 0.12

(c) FAC3 =
$$
\frac{VSS}{OC (particulate)} = \frac{VSS}{TOC - SOC}
$$
 = 2.09
with a standard deviation of 0.11

Eckenfelder (8) derived the value of 1.85 for FACl. Smith (46) used the value 1.87 for FACl. These two values agreed with our data. Smith reported a value of 0.80 for FAC2, whilst our value of 1.24 is much higher. The value of 2.13 is used by Smith for FAC3, which is close to our value of 2.09.

9.2 $pH - Alkalinity$ Relationships

The major contributors to the alkalinity of municipal wastewaters are the species in the carbonate buffer system and ammonia. To a lesser extent, the phosphates and borates could contribute to the alkalinity.

The equilibrium conditions existing in solution can be described by (49):

For the carbonate system

$$
H_2CO_3^* \rightleftharpoons H^+ + HCO_3^- \tag{9.2.1}
$$

 $HCO₃^T \rightleftharpoons H⁺ + CO₃$ $(9.2.2)$

where $[L_{2}CO_{3}^{*}] = [CO_{2}$ dissolved] + $[L_{2}CO_{3}]$ (9.2.3)

For the ammonium system

$$
\text{NH}_4^+ \rightleftharpoons \text{NH}_3 + \text{H}^+ \tag{9.2.4}
$$

$$
H_2O \rightleftharpoons H^+ + OH^-\tag{9.2.5}
$$

The equilibrium constants for the above equations

are:

$$
k_1 = \frac{\begin{bmatrix} H^+ \end{bmatrix} \begin{bmatrix} HCO_3 \end{bmatrix}}{\begin{bmatrix} H_2CO_3 \end{bmatrix}} = 10^{-6.3}
$$
 (9.2.6)

$$
k_2 = \frac{\begin{bmatrix} H^+ \end{bmatrix} \begin{bmatrix} C O_3 \end{bmatrix}}{\begin{bmatrix} HCO_3 \end{bmatrix}} = 10^{-10.3}
$$
 (9.2.7)

$$
k_{n} = \frac{\begin{bmatrix} H^{+} \end{bmatrix} \begin{bmatrix} NH_{3} \end{bmatrix}}{\begin{bmatrix} NH_{4}^{+} \end{bmatrix}} = 10^{-9.3}
$$
 (9.2.8)

and
$$
k_w = [H^+] [OH^-] = 10^{-14.0}
$$
 (9.2.9)

The definition for alkalinity can then be expressed as

$$
\begin{bmatrix} \text{ALK} \end{bmatrix} = \begin{bmatrix} \text{HW}_3^- \\ \text{H}^+ \end{bmatrix} + 2 \begin{bmatrix} \text{CO}_3^+ \end{bmatrix} + \begin{bmatrix} \text{NH}_3 \end{bmatrix} + \begin{bmatrix} \text{OH}^- \end{bmatrix}
$$
\nwhere
$$
\begin{bmatrix} \text{ALK} \\ \text{ALK} \end{bmatrix} = \text{alkalinity equivalents/litre}
$$
\n
$$
\begin{bmatrix} \text{HCO}_3 \\ \text{CO}_3 \end{bmatrix} = \text{bicarbonate ion concentration moles/litre}
$$
\n
$$
\begin{bmatrix} \text{OH} \\ \text{NH}_3 \end{bmatrix} = \text{free ammonia concentration moles/litre}
$$
\n
$$
\begin{bmatrix} \text{OH}^+ \\ \text{OH}^- \end{bmatrix} = \text{hydroxyl ion concentration moles/litre}
$$
\nand
$$
\begin{bmatrix} \text{H}^+ \end{bmatrix} = \text{hydrogen ion concentration moles/litre}
$$

Due to the conservation of mass, we note that

$$
\begin{bmatrix} H_2CO_3^* \end{bmatrix} + \begin{bmatrix} HCO_3^- \end{bmatrix} + \begin{bmatrix} CO_3^- \end{bmatrix} = \text{constant} = \begin{bmatrix} C_T \end{bmatrix} \quad (9.2.11)
$$

and
$$
\begin{bmatrix} NH_3 \end{bmatrix} + \begin{bmatrix} NH_4^+ \end{bmatrix} = \text{constant} = \begin{bmatrix} C_N \end{bmatrix} \quad (9.2.12)
$$

Hence, from Equations 9.2.6 and 9.2.7, as well as from 9.2.11

$$
\begin{bmatrix} C_T \end{bmatrix} = \begin{Bmatrix} \left[\frac{\mu^+}{2} \right] & \mu^+ \mu^+ \mu^- \mu^+ \end{Bmatrix} \begin{bmatrix} HCO_3^- \end{bmatrix}
$$
\n
$$
\text{or } \begin{bmatrix} HCO_3^- \end{bmatrix} = \begin{bmatrix} C_T \end{bmatrix} \begin{Bmatrix} \frac{1}{\mu^+} \\ \frac{\mu^-}{2} \\ \mu^- \mu^- \mu^- \mu^- \mu^+ \end{Bmatrix}
$$
\n
$$
(9.2.13)
$$

Similarly

$$
\begin{bmatrix} \text{CO}_3^{\equiv} \end{bmatrix} = \begin{bmatrix} \text{C}_{\text{T}} \end{bmatrix} \left\{ \frac{\text{k}_2}{\text{k}^+} \right\} \left\{ \frac{1}{\text{k}^+ + 1 + \frac{\text{k}_2}{\text{k}^+ + 1 + \frac{\text{k}_2}{
$$

Therefore

$$
\begin{bmatrix} \text{ALK} \end{bmatrix} = \begin{bmatrix} C_T \end{bmatrix} \begin{Bmatrix} 1 + \frac{2k_2}{\mu^+} \end{Bmatrix} \begin{Bmatrix} \frac{1}{\mu^+ + 1 + k_2} \\ k_1 \end{Bmatrix}
$$

$$
+\left[C_{\rm N}\right] \left\{\frac{1}{1+\left[\rm H^{\dagger}\right]} \left\} + \left[\rm H^{\dagger}\right] - \left[\rm H^{\dagger}\right] \right\} \qquad (9.2.16)
$$

This is the defining equation used to relate the four variables [ALK], [C_T], [CN] and pH. Given three of the variables, the fourth may be calculated. Note also that *if* streams are mixed, and assuming a closed system, there must be conservation of $\begin{bmatrix} ALK \\ C_T \end{bmatrix}$ and $\begin{bmatrix} C_N \end{bmatrix}$.

On expanding Equation 9.2.16, we obtain a polynomial of the fifth order in H^+ : $[-4]$ $[-4]$ $[-4]$ $[-4]$ $[-4]$ $[-4]$ $[-4]$ $[-4]$ $[-4]$ $[-4]$

$$
\begin{aligned}\n\begin{bmatrix}\nF^T\n\end{bmatrix}^3 + \begin{bmatrix}\nF^T\n\end{bmatrix}^2 \left\{\n\begin{bmatrix}\n\Delta L K\n\end{bmatrix} + k_1 + k_N\n\end{bmatrix}\n+ \begin{bmatrix}\nF^T\n\end{bmatrix}^3 \left\{\n\begin{bmatrix}\n\Delta L K\n\end{bmatrix} (k_1 + k_N) + k_1 k_N + k_1 k_2 - k_w - k_1 \begin{bmatrix} C_T \end{bmatrix} - k_N \begin{bmatrix} C_N \end{bmatrix}\n\right\} \\
+ \begin{bmatrix}\nF^T\n\end{bmatrix}^2 \left\{\n\begin{bmatrix}\nk_1 \Delta L K\n\end{bmatrix} (k_2 + k_N) + k_1 k_2 k_N - k_N k_w - k_1 k_N \begin{bmatrix} C_T \end{bmatrix}\n\right. \\
- k_1 k_w - 2k_1 k_2 \begin{bmatrix} C_T \end{bmatrix} - k_1 k_N - 2k_1 k_2 k_N \begin{bmatrix} C_T \end{bmatrix} \\
- k_1 k_2 k_w - k_1 k_2 k_N \begin{bmatrix} C_N \end{bmatrix}\n\right\} \\
- k_1 k_2 k_w - k_1 k_2 k_N = 0\n\end{aligned}
$$
\n(9.2.17)

We could simplify the above equation by making some order of magnitude simplifications. Roughly, $\left[\Delta L K\right] \approx \left[C_{\text{N}}\right] \approx \left[C_{\text{T}}\right] \approx 10^{-3}$.

This means that in the pH range of 5-9, the terms $\begin{bmatrix} \text{OH}^{\text{-}} \end{bmatrix} - \begin{bmatrix} \text{H}^{\text{+}} \end{bmatrix}$ \ll \lceil ALK \rceil in Equation 9.2.16. Hence we can drop out these two terms resulting *in:*

$$
\begin{bmatrix} C_T \end{bmatrix} \begin{bmatrix} 1 + \frac{2k_2}{\mu^+} \end{bmatrix} \begin{bmatrix} \frac{1}{\mu^+} + 1 + \frac{k_2}{\mu^+} \end{bmatrix} = \begin{bmatrix} \lambda LK \end{bmatrix} - \begin{bmatrix} C_N \end{bmatrix} \begin{bmatrix} \frac{1}{\mu^+} \end{bmatrix}
$$

On expansion, a third order polynomial in $[H^+]$ results:

$$
\begin{bmatrix} H^+ \end{bmatrix}^3 \begin{bmatrix} ALK \end{bmatrix} + \begin{bmatrix} H^+ \end{bmatrix}^2 \begin{bmatrix} k_N \end{bmatrix} \begin{bmatrix} ALK \end{bmatrix} - \begin{bmatrix} C_N \end{bmatrix} + k_1 \begin{bmatrix} \begin{bmatrix} ALK \end{bmatrix} - \begin{bmatrix} C_T \end{bmatrix} \end{bmatrix}
$$

$$
+ \begin{bmatrix} H^+ \end{bmatrix} \begin{bmatrix} k_1 k_N \begin{bmatrix} \begin{bmatrix} ALK \end{bmatrix} - \begin{bmatrix} C_N \end{bmatrix} - \begin{bmatrix} C_T \end{bmatrix} \end{bmatrix} + k_1 k_2 \begin{bmatrix} \begin{bmatrix} ALK \end{bmatrix} - 2 \begin{bmatrix} C_T \end{bmatrix} \end{bmatrix} \end{bmatrix}
$$

$$
+ k_1 k_2 k_N \begin{bmatrix} \begin{bmatrix} ALK \end{bmatrix} - \begin{bmatrix} C_N \end{bmatrix} - 2 \begin{bmatrix} C_T \end{bmatrix} \end{bmatrix} = 0
$$

Equation 9.2.19 has been found for the several cases tried to *give* only one real root and two imaginary roots. This *is* very fortunate and helps simplify the solution of the cubic equation, for which a simple interval bisection technique *is* used. The bisection subroutine uses the geometric mean of the left and right estimates of the solution, in its convergence.

9.3 Inter~conversion Between Mass flow and Concentration

Although the units used for the stream variables are for mass flow, it is frequently useful to convert them into concentration units. The model for the aerobic biological processes has its rate equations all in terms of component concentrations. The summary report. generated has all the values expressed *in* concentrations, which *is* more meaningful than mass flows.

Due to the analytical procedures used *in* wastewater treatment, soluble components are usually expressed as milligrams per litre of water, while solid components are expressed *in* milligrams per litre of sample. In the case where the concentrations involved are less than 1000 mg/1 {0.1%) the difference between the two *is* negligible. However, *since* sludges of up to 80,000 mg/1 (8%) are handled in our system, the distinction should be made.

For soluble components we then use

$$
C_{\underline{i}} = \frac{F_{\underline{i}}}{W} 10^{-6}
$$
 (0.3.1)

where C_i = concentration of soluble component *i* (mg/1) *Fi* = mass flow of soluble component *i* (lbs/hr) and $W =$ mass flow of water (lbs/hr)

For particulate components we use

$$
C_{\underline{i}} = \frac{F_{\underline{i}}}{Q} \times 10^{-5}
$$
 (9.3.2)

where C_i = concentration of particulate component i (mg/1) F_i = mass flow of particulate component i (lbs/hr) and $Q =$ total volumetric flow of stream (gal/hr)

CHAPTER 10

10. WASTEWATER TREATMENT COSTS

Besides the technical aspects of wastewater treatment, we have to concern ourselves with the costs that such a program would entail. In the expectation of increased activity in the area of pollution abatment, it is becoming more essential that we have accurate cost estimates to base our designs on. A large part of current practice has been to use "rules of thumb" which have fairly large safety factors built into them. An optimal design for a treatment plant, subject to the requirements of the regulatory bodies, can be arrived at by combining the technical section of our simulation with the corresponding cost estimates.

For each unit process in wastewater treatment, capital costs as well as operating and maintenance costs are available as a function of the most prominent capacity.factor. For example, the capital cost of the activated sludge tanks is derived as a function of its volume. Most of the correlations are based on the same idea as the six-tenths capacity factor used in Chemical Engineering.

The bulk of the correlations come from the work of Russell and Swanson as reported in Smith's paper in 1969 (46), and from Eckenfelder and Barnard (2}. Industrial waste treatment costs are also given in the latter report.
The capital costs are all updated to 1969 costs by the use of the Engineering News Record Index of 1120. This index, being based on changes in the price of steel, cement, lumber and common labour, is most appropriate for wastewater treatment process equipment. The capital cost correlations used are summarized in Table 10 .1.

The total cost of the unit processes (installed) is then the sum of the separate costs. It is assumed that the duplication of a unit merely doubles the cost for the unit. This does not take into account that there may be savings due to shared pipings or channels or other factors.

The Physical Plant Cost can then be found by adding the cost of the control house and site improvements on to the total unit processes cost. The Engineering Costs added to the.Physical Plant Cost will give the Direct Plant Cost. The Engineering Cost factor is obtained as a fraction of the Physical Plant Cost. As given in Smith (46)

> $, 0.146$ CENG = 8.0 ($1,000,000$)^{0.140} (10.1) Physical Plant Cost

where $CENG =$ Engineering Cost factor.

The Fixed Capital Cost can then be computed by adding the Contractor's fee (10% of Direct Plant Cost} and the contingency costs (15% of Direct Plant Cost} to the Direct Plant Cost. These figures are suggested by Smith (46) and

Table 10.1. Capital Cost Correlations

1-' *w* N

FIXED CAPITAL ESTIMATE

 $\gamma \to \gamma \gamma$.

TREATMENT PLANT - BURLINGTON. CAPITAL COST IN 1962 **DRURY SEWAGE** ANE $\frac{2\pi}{4\pi}$ $\frac{1}{\sqrt{2}}$

Table 10.2. Capital Cost for Drury Lane Plant

are about average in the Chemical Process Industry. The derivation of the Fixed Capital Cost is summarized in Table 10.2, for the case of the Drury Lane, Burlington Water Pollution Control Plant. The actual plant cost in 1962 was \$676,033.78, while the estimated cost was \$671,595.03. The difference is less than 1%.

The operating and maintenance costs of the unit processes are normally reported in the literature together as one cost. Smith (46} and Eckenfelder and Barnard (2) have reported them in terms of annual costs (utilities excluded) . This necessitates the use of a "Labour Index" to bring these . costs up to date. However, we can avoid this by converting the annual costs to man-hours required per year, by dividing by the labour cost $\frac{2}{\pi}$ /man-hours for the year in which the correlations were obtained. This was calculated from the average wage of \$110/week or approximately \$3.00/man-hour in 1966 (26). The annual cost is then computed by multiplying the man-hours required by the current labour cost. Table 10.3 summarizes the operating and maintenance man-hours used in the program.

Again we have assumed that the duplication of a unit will double the number of man-hours required to operate and maintain the units.

The sum of the Operating and Maintenance Costs for the unit processes will give the total labour cost. This,

together with supervision, utilities, sludge haulage, chemical and plant supplies, will give the Direct Operating and Maintenance Cost. Supervision_is assumed to be 10% of the Total Labour Cost. Chemical and plant supplies are assumed to be 6% and 5% of the Total Labour Cost, repectively. These estimates are obtained from the Ontario Water Resources Commission annual reports on their plant operations. The estimates of power consumption provided by Michel (26) can be correlated to give the following annual power consumption *P* in kwh:

> For Primary Plant $P = 101000 \Omega^{0.65}$ For Trickling Filter Plant $P = 16200000^{0.65}$

For Activated Sludge Plant P = 378000 $Q^{0.65}$

where Q is the plant flow in migd.

Sludge haulage fees in Ontario vary and from the annual reports of the Ontario Water Resources Commission, an approximate correlation is obtained as:

Sludge haulage costs = $8000 (Q)^{0.5}$ \$/yr. where $Q =$ sludge volume hauled, mig/yr.

Indirect costs are generally not significant, the most important of which is for laboratory analyses. The bigger plants usually conduct their own analyses, while the smaller ones tend to send them away to be done. The Net Operating and Maintenance cost will then be the sum of the Direct and Indirect costs.

To obtain the true annual cost, we have to take into account the cost of the Fixed Capital Investment. In the case of owned capital, we have to include depreciation and opportunity costs. The capital recovery factor to use is:

$$
CRF = \frac{S (1 + S)^{n}}{(1 + S)^{n} - 1}
$$
 (10.2)

where $CRF = Capital Recovery Factor$

S = interest rate that owned capital can obtain elsewhere

and $n =$ expected life of plant.

The cost of the Fixed Capital Investment to the municipality of company, annually, is then

 $R = CRF (L_0 - I_c)$

where $R =$ amortization

 I_{Ω} = Fixed Capital Investment

and I_s = scrap value of project at the end of n years.

In the case of borrowed capital, which is the usual case with most municipalities, we have to add debt retirement plus accrued interest to the Net Operating and Maintenance Cost to obtain the true Annual Cost. The factor can be calculated from the same equation, but the interest rate on the loan s and the debt reitrement period n may be different.

The derivation of Total Annual Cost is shown in Table 10.4, again, for the case of the Drury Lane, Burlington Water Pollution

Control Plant. The actual operating cost in 1969 was \$103,520.52, while the projected cost was \$109,116.34, which is about 5% higher.

Table 10.3. Annual Operating and Maintenance Man-Hour Requirements

I-' *w* 00

Table 10.4. Operating Costs for Drury Lane Plant

139

 \mathbb{R}^2

CHAPTER 11

11. INTER-ACTIVE VERSION

An inter-active version of the simulation was written to enable the user to set up a dialogue with the computer. This can be used in the following ways:

- (1) As a teaching tool to train operators and other students,
- (2) To enable plant personnel to predict plant performance due to changes in feed flows and/or operating conditions, and
- (3) As a quick and accurate aid to plant designers.

An important feature of the inter-active version is that it requires only a very minimum knowledge of the GEMCS system for its use. This is achieved through a series of questions and answers posed by the computer. Of course, a user with a good knowledge of GEMCS, can manipulate and use the inter-active version with greater flexibility.

The inter-active version consists of four subroutines and a slightly modified version of the main program of GEMCS. The four subroutines are FLWCHT, CASET, ICLOAD and ENSET. Their functions will be described below.

FLWCHT enables the user to set up his own plant layout, together with the physical dimensions of the various units. It has an enlarged process flow diagram covering most of the common unit processes encountered in wastewater treatment

plants. This is shown in Figure 11.1. Ths units are selected or by-passed by the use of the mixer-splitter module. The majority of the data set is read in by DLOADl, to minimize the use of the teletype to input such information. Hodule numbers, module types, input and output stream numbers and s o on are predetermined for all the units in the process flow diagram and hence can be read in by DLOADl. The calculation order and the physical dimensions of the various units are determined by a dialogue with the user.

CASET is used to generate a random case study. It generates a random feed by multiplying the average value of each component flow by a random factor ranging between 0.5 and 2.0. It also generates a failure in one of the plant units. The purpose of the subroutine is to teach operators what to do in the event that such a flow or plant failure occurs, as well as to locate the cause of such a failure.

ICLOAD is the analytical laboratory of the simulation. All information regarding stream flows and component concentrations are supplied through ICLOAD. A charge is levied for each analysis performed, the object of the game being to locate and correct a plant failure with the minimum number of analyses. A surcharge is also placed on any unsatisfactory effluent. Five cents is levied on each pound of suspended solids and Biochemical Oxygen Demand, above the regulatory level of 20 mg/1 for both. This provides the incentive to correct a plant failure with the minimum delay.

ENSET enables the user to change any operating conditions by changing the appropriate values of the EN vectors. The process flow diagram however, cannot be changed. This must be done through FLWCHT.

In addition to the above four subroutines, slight changes are made to the other modules, primarily to suppress excessive printing.

Figure 11.1. Process Flow Diagram for the Interactive Version

Key to Figure 11.1. 144

CHAPTER 12

12. TEST RUNS

The two plants selected for simulation are the Drury Lane and the Skyway Sewage Treatment plants. The former is a conventional activated sludge process, whilst the latter employs the extended aeration modification. In addition to testing the various models developed, the two simulations should show that the same kinetics can be used for the activated sludge process, regardless of the process modification. •

The base cases used will be the average flow and operating conditions for 1969, obtained from plant data and also from the Ontario Water Resources Commission's Annual Summaries. The 1969 data were chosen, as the operating and flow conditions were fairly constant throughout the year. Due to the diurnal variation in the feed flows, the steadystate models can, at best, predict the average performance of the various units. It is for this reason that no attempt was made to collect base case data on any particular day, as this would require at least a twenty four hour sampling schedule, for a week.

12.1 Simulation of the Drury Lane, Burlington Sewage Treatment Plant

The process flow diagram for the plant is shown in Figure 12.1.1, and the design data is summarized in Appendix I.

Key to Figure 12.1.1.

It was originally designed for a flow of 2.5 migd. However, due to the expansion of the Skyway plant, part of the flow was diverted to the latter plant, and the actual flow received was 1.6 migd, in 1969. The average BOD and TSS for the raw sewage is 215 mg/1 and 299 mg/1, respectively.

The pretreatment section consists of 1-inch bar screens and grit channels with a detention time of approximately one minutes. Primary treatment consists of two tanks with dimensions 29.3 ft. x 18 ft. x 12.25 ft. Secondary treatment is provided by two triple-pass activated sludge tanks, each with a total length of 321.5 ft., width of 18 ft. and depth of 10.7 ft. Total air blower capacity is 3000 scfm. Two circular secondary clarifiers are employed, each with a radius of 25 ft. and depth of 10.6 ft. A two-stage digestion system is used to handle the waste sludge generated. The first stage consists of two 40 ft. diameter tanks with a depth of 20 ft., and the second stage consists of one 40 ft. diameter tank with a depth of 18 ft. A summary of the operating conditions and the parameters used for the simulation is shown in Table $12.1.1.$

The data set used for the simulation is shown in Appendix I. The computer printout for the case is also included *in* Appendix I. A summary of the results from the simulation, together with the plant data is shown in Table 12.1.2.

The removal of Total Suspended Solids by the primary clarifier was predicted to be 49% while the observed removal

was 48%. The BOD removal predicted was 30% compared to the actual removal of 33%. Both removals were predicted very well by the primary clarifier model.

The Total Suspended Solids in the final effluent is identical with the value observed, which was 20 mg/1. The BOD of the final effluent was predicted to be 22 mg/1, while the observed value was lower at 15 mg/1. The agreement is quite good.

The predicted Total Suspended Solids of the return activated sludge was 11,700 mg/1 with a volatile content of 57%. The observed values were 12,000 mg/1 and 60% respectively.

The simulation predicted that a greater volume of sludge should be sent to the digester than was actually observed. A raw sludge flow of 7600 igpd was calculated, while the reported flow was much lower at 5300 igpd. This is reasonable since a considerable amount of solids is lost during a rain, but which is not accounted for. The figures imply that about 30% of the solids which should go to the digester was lost over the wiers of the secondary clarifiers, either during a storm or due to a plant upset.

The simulation predicted a digested sludge flow of 3000 igpd at a Total Suspended content of 5.8% as compared to the volumes of 2300 igpd hauled away for disposal. The anaerobic digester model predicted a much higher volatile solids

destruction than was observed. This *is* reflected in the lower' volatile solids concentration predicted for the digested sludge.

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RAW SEWAGE:

average $flow = 1.6$ migd average TSS $= 299 \text{ mq/1}$ average VSS $= 195 \text{ mg/l}$ percent of voltile solids settleable = (77%) percent of inorganic solids settleable = (46%) average BOD $= 215 \text{ mg/l}$ average DOC $=$ (61) $mq/1$

PRIMARY SETTLING TANKS: (using nomenclature of Chapter 4)

 W_T = \emptyset exp $(-kt^{\alpha}/H^{\beta})$ where $k = 0.23$ α $= 0.5$ $= 0.25$ ß and *¢* = 0.85

average underflow solids concentration = $60,000$ mg/l ACTIVATED SLUDGE TANKS: (using nomenclature of Chapter 5)

$$
k_1 = 0.0028 \text{ hr}^{-1}
$$

\n
$$
k_2 = 0.0014 \text{ hr}^{-1}
$$

\n
$$
k_3 = 0.000002 \text{ hr}^{-1}
$$

\n
$$
k_4 = 0.0010 \text{ hr}^{-1}
$$

\n
$$
k_5 = 2.2 \text{ hr}^{-1}/(\text{cu.ft.air}/\text{cu.ft.reactor})
$$

 $y = 0.54$

 $MLSS = 2200$ mg/1.

RTD model: 3 CSTR's in series.

SECONDARY SETTLING TANKS: (using nomenclature of Chapter 6) 556 (GSS 0.494) \times 1.5 XRSS =

 $(MLSS$ 1.82) $(TM$ ^{1.5})

 $SVI = 56.1 + 113 (FM) (1.05T-20)$

fraction of time return sludge pumps used = 0.3 ANAEROBIC DIGESTER: (using nomenclature of Chapter 7)

1st stage at high-rate, $k = 0.082$

2nd stage at conventional rate, $k = 0.026$ day⁻¹

fraction of flow to supernate = 0.6

Figure 12.1.1. Operating Conditions and Parameters Used

In the Simulation of the Drury Lane Plant

Table 12.1.2. Simulation of the Drury Lane, Burlington, Water Pollution Control Plant

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12.2 Simulation of the Skyway, Burlington, Sewage Treatment '

· Plant

The process flow diagram for the plant is shown in Figure 12.2.1, and the design data is summarized in Appendix J. The average flow *in* 1969 was 3.3 migd, but due to an expansion completed in 1970, it *is* now receiving about 6. 0 migd.

Pretreatment consists of a $2\frac{1}{2}$ inch coarse screen followed by a 1 inch screen, both mechanically cleaned. Grit removal is achieved by an aerated grit chamber with a detention time of approximately 20 minutes. There are no primary tanks in the Skyway Plant. The activated sludge process employed is that of the extended aeration modification, and consists of six single-pass tanks with dimensions of 270 ft. x 27 ft. x 15 ft. The total blower capacity is 18000 scfm. Secondary clarification is provided by four settling tanks, 60 ft. square and 12 ft. deep. Waste sludge is thickened in a circular tank, 20 ft. in diameter and 9 ft. deep. A summary of the operating conditions and the parameters used in the simulation is shown in Table 12.2.1.

The data set for the simulation *is* shown in Appendix J, together with the computer printout for the case. A summary of the results from the simulation together with the plant data is shown in Table 12.2.2.

Key to Figure 12.2.1.

The simulation predicted a slightly better final effluent than was observed. The BOD and TSS predicted were 8 mg/1 and 10 mg/1 respectively, while the observed values were $12 ~mg/1$ and $12 ~mg/1$ respectively.

From plant data, the average suspended solids concentration *in* the return sludge was 10,000 mg/1, while the simulation predicted a value of 9,500 mg/1. The agreement *is* very good. However, the average volatile content of the sludge observed was 60% while the simulation gave a value of 42%.

A greater discrepancy occured here than was with the case of the Drury Lane Plant, *in* the volume of sludge hauled away. The simulation predicted that a volume of 22,000 igpd at a solids content of 2.1% should be hauled away, but actual plant figures indicated that only 4000 igpd at 2.0% solids were hauled away. Although upsets are frequent *in* a plant this size, the almost five-fold difference *in* the thickened sludge for disposal *is* unexpected. From a rough balance on the solids over the entire plant, the volume of sludge produced should be approximately

3.3 x 10^6 x $(185 - 12)$.

20,000

 $= 28,000$ igpd, at 20% solids

This figure is in better agreement with the predicted value. Continuous sampling of the final effluent will probably show up the difference.

 $\frac{y}{B}$

average flow $=$ 3.3 migd average TSS = $185 \text{ mg}/1$ average $VSS = 85$ mg/1 average BOD = $155 \text{ mg}/1$ average DOC = (62) mg/l

ACTIVATED SLUDGE Tanks: (using nomenclature of Chapter 5)

 k_1 = 0.0028 hr⁻¹ k_2 = 0.0014 hr⁻¹ k_3 = 0.000002 hr⁻¹ $k_A = 0.0010 \text{ hr}^{-1}$ k_{5} = 2.2 hr⁻¹ $Y = 0.54$

 $MLSS = 4000 \text{ mq/1}$

RTD model: 3 CSTR's in series

SECONDARY SETTLING DATA: (using nomenclature of Chapter 6)

\n
$$
\text{XRSS} = \frac{556 \, (\text{GSS} \, 0.494) \times 1.5}{(\text{MLSS} \, 1.82) \, (\text{TM} \, 1.5)}
$$
\n

\n\n $\text{SVI} = 56.1 + 113 \, (\text{FM}) \, (1.05 \, \text{T}^{-20})$ \n

Fraction of time return sludge pumps used = 0.4 THICKENER: Fraction of liquid to overflow= 0.6

Fraction of solids to overflow = 0.1

Table 12.2.1. Operating conditions and Parameters Used in the Simulation of the Skyway Plant

Table 12.2.2. Simulation of the Skyway, Burlington, Water Pollution Control Plant

CHAPTER 13

13. DISCUSSION OF RESULTS

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13.1 Discussion of Experimental Data

13.1.1 Primary Settling Data

Our settling curve analysis is based on the settleable solids present in the raw waste. The same analysis when applied to the total suspended solids gave less. consistent results between the runs, as can be seen from Table 13.1.1.

Table 13.1.1. Correlations for the Batch Settling

Data, Based on Total Solids

An assumption made in our model is that the removal obtained is the same for all classes of settleable solids. The validity of the assumption, although not tested, can be checked by taking volatile suspended I :solids measurements, together with the usual suspended I j. solids readings.

A shortcoming of our data is the fact that our batch settling column is only 8 ft. tall. This means that an effective liquid depth of only 6 ft. may be used. Most primary settling tanks have liquid depths between 8 ft. and 12 ft., thereby necessitating the extrapolation of our data, to predict their performance.

13.1.2 Tracer Studies of Primary Sedimentation Tanks

In an attempt to evaluate the level of turbulence existing in the primary settling tanks, tracer studies were performed. The results were discouraging. Tracer recoveries were of the order of 80% and consequently the detention times estimated from the exit age distributions were much lower than the hydraulic detention times. The loss of the tail end of the exit age distribution also affects whatever model we try to fit to the flow pattern. Another tracer system should have been used where the recovery obtained is much higher.

The use of ^a pulse input is also a poor choice as it generates a tail end in the exit age distribution, which is very important but where the accuracy of measurement is poor. Time Series Analysis, using a series of random pulse inputs would largely remove this source of error.

We had to revert to a simplified treatment to correct for the effects of turbulence by defining the efficiency factor. The fact that a fairly constant efficiency factor exists for the primary tanks in both the Dundas and Drury Lane plants seem to suggest that a correlation *is* possible, relating the efficiency factor with a parameter that represents the level of turbulence in the tanks.

13.1.3 Data From the Chemostat Experiments

Our Chemostat set-up is rather simple and lacks a good control over the operating conditions. A closer control should have been kept on the temperature in the reactor. With proper temperature control, the experiments could be repeated at other temperature levels. The flow rate from the constant head tank was found to vary and had to be adjusted about twice a day.

We have assumed in our calculations that the original solids in the feed are not significantly solubilized during the experiment, and that the increase in solids *in* the effluent *is* mainly due to synthesis of microbial cells. There will be a slight solubilization of the feed solids, and hence our calculated value for M, the synthesized microbial cell mass, will be low. This problems can be corrected by filtering the raw waste

through a filter press or by using a "synthetic" raw sewage, having only the soluble components.

Another assumption made was that the effluent soluble organic carbon was entirely degradable. This was not checked by taking the Biochemical Oxygen Demands of the filtered effluent. However, the least squares fit of the data points gave a correlation $u = 1.04$. $x 10^{-3}$ (C + 0.55). The intercept on the horizontal axis *is* not significantly different from zero (but *is in* fact even negative). A significant positive intercept of say 5 mg/l or greater will indicate that not all . of the effluent soluble carbon *is* degradable.

13.1.4 Data on the Aeration Studies of Activated Sludge

Since some of the biological rates *in* the kinetic scheme proposed *in* Chapter 5 are expected to be very slow, we had to aerate the return activated sludge for at least two weeks. No raw sewage was added to the return sludge. The reason *is* that the soluble degradable carbon *in* the raw sewage will have a very rapid rate of assimilation and hence creating a "stiff" condition *in* the solution of the differential equation describing its rate of reaction.

Several assumptions were made *in* the analysis of the results. The conversion factors obtained *in* Section 9.2, were used in calculating the soluble degradable carbon and in calculating the carbon equivalent of the volatile solids. Equation 5.1.12, relating the oxygen uptake rate to the rate of carbon loss due to respiration, and hence to the concentration of microorganisms, was assumed to hold, The above assumptions can be tested by integrating the area under the oxygen uptake . rate curve (which *is* the total oxygen consumption) , and comparing with the amount of carbon loss. for Run No. $D-1$,

 Δ O₂ $\frac{6200}{ }$ = 3.65 6C 1700

and for Run No. D-2,

$$
\frac{\Delta O_2}{\Delta C} = \frac{5800}{1500} = 3.86
$$

Both values are close to the factor of 3.73 obtained *in* Section 5.1.

A comparison of the results from the chemostat experiments and the extended aeration studies showed that the unit growth rate factor, k_A , was much lower in the latter case. The value of k_A obtained in the chemostat experiments was 0.0010 hr $^{-1}$ (mg/1) $^{-1}$ whilst that obtained from the extended aeration studies was 0.0006 hr^{-1} (mg/l) $^{-1}.$ The former experiments were

run under conditions where the substrate concentrations were high, whilst the latter experiments were made under food-deficient conditions. This suggests that an extended food~deficient condition could lead to decreased microbial activity, in what is commonly known as a "lag-phase". This implies then that k_A is not actually a constant but could depend on the length of substrate privation. However, in activated sludge processes, the former condition normally occurs. The food-deficient condition would occur if we use the aerobic biological reaction in a solids digestion process.

Only a few readings were taken of the soluble degradable organic carbon throughout the duration of the runs. This was because the BOD measurements required a fairly large volume of sample. A larger reactor should have been used.

13.2 Discussion of Plant Simulations

13.2.1 Simulation of *Drury Lane, Burlington, Sewage Treatment* Plant

The agreement between the predicted and observed values for the primary and final effluents were very good. The results show that primary sedimentation tanks can be modelled quite successfully *if* a sufficiently accurate

and detailed breakdown of the feed solids are provided. But normally a complete solids analysis, as required by our stream list is never performed, and the simulation may require the use of a less detailed model.

The activated sludge process was also modelled very successfully. The kinetic rates, measured experimentally, predicted the final effluent very well. An examination of the converged values for the return sludge stream showed that 42% of the volatile solids is composed of microorganisms. This is in agreement with the value of approximately 50% obtained *in* Section 5.3, for the return sludge from the Skyway Plant,

Several other case studies were made under different operating and flow conditions. Table 13.2.1 shows the individual effects of increasing capacities in the primary clarifiers, aeration tanks, secondary clarifiers and of just increasing the mixed liquor suspended solids in the aeration tanks to 3000 mg/1.

The effect of adding an extra primary clarifier was merely to improve the primary effluent to a BOD of 146 mg/1 and a TSS of 141 mg/1. There was no significant improvement in the final effluent.

The effect of adding an extra aeration tank improved the final effluent to a BOD of 15 m/gl and a TSS of 17 mg/1. No improvements were expected to occur in the primary effluent.

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Table 13.2.1. Effect of Plant Alterations, at the Same Flow.
The extra secondary clarifier reduced the TSS in the final effluent to 17 mg/1, with not much improyement in the BOD.

The results of merely increasing the mixed liquor suspended solids in the aeration tanks to 3000 mg/l were quite surprising. The final effluent improved to a BOD of 16 mg/l and a TSS of 16 mg/l. This is just like having an extra aeration tank! This is quite logical since about 50% more solids will be carried in the aeration tanks at 3000 mg/l as compared to the operating value of 2200 mg/1.

Table 13.2.2 shows the effect of increasing the raw waste flow to 2.0 and 2.4 migd., with the same influent BOD and TSS, and with no change in equipment capacity. It should be noted that the plant was originally designed for a flow of 2.5 migd with an influent BOD of 200 and an influent TSS of 180 mg/1. However, both the influent BOD and TSS have increased over the years. The reported influent BOD and TSS In 1969 were, as mentioned before, 215 mg/1 and 299 mg/1 respectively.

At the operating conditions of 1969, with a feed flow of 1.6 migd, the final effluent BOD and TSS were predicted to be 22 mg/1 and 20 mg/1 respectively. This is just bordering on the limits set by the Ontario Water

Resources Commission. At a feed flow of 2.0 migd, the primary effluent deteriorated slightly to a BOD of 153 mg/1 and a TSS of 160 mg/1, while the final effluent BOD and TSS rose to 28 mg/1 and 25 mg/1 respectively. At a feed flow of 2.4 migd, the primary effluent deteriorated further to a BOD of 156 mg/1 and a TSS of 166 mg/1 while the final effluent BOD and TSS predicted were 34 mg/1 and 30 mg/1 respectively, which is completely unsatisfactory.

The limiting equipment was found to be in the aeration tanks. If expansion is to be contemplated at the Drury Lane Plant to handle a higher flow, top priority should be given to expand the aeration tank capacity. The problem can be alleviated by maintaining a higher mixed liquor.suspended solids in the activated sludge process.

Table 13.2.2. Effect of Increased Plant Flows, with no

Change in Present Equipment Capacities

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13.2.2 Simulation of the Skyway, Burlington, Plant

In this case the activated sludge process model predicted a slightly better final effluent than was observed. However, the results show that the extended aeration process is not something special, but is merely the same activated sludge process, under a different name. The results also suggest that the 18 hour detention time required by the Ontario Water Resources Commission and other public agencies for a treatment plant without a primary clarifier is not necessary.

To show the effects of decreased detention time, or increased feed flow, on the final effluent, case studies were made at 6, 8 and 10 migd using the same feed composition. The two new final clarifiers, which are now completed, are added to the process flow diagram, together with the new {third) sludge return pump. The resulting detention times are 16, 12, and 10 hours respectively, all excluding sludge recycle.

The results are shown *in* Table 13.2.3. The final effluent at 10 migd feed flow, *is* predicted to have a BOD of 15 mg/1 and a TSS of 20 mg/1, which is still acceptable. It should be pointed out that the feed to the plant (based on 1969 values) which has a BOD

of 155 mg/l and a TSS of 185 mg/l is approximately equivalent to the primary effluent of many conventional activated sludge plants, and hence a detention time of 8-12 hours, which is sufficient for a conventional activated sludge plant, should also be sufficient for the "extended aeration" modification. The only justification for requiring more aeration time in a "extended aeration" plant is when the influent BOD and TSS are both higher than say 200 mg/1.

Stream	BOD	TSS	DOC
Raw Sewage	155	185	62
CASE STUDY WITH 6.0 migd FEED FLOW			
Final Effluent	11	13	5
CASE STUDY WITH 8.0 migd FEED FLOW			
Final Effluent	13	17	6
CASE STUDY WITH 10.0 migd FEED FLOW			
Final Effluent	15	20	

Table 13.2.3. Effect of Increased Feed Flows, at the

Skyway Plant.

CHAPTER 14

14. CONCLUSION

Simulation models for primary sedimentation, the activated sludge process, secondary sedimentation, anaerobic digestion and several other unit processes have been developed. The models were based on known and developed theories, supported by laboratory and plant data.

The primary clarifier was modelled on correlations obtained from batch column settling studies. It was found that if settleable suspended solids were used rather than the total suspended solids, better correlations resulted. This is to be expected as the total suspended solids include the non-settleable solids as well.

For the modelling of the activated sludge process, a new reaction scheme, based on the physiological activities of the bacterial and the bacterial cell mass, was proposed. Batch aeration studies of activated sludge were used to obtain estimates of the parameters used in the reaction scheme. This is an improvement over the use of mixed liquor suspended solids, as the fraction of "active mass" in the sludge is dependent on feed compositions and operating conditions in the plant.

For the secondary clarifier model, use was made of correlations developed by Villiers and the Rex Chainbelt

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Company, for the fraction of solids escaping over the wiers and the Sludge Volume Index of the sludge produced, respectively. The anaerobic digester model assumed a first order rate for the liquefaction of organic solids, the rate constants being obtained from experimental data published in the literature.

Two plants were used in the simulation studies. The first is a conventional activated sludge plant, while the second is an extended aeration plant. The results from the simulation studies were in very good agreement with plant data for the liquid streams. The actual sludge volumes produced from both plants were lower than those predicted by the simulation. The loss of solids over the wiers of the secondary clarifiers, during plant upsets or storm flows, probably accounted for the lower production of sludge from both plants,

An inter-active version of the computer simulation was also developed and tested under varying conditions. This can be used as an effective teaching aid for operators and students in the wastewater treatment area.

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APPENDIX A

PRIMARY SETTLING DATA ON BATCH COLUMN

- RUN #A.l Raw Sewage from Dundas W.P.C.P.
- RUN #A. 2 Raw Sewage from Drury Lane, Burlington. W.P.C.P.
- RUN #A. 3 Raw Sewage from Dundas W.P.C.P., 1970 Data.
- RUN #A. 4 Raw Sewage from Dundas W.P.C.P., 1969 Data.

RUN #A.

Raw Sewage from Dundas W.P.C.P.

Settling Data: Original suspended solids average of duplicates.

Average of initial solids = 250 mg/l (rounded up) Average of final non-settleable solids = 82 mg/1 {rounded up) Hence total settleable solids = $168 \text{ mg}/1$ $% of settleable solids = 100 (SS - 82)/168$

Weight % settleable solids as a function of time and depth:

181

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Fraction of settleable solids removed, as a function of time and depth, using Equation 4.1.6:

 $(Con't.)$

 RUN $#A.2.$

Raw Sewage from Drury Lane, Burlington W.P.C.P. Settling Data: Original suspended solids average of duplicates

Average total *initial* suspended solids = 275 mg/1 Average final suspended solids = 88 mg/l Total settleable solids = $187 \text{ mg}/1$ Weight % settleable solids = 100 (SS - 88)/187 Temperature = 20° C

Weight % settleable solids as a function of time and depth:

Weight fraction of settleable solids removed, as a function fo time and depth, using Equation 4.1.6:

186

 $(Con't.)$

Ń

RUN #A.3.

Raw Sewage from Dundas W.P.C.P., data

of 1970.

Settling Data: Original suspended solids average of duplicates.

Average total initial suspended solids = 250 mg/l Average final suspended solids = $73 \text{ mg}/1$ Total settleable solids = 177 mg/l Weight % settleable solids = 100 (SS - 72)/177

Weight % settleable solids as a function of time and depth:

Weight fraction of settleable solids removed, as a function of time and depth, using Equation 4.1.6:

 \bullet

RUN #A.4.

Raw Sewage from Dundas W,P.C.P.

Settling Data: Original suspended solids average of duplicates

- data of Hudspith et al, 1969

Average of initial solids = 246 mg/l Average of final non-settleable solids = 92 mg/l Hence total settleable solids = 154 mg/1 % settleable solids = 100 (SS - 92)/154

*Insufficient mixing?

Weight % settleable solids as a function

of time and depth:

 $\ddot{}$

Weight fraction of settleable solids removed as a function of time and depth, using Equation 4.1.6:

APPENDIX B

Exit Age Distributions of Primary Tanks RUN #B.1-3 Dundas Plant RUN $#B.4-6$ Drury Lane Plant SETTLEABLE SOLIDS REMOVAL DATA

TRACER CONCENTRATION (ppm)

TIME (minutes)

Table B.l. Settleable Solids Removal· Data

APPENDIX C

DATA FROM THE CHEMOSTAT EXPERIMENTS

Average value of $Y = 0.57$

Standard deviation = 0,037

Temperature = $25^{\circ}C + 3^{\circ}C$

Source: Filtered raw sewage from Skyway, Burlington

 $(MOTE: M is obtained from the increase in suspended solids by the conversion$ factor of 2.1. Both C and Mare expressed in terms of carbon).

 $\ddot{}$

APPENDIX D

EXTENDED AERATION STUDIES ON ACTIVATED SLUDGE

- {a) Two runs using return sludge from the Skyway, Burlington W.P.C,P.
- (b) Replicates analyses using five simulaneously \sim drawn samples.

Aeration of return activated sludge from the

Skyway, Burlington W.P.C.P.

Original Data:

Temperature = $23 + 2^{\circ}C$

 $pH = 7.5 \pm 0.3$

Air Rate = 1.0 litre/hr/litre reactor volume Ash content of sludge:

initial = 1650 mg/l $final = 1480 mg/l$ average = 1520 mg/1

Converted Data:

VSS = Volatile suspended solids (mg/ 1) {see Section $9.1)$ $=$ (Total Suspended Solids - 1520)/2.1

 \mathcal{L}

 $SNDOC = Soluble non-degradable organic carbon (mhC/1)$ = soluble organic carbon - SDOC

Aeration of return activated sludge from the

Skyway, Burlington W.P.C.P.

Original Data:

Temperature = $23 \pm 2^{\circ}$ C $pH = 7.4 \pm 0.4$ Air Rate = 1.0 litre $\ar{air/hr/l}$ itre reactor volume Ash content of sludge: initial = 1700 mg/l $final = 1890 mg/l$ average = 1800 mg/1

Converted Data:

 VSS = volatile suspended solids (mgC/1) (see Section 9.1)

- \texttt{SDOC} = soluble degradable organic carbon (mgC/1) = BOD_5 (filtrate)/1.9
- SNDOC = soluble non-degrddable organic carbon $(mgC/1)$

soluble organic carbon - SDOC \equiv

(b) Replicate Analyses on Five Simultaneously-drawn

Samples:

. Original Data:

NOTE: (a) 10 ml samples were used for VSS (b) dilution for BOD_5 is 25%. Filtrate
is close to saturation. Hence no is close to saturation. need for $0₂$ correction.

The above data is converted to the similar values used in the four responses for the objective function:

 (c) SNDOC = soluble organic carbon - SDOC

Inverse of Variance - Covariance Matrix

APPENDIX E

DATA FOR THE STRIPPING RATE OF PURGEABLE SOLUBLE ORGANIC CARBON BY AERATION WITH AIR

RUN #E.l.

Air Rate = 2.1 cu.ft.air/hr/cu.ft. reactor

Soluble non-purgeable organic carbon assumed to be = 99 mg/l soc = total soluble organic carbon SPOC = soluble purgeable organic carbon $SPOC = SOC - 99$

RUN #E.2.

Air Rate= 1.0 cu.ft.air/hr/cu.ft. reactor

Soluble non-purgeable organic carbon assumed to be = 106 mg/l SOC = total soluble organic carbon SPOC = soluble purgeable organic carbon $SPOC = SOC - 106$ $\mathcal{L}_{\mathcal{A}}$

RUN #E.3.

Air Rate = 0.5 cu.ft.air/hr/cu.ft. reactor

Soluble non-purgeable organic carbon assumed to be = $111 \text{ mg}/1$ soc = total soluble organic carbon SPOC = soluble purgeable organic carbon $SPOC = SOC - 111$

RUN #E.4.

Air Rate = 1.5 cu.ft.air/hr/cu.ft. reactor

Soluble non-purgeable carbon assumed to be = 98 mg/l soc = total soluble organic carbon SPOC = soluble purgeable organic carbon $SPOC = SOC - 98$

APPENDIX F

DATA OF ACTIVATED SLUDGE

SETTLING FROM *P.* LEUNG

Data of Peter Leung

The predicted effluent SS_e was found to be

 $SS_{\circ} = \frac{2560}{\cdots}$ ss e $(MLSS)^{0.57}$ (TA)^{0.20} 2560 ors_{e} $\begin{array}{ccc} & \text{ss}_{\text{e}} \\ \text{or} & \text{xRSS} & = & \end{array}$ (MLSS) $(MLSS)^{1.57}$ (TA)^{0.20}

The SVI was not found to correlate well.

Figure F.1. Comparison of Observed Effluent Solids with Correlation $\overline{}$

(Data of P. Leung)

Predicted Effluent SS (mg/1)

220

APPENDIX G

ANAEROBIC DIGESTION DATA

- (a) Conventional Digester -Data of Rankine (32).
- (b) High Rate Digester -Data of Torpey (50), Roy and Sawyer (34) and Estrada (12).

(a) Conventional Digester

Data of Rankine (32).

(b) High Rate Digester

APPENDIX H

INTER-RELATIONSHIPS BETWEEN ORGANIC CARBON,

BIOCHEMICAL OXYGEN DEMAND AND VOLATILE SUSPENDED SOLIDS

TBOD = total BOD_5 (mg/1)

DBOD = BOD_5 of soluble organics (mg/1)

 $TOC = total organic carbon (mg/1)$ U

 $SOC = soluble organic carbon (mg/l)$

APPENDIX I

SIMULATION DATA AND RESULTS FOR THE DRURY LANE,

BURLINGTON, SEWAGE TREATMENT PLANT

227

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DESIGN DATA 228

PRIMARY TREATMENT

Screening

1" bar screens

Grit Hemoval

Type: Grit channels Helention: 0. 8 min

Primary Sedimentation

Type: Walker Process Size: Two 49.3' x 18' x 12.25' (135, 700 gal) Retention: 1.3 hr Loading: Surface, $1400 \text{ gal/ft}^2/\text{day}$ Weir, $17,100$ gal/ft/day

SECONDARY TREATMENT

Aeration Tanks

Type: Diffused air; triple-pass Size: Two tanks, each with 2 passes 118' x 18' x 10. 7' 1 pass 85. 5' x 181 x 10.71 $(833, 000 \text{ gal.} \text{ total})$ Retention: 8.0 hours

Air Supply

One Sutorbilt- 1500 cfm Two Roots-Connerville- 750 cfm

Diffusers - (each tank)

1) 132 Schumacher Brandel tubes in first two passes

2) 41 Spargers on 2' centres in third pass

Secondary Sedimentation

Type: Rex Unitube Tow-Bro Size: Two $50'$ dia x $10.6'$ swd $(260, 000 \text{ gal})$ Retention: 2. 5 hr Loading: Surface, $1000 \text{ gal/ft}^2/\text{day}$ Weir, 8500 gal/ft/day

CHLORINATION

Type: Kent

Chlorine Contact Chamber

in outfall

OUTFALL

to Lake Ontario

SLUDGE HANDLING

Digestion System

Type: Two-stage

Primary- Size: Two 40' dia tanks (313, 000 gal total), depth = $20'$ Loading: $2.7 \text{ lb/ft}^3/\text{mo}$

Secondary $-$ Size: One $40'$ dia tank $(143, 000 \text{ gal})'$ Loading Total: 1.9 lb/ft3/mo depth = $18'$

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RUN WITH AVERAGE FLOW CONDITIONS -1969

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SUMMARY REPORT ON PLANT OPERATIONS - CASE

APPENDIX J

SIMULATION DATA AND RESULTS FOR THE SKYWAY,

BURLINGTON, SEWAGE TREATMENT PLANT

238

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DESIGN DATA

PRETREATMENT

Coarse Screening

Type: Link Belt, mechanically-cleaned Size: $2\frac{1}{2}$ ¹¹ space

Lift Station Pumps (@ 30' tdh)

2 @ 6 mgd each; 2 @ 5 mgd each

Screening

Type: Link Belt, mechanically-cleaned Size: 1" space

Grit Removal

Type: Air degritter Size: One 20. 3' x 25' x 14. 2' Retention: 20. 8 min

SFCONDARY TREATMENT

Aeration Tanks

Type: Diffused air, single pass Size: Six 270' x 27' x 15' (660,300 ft.,or 4.12 mil gal) Retention: 16.5 hr Loading: 9.5 lb BOD/1000 ft³

Air Supply

Type: Hoffman multi-stage

centrifugal blowers Size: 3- 4000 cfm (max); 1-6000 cfm (max) Diffusers

Type: Saran-covered flexofusers Spacing: 2 50 tubes per tank

Secondary Sedimentation

Type: Eimco Size: Four $60' \times 60' \times 12'$ swd $(538, 000 \text{ gal})$ *(only two completed)* Retention: 4.1 hours Loading: Surface, 434 gal/ft²/day Weir, $7,100 \text{ gal/ft/day}$

CHLORINATION

Type: Fischer & Porter Size: Two 2000 lb/day

Chlorine Contact Chamber

-nil

- chlorination in outfall

OUTFALL

- to Hamilton Harbour

SLUDGF HANDLING

Type: Thickening tank, decanted Size: One 20' dia x 9' depth $(2830 \text{ ft}^3 \text{ or } 1760 \text{ gal})$

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RUN WITH AVERAGE FLOW CONDITIONS IN 1969

FINAL OUTPUT STREAMS FOR MODULE 18888

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0.000 -983 $.001$ 0.000 0.000 $0 - 000$ $8,550$ 0.000 \$\$\$\$\$ MIXER1

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ويردعان المحامين **Automaker**

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 $\label{eq:1} \delta \theta(t) = \frac{1}{2} \int_{0}^{t} \left(\frac{d\theta}{dt} \right) \left(\frac{d\theta}{dt} \right) \, dt$

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 $\label{eq:1} \left\langle \mathbf{v}^{\dagger}\right\rangle =\left\langle \left\langle \left\langle \mathbf{v}^{\dagger}\right\rangle \mathbf{v}^{\dagger}\right\rangle \right\rangle +O_{\mathcal{A}}$

 $\label{eq:1} \phi_{\alpha\beta}(\theta) = \frac{1}{\sqrt{2\pi\sigma_{\alpha\beta}}}\,.$

 $\label{eq:1} \frac{1}{\sqrt{2\pi}}\int_{0}^{\frac{\pi}{2}}\frac{d\mu}{\sqrt{2\pi}}\left(\frac{d\mu}{2\pi}\right)^2\frac{d\mu}{2\pi}\left(\frac{d\mu}{2\pi}\right)^2\frac{d\mu}{2\pi}\left(\frac{d\mu}{2\pi}\right)^2\frac{d\mu}{2\pi}\left(\frac{d\mu}{2\pi}\right)^2\frac{d\mu}{2\pi}\left(\frac{d\mu}{2\pi}\right)^2\frac{d\mu}{2\pi}\left(\frac{d\mu}{2\pi}\right)^2\frac{d\mu}{2\pi}\left(\frac{d\mu}{2\pi}\right)^2\frac$

 $\label{eq:2.1} \frac{1}{\sqrt{2}}\int_{0}^{\infty}\frac{1}{\sqrt{2\pi}}\left(\frac{1}{\sqrt{2\pi}}\right)^{2\alpha} \frac{1}{\sqrt{2\pi}}\int_{0}^{\infty}\frac{1}{\sqrt{2\pi}}\frac{1}{\sqrt{2\pi}}\frac{1}{\sqrt{2\pi}}\frac{1}{\sqrt{2\pi}}\frac{1}{\sqrt{2\pi}}\frac{1}{\sqrt{2\pi}}\frac{1}{\sqrt{2\pi}}\frac{1}{\sqrt{2\pi}}\frac{1}{\sqrt{2\pi}}\frac{1}{\sqrt{2\pi}}\frac{1}{\sqrt{2\pi}}\frac{1}{$

 $\label{eq:2.1} \frac{1}{\sqrt{2}}\int_{0}^{\infty} \frac{dx}{\sqrt{2\pi}}\,dx\leq \frac{1}{2}\int_{0}^{\infty} \frac{dx}{\sqrt{2\pi}}\,dx.$

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 $\label{eq:1} \mathcal{A}(\mathbf{z}) = \mathcal{A}(\mathbf{z}) \mathcal{A}(\mathbf{z}) = \mathcal{A}(\mathbf{z}) \mathcal{A}(\mathbf{z})$

 $\label{eq:2} \hat{f}_{\text{in}}(t) = \frac{1}{2} \exp\left(-t\right) \exp\left(-t\right) \exp\left(-t\right) \exp\left(-t\right) \exp\left(-t\right) \exp\left(-t\right)$

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constraint the field $\hat{f}_{\rm{max}}(x)$, and $\hat{f}_{\rm{max}}(x) = \hat{f}_{\rm{max}}(x)$

 $\label{eq:2.1} \gamma_{\mu\nu}(\tau) = \gamma_{\mu\nu} \gamma_{\mu\nu}(\tau) \; \text{and} \; \; \text{isomorphism of } \; \eta_{\mu\nu}(\tau) = \tau_{\mu\nu}(\tau) \; \; \forall \; \alpha \in \partial \Omega,$

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 $\label{eq:1.1} \mathbf{F}(\alpha)=\mathbf{F}(\alpha)+\mathbf{F}(\alpha)\mathbf{F}(\alpha)\mathbf{F}(\alpha)\mathbf{F}(\alpha)\mathbf{F}(\alpha)$ and α

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SUMMARY REPORT ON PLANT OPERATIONS - CASE

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APPENDIX K

Computer Listings

LISTING OF THE GEMCS EXECUTIVE

PROGRAM TST (INPUT, OUTPUT, TAPF5=INPUT, TAPF6=OUTPUT) GFMCS/6400 ***** PRINTING CONTROL CHARACTERS ***** KPRNT(1)=1 CAUSES PRINTING OF NODITIEER AND OUTPUT STREAMS
KPRNT(2)=1 CAUSES PRINTING OF INITIAL STREAMS
KPRNT(2)=1 CAUSES PRINTING OF INITIAL STREAMS
KPRNT(3)=1 CAUSES PRINTING OF MODULES SETS
KPRNT(3)=1 CAUSES PRINTING O AFTER TWO LOOPS COMMON DECLARATIONS - JUNE 8 ***** ***** LLST(50), NS(100), EN(100), SI(4,30), SO(4,30), KPRNT(10) COMMON COMMON ISSNÉSJÚSĽOOPSNINSNOUTSMÁNSISPSNCSIIISNCALCSNOCOMPSNSR
COMMON MODESNPLNTSLOOPC
COMMON SN(25,30)SEEN(600)SNPOINT(25,2) DIMENSION NAME(20) DIMENSION NAME(20)
NAME(2)=8H CONTLI
NAME(3)=8H SETSTI
NAME(4)=8H SETSTI
NAME(5)=8H SETSPI
NAME(5)=8H SEPACI NAME (6)=8H PRISET1
NAME (7)=8H ACTSL1
NAME (8)=8H DPTR1
NAME (9)=8H SECLAR1
NAME (10)=8H TRICK1 PRISET1
ACTSL1 NAME (11) = 8H
NAME (12) = 8H
NAME (13) = 8H
NAME (14) = 8H GRITI ANDIG1 CHLOR1 **COSTI** NAME (15) = 8H
NAME (16) = 8H
NAME (17) = 8H
NAME (18) = 8H SCREEN1
REPT01 CONT_C2 CONVOI
CASET $NAME(19) = 8H$ NAME $(20) = 8H$ THICK1 $III = 25$ CALL DLOAD1
LOOPC=1
WRITE(6,196)_LOOPC READ(5,198) TITLE
WRITE(6,200) TITL TITLE CALCULATING EQUIPMENT IN CALCULATION ORDER LIST S USED AS AN INDICATOR
FOR A STRAIGHT THROUGH CALCULATION
FOR A RECYCLE OPERATION IK IS USED $\overline{K} = \overline{1}$
 $\overline{K} = 0$ \overline{I} K=0 IF(LOOP.EQ.999) IK=1 CONTINUE $NC = 1$ MODĒ=0
KTEST=0 $LOOP = 1$ IFIIK.EQ.0) GO TO 98

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> CCCCC $\frac{c}{100}$

MODE=1
KTEST=1
LOOP=999 56
 502 **CONTINUE** READ EN VECTOR IF (KPRNT(5)) 108,108,104 $\overline{1}04$ CONTINUE $(6, 182)$ LO
6 IKE=1,III WRITE LOOP DO 106 IF (SN(IKE,1).LE.0.0) GO TO 106
WRITE (6,194) (SN(IKE,J),J=1,JJ) $rac{1}{c}$ ⁰⁶ CONTINUE NE=LLST(NC)
MM=NE $\overline{1}08$ CALL DISKIO (1, MM)
NN=EN(3)+.001 $NIN = EN(6) + .001$ NOUT=EN(11)+.001
NTYPE=ABS(EN(2))+.001 $KSW = 0$

IF (E)

IF (L) IF (EN(2).LE.0.) KSW=1
IF (LOOP.GT.2) ISP=0
IF(LOOP.LE. 2) ISP=1 $\frac{c}{c}$ FINDING INPUT STREAMS IF(NIN-EQ.U) GO TO 128
DO 122 I=1,NIN
S=EN(I+6) CALL STRÉAM (S)
IF (IS-III) 114,114,110 $15 \text{ N}111111$

CONTINUE

SI(1, 1) = S

DO 112 J=2, J

SI(1, J) = 0.

CONTINUE 110 112 GO' TO 122
DO 116 J=1, JJ
SI(1, J)=SN(1S, J)
CONTINUE 114 116 $(INT(5+.001)) 122.122.118$ ĪĔ CONTINUE
M=S+.001
IF (NS(M)-6) 122,120,120 118 120 CONTINUE SN(IS, 1)=0.
CONTINUE 122 $IF (ISP)$ 128, 128, 124 WRITE (6,184) NE , NAME(NTYPE) 124 DO 126 IKE=1,NIN
WRITE(6,194) WRITE (6 $(6,194)$ (SI(IKE, J), J=1, JJ) $\frac{126}{128}$ **CONTINUE** خ
C CALL MODULE (NTYPE) CCC
CCC STORING OUTPUT STREAMS AND PRINTING IF(NOUT) 166,166,136
CONTINUE
DO 150 I=1,NOUT
S=C(1,1)=S
SO(1,1)=S
M=5++001
LE(NS(M)=6) 140,150, 136 $\ddot{}$ $IF(\overline{NS(M)}-6)$ $140, 150, 140$ CALL STREAM
IF (IS-III)
CALL STREAM
IF (IS-III) 140 (-5) $146, 146, 142$ $(0, 1)$ 142 $146, 146, 144$

WRITE (6,190)
GO TO 150
DO 148 J=1, JJ
SN(IS, J)=SO(I, J)
CONTINUE 250 144 146 148 $\frac{1}{2}$ 50 CONTINUE IF (ISP) 158,158,154
WRITE (6,186) NE , NAME(NIYPE)
DO 156 IKE=1,NOUT \cdot IF 154 WRITE(6,194)
WRITE (6,194) (SO(IKE,J),J=1,JJ) CONTINUE 156 $\frac{2}{158}$ IF (LOOP-999) 166,160,160
IF (KSW.NE.U) GO TO 166
WRITE(6,188) NE, NAME(NTYPE)
DO 164, IKE=1,NOUT 160 WRITE(6,194)
WRITE (6,194) (SO(IKE,J),J=1,JJ) 164 CONTINUE 166 $NC=NC+1$ IF (NC-NCALC) 102,102,168 $\frac{C}{168}$ CONTINUE
IF (LOOP-999) 170,172,172
CONTINUE 170 $NC = 0$ TO 166
(KTEST) 174,174,176 GO TO IF (KTĒ
NC=0
KTEST=1 $\frac{172}{174}$ KIESI=1

MODE=1

GO TO 166

LOOPC=LOOPC+1

IF (LOOPC-NPLNT) 178,178,180

WRITE(6,196) LOOPC

CALL DLOAD3

GO TO 100 176 178 $\frac{6}{180}$ WRITE (6,192) **STOP** \subset $(1H0, 32H5555)$
 $(1H0, 3H5555)$ SN TABLE ON ENTERING LOOP , 15,7H \$\$\$\$\$)
INPUT STREAMS FOR MODULE , 15,4H - , 182 FORMAT 184 FORMAT \$\$\$\$\$) $1A8,7H$ FÖRMAT (1HŰ, 32HSSSSS)
'7H SSSSS) OUTPUT STREAMS FOR MODULE . 15,4H - . A8, 186 2`7H``^\$\$\$\$\$`)
`FORMAT`{1H0}
}_A8?7H` \$\$\$\$ \overline{z} FINAL OUTPUT STREAMS FOR MODULE , 15,4H 188 3 190 FORMAT ERROR IN SN ***** FORMAT (1HO, 38H************ END OF
FORMAT (1H , 10F12.3)
FORMAT (1H1, 28H\$\$\$\$\$\$\$\$\$\$\$\$\$\$\$\$\$\$\$ $\frac{192}{194}$ END OF EXECUTION ********** -1 55555555555 CASE 196 $,14,24H$ 1\$\$\$\$\$\$\$\$`,7 $, / /$ 198 200 FORMAT(1H0,10A8) END

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SUBROUTINE STREAM (S)

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7 C COMMON LLST(50),NS(100),EN(100),SI(4,30),SO(4,30),KPRNT(10)
COMMON IS,NE,JJ,LOOP,NIN,NOUT,MSN,ISP,NC,III,NCALC,NOCOMP,NSR
COMMON MODE,NPLNT,LOOPC
COMMON SN(25,30),EEN(600),NPOINT(25,2) DO 2 IS=1, III
IF (ABS(SN(IS, 1)-ABS(S))-.001) 4,2,2 $\begin{array}{c}\n\stackrel{\frown}{\text{Conv}}\stackrel{\frown}{\text{Conv}}\stackrel{\frown}{\text{Conv}}\stackrel{\frown}{\text{Conv}}\stackrel{\frown}{\text{Conv}}\stackrel{\frown}{\text{Conv}}\stackrel{\frown}{\text{Conv}}\stackrel{\frown}{\text{Conv}}\stackrel{\frown}{\text{Conv}}}\n\end{array}$ RETURN **END**

```
\mathcal{L}SUBROUTINE DISKIO (IPNT, MM)
COMMON LLST(50),NS(100),EN(100),SI(4,30),SO(4,30),KPRNT(10)<br>COMMON IS,NE,JJ,LOOP,NIN,NOUT,MSN,ISP,NC,III,NCALC,NOCOMP,NSR<br>COMMON MODE,NPLNT,LOOPC<br>COMMON SN(25,30),EEN(600),NPOINT(25,2)
THIS SUBROUTINE SIMULATES DISKIO ON360/30<br>IF IPNT=1. READS FROM MODULE SETS TABLE<br>IF IPN=2. WRITES ONTO MODULE SETS TABLE<br>MQ=NPOINT(MM,1)<br>ML=NPOINT(MM,2)<br>IF (IPNT-1) 6,2,6
DO 4 I = 1.1MLEN(I)=EEN(MQ+I)<br>CONTINUE<br>GO TO 14<br>IF (IPNT-2) 12,8,12
 \overline{D}O 1\overline{O} \overline{I}=1, ML
EENIMQ+IEEN(I)CONTINUE<br>GO TO 14<br>WRITE (6,16) IPNT, MM
RETURN
FORMAT (1HU,22H DISKIO ERROR-IPNT, MM=,15,1X,15)
END
```

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SUBROUTINE DLOAD1
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CONTINUE READING MODULE SETS, NOE IN NUMBER READ (5,100) ANOE NOE=ANOE

IF (KPRNT(3)-1) 64,62,64

WRITE (6,104) NOE

DO 72 I=1,NOE $I \stackrel{\uparrow}{\uparrow} \stackrel{\sim}{\uparrow} \stackrel{\sim}{\uparrow} \stackrel{\sim}{\uparrow} \stackrel{\sim}{\uparrow} \stackrel{\sim}{\uparrow} \stackrel{\sim}{\uparrow} \stackrel{\sim}{\uparrow} \stackrel{\sim}{\downarrow} \stackrel{\sim}{\down$ ĐŌ $EN(IZ) = 0$.
CONTINUE READ (5,100) (EN(N), N=1,5)
NN=EN(3) NN=EN(3)

NCOUNT=NCOUNT+NN

READ (5,100) (EN(N),N=6,NN)

IF (KPRNT(3)-1) 70,68,70

WRITE (6,102) (EN(N),N=1,NN)

WRITE (6,102)

MM=EN(1)+.001

NPOINT(MM,1)=NCOUNT-NN

NPOINT(MM,1)=NCOUNT-NN

NPOINT(MM,1)=NCOUNT-NN

NPOINT(**CONTINUE** (KPRNT(4).GT.0) GO TO 74
TO 78 $F_{\rm GO}$ $(6, 106)$
 $I = 1, 25$
 $(6, 90)$ WRITE
DO 76 I=1,25
WRITE (6,990) I,NPUIN.
CONTINUE
CONTINUE
CONTINUE.
CORMAT STATEMENTS FOR DLOAD WRITE I , NPOINT(I,1), NPOINT(I,2) 2888
FOR -1 : ORMAT(1H0,1H\$,19X,10A8,19X,1H\$)
'ORMAT (1H0,22HPRINTING NCONT,LLST,NS)
'ORMAT (1H0,24HPRINTING INITIAL STREAMS)
'ORMAT (1K,316)
'ORMAT (1X,316) 88889999999111111111111 FORMAI FORMAT FORMAT (IHO,20HPRINTING MODULE)
FORMAT (IHO,41HNUMBER OF MODULE)
FORMAT (IHO,41HNUMBER OF MODULE)
FORMAT (IHO,27HCALCULATION ORDE)
FORMAT (IHO,23HNUMBER OF COMPON)
FORMAT (IHO,33HNUMBER OF STREAM)
FORMAT (IHO,33HNUMBER OF MODULES
DN ORDER IN CALCULATION ORDER = $, 15)$ COMPONENTS = , 15)
STREAM CODES READ =
INITIAL STREAMS READ $,15,7)$ $,15,7)$ $=$ SETS RI
LENGTH
CASES $READ =$ $,15,7)$ $\frac{7}{15}$ **END**

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68 70

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74

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SUBROUTINE MODULE(NT)

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SUBROUTINE DLOAD3

COOOO WRITTEN BY P. TAN 1971
MODULE TO RUN NEW CASES
READS IN NEW STREAMS AND MODULE SETS COMMON LLST(50),NS(100),EN(100),SI(4,30),SO(4,30),KPRNT(10)
COMMON IS,NE,JJ,LOOP,NIN,NOUT,MSN,ISP,NC,III,NCALC,NOCOMP,NSR
COMMON MODE,NPLNT,LOOPC
COMMON SN(25,30),EEN(600),NPOINT(25,2)
DIMENSION TITLE(10) $\frac{c}{c}$ READ(5,100) TITLE
FORMAT(10A8)
WRITE(6,101) TITLE 100 $\mathcal{\check{E}}^{01}$ FORMAT(1H0,10A8) READ NUMBER OF STREAM CHANGES READ(5,112) ANSR
FORMAT(5F12,4) 112 E FORMATISFIZACI

NSR=ANSR+0.001

IF(NSR.LE.0) GO TO 300

WRITE(6,111) NSR

111 FORMAT(1HC,15,25HINITIAL STREAMS ARE READ -1 $\frac{c}{c}$ READ STREAMS
DO 530 I=1,NSR
BORROWS VECTOR FROM SO(4,J), J=1,JJ
READ(5,112) (SO(4,J),J=1,JJ)
CALL CONVERT(2,2,4,2,4) \overline{C} CALL BALNCE(2,4) WRITE(6,113)
WRITE(6,113) (SO(4,J),J=1,JJ)
113 FORMAT(1X,10F12.4) S=SO(4,1)
CALL STREAM(S)
IF(III-IS) 400,401,401 400 CALL STREAM(0.0)

IF(111-15) 403,401,401

401 DO 402 J=1,JJ

402 SN(15,J)=SO(4,J) $53\overline{0}$ CONTINUE $\frac{c}{c}$ READ NUMBER OF MODULE CHANGES
READ(5,112) ANOE
NOE=ANOE+0.001 300 IF (NOE.LE. 0) GO TO 301 114 FORMAT(1HU, 15, 21HMODULE SETS ARE READ λ $\frac{c}{c}$ READ MODULES DO 540 I=1, NOE
DO 541 IZ=1, 100
EN(IZ)=0.0 541 READ(5,112) (EN(N), N=1,5) $NN = EN(3) + 0.001$ MM=EN(1)+0.001
READ(5,112) (EN(N),N=6,NN) WRTTE $(6, 113)$
WRITE $(6, 113)$ (EN(N), N=1, NN)
CALL DISKIO(2, MM) CALL DIS
CONTINUE 540 301 CONTINUE **RETURN** C 403 WRITE(6,405)
405 FORMAT(1H0,* ERROR IN SN - VECTOR SPACE INSUFFICIENT *,//) C REIURN END

SUBROUTINE SETST1

COMMON LLST(50),NS(100),EN(100),SI(4,30),SO(4,30),KPRNT(10)
COMMON IS,NE,JJ,LOOP,NIN,NOUT,MSN,ISP,NC,III,NCALC,NOCOMP,NSR
COMMON MODE,NPLNT,LOOPC
COMMON SN(25,30),EEN(600),NPOINT(25,2)

ALTERS THE TEMPERATURE IN ANY STREAM
EN(4) = CHANGE IN TEMP (+VE FOR INCREASE AND -VE FOR DROP)
WRITTEN BY P. TAN
FOR WWTP SIMULATION FOR WWTP

DO 1 $I = 2$, U
SO(1, I) = SI(1, I)
SO(1, 4) = SI(1, 4) + EN(4)

RETURN END

SUBROUTINE SETSP1

COMMON LLST(50), NS(100), EN(100), SI(4,30), SO(4,30), KPRNT(10)
COMMON IS, NE, JJ, LOOP, NIN, NOJT, ASN, ISP, NC, III, NCALC, NOCOMP, NSR
COMMON MODE, NPLNT, LOOPS COMMON SN($\overline{2}5$, 30), EEN(600), NPOINT(25, 2)

WRITTEN BY P. TAN
FOR WWTP SIMULATION
ALTERS THE HEAD IN ANY STREAM
EN(5)= CHANGE IN HEAD (+VE FOR GAIN, -VE FOR LOSS)

DO 1 I=2, JJ
SO(1, I)=SI(1, I)
SO(1, 5)=SI(1, 5)+EN(5) **RETURN** END

SUBROUTINE CONTL1

LLST(50), NS(160), EN(100), SI(4, 30), SO(4, 30), KPRNT(10)
IS, NE, JJ, LOOP, NIN, NOUT, MSN, ISP, NC, III, NCALC, NOCOMP, NSR **COMMON COMMON** COMMON MODE , NPLNT , LOOPC COMMON SN(25,30), EEN(600), NPOINT(25,2)

WRITTEN BY A.I.JOHNSON AND T.TOONG MAY 1968

EN

- $\frac{1}{2}$.
-
- $\overline{4}$.
- LIST
EQUIPMENT NUMBER
EQUIPMENT TYPE -2
LENGTH OF EN LIST-21
LARGEST LOOP IDENTIFICATION NUMBER IN PROCESS FLOW DIAGRAM
PRINTING CONTROL-NUMBER OF LOOPS BETWEEN PRINTING OF THE ST
UNDERFIEST JONLY NON ZERO COMPONENTS* AND **STREAM** 5.4 UNDER'TEST JONLY NON ZERO COMPONENTS* AND THEIR FRACTIONALS
-15. TEN ZEROES
-15. TEN ZEROES
NO. OF MODULES IN CALC ORDER LIST CONTROLLED BY CONTL1
MAXIMUM NO. OF LOOPS
STREAM NUMBER OF STREAM TESTED
FRACTIONAL TOLERANCE
LO
- $6 15$.
- $\frac{16}{17}$.
-
- $^{18}_{19}$.
- $20.$
- $21 \bar{2}\bar{2}\cdot-(19+JJ) = 0.0$
-

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SUBROUTINE CONV01

 $\overline{\mathbf{1}}$

 $\frac{c}{1}$ 600 610 6 3 O· COMMON LLST(50l,NS(100l,EN(100l,SI(4,30l,S0(4,30l ,KPRNT(10l COM~ON IS,NE,JJ,LOOP,NIN,NOUT,MSN,ISP,NC,III,NCALC'NOCOMP'NSR COMMON MODE,NPLNT,LOOPC
COMMON SN(25,30),EEN(600),NPOINT(25,2) WRITTEN BY P. TAN ... 25/6/71 FOR WWTP SIMULATION CONVERGENCE PROMOTION ON A STREAM EN VECTOR
********* ********* 1• = MODULE NUMBER 2. MODULE TYPE (18)
3. MODULE LENGTH $4. = LQQP \cdot A$ WHICH 5. = STREAM NUMBER 5•-15• = TEN ZEROES
16• LOOP COUNTER -
17• - (16+(JJ-2)*2) (16 + (<mark>JJ-2)*2</mark>
MODULE IS EFFECTIVE OF STREAM TO BE CONVERGENCE-PROMOTED INITIALLY SET AT ZERO STORAGE SPACE FOR STREAM NLOOP = EN(4)+0.01 IFILOOP.LT.NLOOPl RETURN KOUNT=EN<16}+0.01 KOUNT=KOUNT+1 S=EN(5) CALL ŠTREAM(\$)
GO TO (1,2,3), KOUNT DO 600 J=<mark>3,JJ</mark>
EN(14+J)=SN**(**IS,J) EN(16)=KOUNT RETURN DO 610 J=3,JJ
EN(12+JJ+J)=SN(IS,J) EN(16)=KOUNT RETURN DO 630 J=13,JJ DENOM=2•0*EN!12+JJ+Jl-EN<14+Jl-SN<IS,Jl IF!DENOM.EQ. 0.01 GO TO 630 SN<IS,Jl=!EN<l2+JJ+Jl*EN!12+JJ+Jl-SNIIS,Jl*EN(l4+Jll/DENOM $EN(14+J)$ =SN(15,J) CONTINUE
EN(16)=1.0 $I = S + 0.01$ CALL BALNCE(1,I) RETURN END

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LISTING OF THE PRIMARY SETTLING TANK MODEL

SUBROUTINE PRISET1 LLST(50),NS(100),EN(100),SI(4,30),SO(4,30),KPRNT(10)
IS,NE,JJ,LOOP,NIN,NOUT,MSN,ISP,NC,III,NCALC,NOCOMP,NSR **EOMMON** MODE , NPLNT, LOOPC
SN(25,30), EEN(600), NPOINT(25,2) COMMON COMMON SIMULATES A PRIMARY SETTLING TANK
WRITTEN BY P. TAN
FOR WWTP SIMULATION
FOR REVISED STREAM LIST 2/9/71 EN VECTOR UNIMPARD FORMAT SINCE IN PARALLEL OF WATER)

STANDARD FORMAT

NUMBER OF TANKS IN PARALLEL OF WATER)

HEAD LOSS THROUGH INREAM (RAW WASTE STREAM)

SECOND INPUT STREAM (RAW WASTE TROM THE DIGESTERS - IF

SECOND INPUT STREAM ********* $1 - 15$ $\frac{4}{5}$. $\frac{1}{8}$ $\frac{1}{2}$ $\frac{1}{2}$ ANY) $\overline{16}$. $17.$ $18.$ $190...$
 2222 $23 - 24$ $25.$ CALL ALKMIX(PH) $[00, 700, J=2, JJ]$
50(1, J)=0.0 $SO(2, J) = 0.0$ $SO(3, J) = 0.0$ IF(NIN.EQ.1) GO TO 40 STREAMS - HENCE SETTLE MUCH
DISP IS THE FRACTION OF SOLIDS
ARE RESUSPENDED. INPUT STREAMS 2 AND 3 ARE SLUDGE
FASTER THAN SOLIDS IN RAW WASTE.
FROM INPUT STREAMS 2 AND 3 WHICH $DISP=EN(22)
DISP1=1.0-DISP$ DO 620 J=13,19
DO 620 I=2,NIN
SO(2,J)=SO(2,J)+SI(I,J)*DISP1
SO(1,J)=SO(1,J)+SI(I,J)*DISP1 DO 61 U J=20, JJ
DO 61 U I=2, NIN
SI(1, J)=SI(1, J)+SI(I, J) SI(1,4)=SI(1,4)*SI(1,3)

DO 630 I=2,NIN

SI(1,3)=SI(1,3)+SI(1,3)

SI(1,4)=SI(1,4)+SI(1,4)*SI(1,3)

SI(1,6)=SI(1,6)+SI(1,6)

IF(SI(1,5).LI.SI(1,5)) SI(1,5)=SI(1,4)=SI(1,4)/SI(1,3) $SI(1,5)=SI(1,5)$

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CONTINUE CALCULATE EXIT HEAD
SO(1,5)=SI(1,5)-EN(5)
SO(2,5)=SI(1,5)-EN(5) $SO(3,5) = SU(1,5) - EN(5)$ $SO(1, 2) = PH$
 $SO(2, 2) = PH$
 $SO(3, 2) = PH$ $SO(1, 4) = SI(1, 4)$
 $SO(2, 4) = SI(1, 4)$
 $SO(3, 4) = SI(1, 4)$ CALCULATE SURFACE AREA
IF(EN(16).LE. 0.0) GO TO 730
AREA=EN(16)*EN(17)*EN(4) CO TO 720
AREA=3.142*EN(17)*EN(17)*EN(4)/4.0
CONTINUE 730
720
C CALCULATING RESIDENCE TIME TBAR (MINUTES) CALCULATING OVERFLOW RATE OR (IGPD/SQ.FT.)
OR=SI(1,3)*24.0/AREA CALCULATE THE THEORETICAL REMOVAL FROM BATCH TEST DATA
REMOVAL=1.0-EXP(-0.23*TBAR**0.57EN(18)**0.25) ACCOUNT FOR THE DECREASE IN EFFICIENCY DUE TO TURBULENCE REMOVAL=REMOVAL*EN(21) MATERIAL BALANCE

SO(2,13)=SO(2,13)+SI(1,13)*REMOVAL

SO(2,15)=SO(2,15)+SI(1,15)*REMOVAL

SO(2,16)=SO(2,16)+SI(1,16)*REMOVAL

SO(2,17)=SO(2,17)+SI(1,17)*REMOVAL

REMI=1.0-REMOVAL

SO(1,13)=SO(1,13)+SI(1,13)*REM1

SO(1,15)= CALCULATE UNDERFLOW FLOW
SO(2,6)=EN(25)*EN(23)*EN(24)*10.0
SSF=(SO(2,13)+SO(2,15)+SO(2,16)+SO(2,17))/SO(2,6)
ASSUME THAT THE MAXIMUM UNDERFLOW CONCENTRATION IS 60000 MG/L
IR(SSF:LE.0.06) GO TO 19
INSUFFICTENT WITHDRAWAL OF $1 \text{NSUFT}\left\{\text{LLEN1} \text{ W1} \right\} \text{HDRAWAL OF} \text{SLUDGE LEADS} + \text{SO(1,13)} = \text{SO(1,13)} + \text{SO(2,13)} * \{1,0-0.06/\text{SSF}}\right\} \text{SO(1,15)} = \text{SO(1,15)} + \text{SO(2,15)} * \{1,0-0.06/\text{SSF}}\right\} \text{SO(1,16)} = \text{SO(1,16)} + \text{SO(2,16)} * \{1,0-0.06/\text{SSF}}\right\} \text{SO(1,17)} = \text$ $\frac{1}{2}$ SET SCUM FLOW
SO(3,18)=SI(1,18)*0.9
SO(1,18)=SO(1,18)+SI(1,18)-SO(3,18) SO(2,18)=0.0
SO(2,18)=0.0
SO(3,6)=SO(3,18)*(1.0-EN(20)*1.0E-6)/EN(20)*2.2E6 EFFLUENT FLOW
SO(1,6)=SI(1,6)-SO(2,6)-SO(3,6) SET SCUM VARIABLES (OTHER THAN SCUM) SIMILAR TO EFFLUENT
F=S0(3,6)/(S0(1,6)+S0(3,6)) $50(3,13)=50(1,13)*F$
S0(1,13)=50(1,13)*F
S0(1,13)=50(1,13)-S0(3,13)
S0(3,15)=S0(1,15)*F
S0(1,15)=S0(1,15)-S0(3,15)

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 $SO(3,16)=SO(1,16)*F$
SO(1,16)=SO(1,16)-SO(3,16)
SO(3,17)=SO(1,17)*F
SO(1,17)=SO(1,17)-SO(3,17) COMPONENTS IN SOLUTION SPLIT ACCORDING TO FLOW CONFUNEINTS IN SOLUTION SPLIT AC

DO 600 I=20, JJ

SO(1,1)=SI(1,1)*SO(1,6)/SI(1,6)

SO(2,1)=SI(1,1)*SO(2,6)/SI(1,6)

SO(3,1)=SI(1,1)-SO(1,1)-SO(2,1)

CONTINUE 600 CONTINUE

SO(1,14)=SI(1,14)*SO(1,6)/SI(1,6)+SO(1,14)

SO(2,14)=SI(1,14)*SO(2,6)/SI(1,6)+SO(2,14)

SO(3,14)=SI(1,14)*SO(3,6)/SI(1,6)+SO(3,14)

SO(1,19)=SI(1,19)*SO(1,6)/SI(1,6)+SO(1,19)

SO(2,19)=SI(1,19)*SO(2,6)/SI(1,6)+S **SET** EXIT OXYGEN CONCENTRATIONS TO ZERO ALL $50(1, 7) = 0.0$
 $50(2, 7) = 0.0$
 $50(3, 7) = 0.0$ CALL BALNCE(2,1)
CALL BALNCE(2,2)
CALL BALNCE(2,3) IF(ISP) 1,1,2

CONTINUE

WRITE(6,300) REMOVAL

FORMAT(1H0,17HSOLIDS REMOVAL = ,F10,3)

WRITE(6,400) OR

FORMAT(1X,17HOVERFLOW RATE = ,F10,3,

WRITE(6,410) TBAR

WRITE(5,410) TBAR

WRITE(5,410) TBAR 300 400 $,F10.3.12H$ IGPD/SQFT 410 FORMAT(1X, I7HDETENTION TIME = $, F10, 3, 10H$ MINUTES λ CONTINUE

RETURN END

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SURROUTINE ACTSL1

LLST(50),NS(100),FN(100),SI(4,30),SO(4,30),KPRNT(10)
IS,NE,JJJLOOP,NIN,NOUT,MSN,ISP,NC,III,NCALC,NOCOMP,NSR COMMON COMMON MODE, NPLNT, LOOPC COMMON-COMMON SN(25+30) FEN(600) > NPOINT(25+2) SIMULATES THE ACTIVATED SLUDGE PROCESS CAN ALSO BE USED TO SIMULATE THE STEP AERATION PROCESS
CALLS THE BASIC CSTR PROGRAM FOR AEROBIC BIOLOGICAL REACTIONS EN VECTOR ********** STANDARD FORMAT
NUMBER OF REACTORS
TOTAL HEAD LOSS TH $1. - 15.$ IN PARALLEL $4.$ NUMBER OF REACTURS IN PARALLEL
TOTAL HEAD LOSS THROUGH ACT. SL. TANKS (FT OF WATER)
NUMBER OF INPUT STREAMS (= 2.0)
FIRST INPUT STREAM (= RETURN SLUDGE FLOW)
SECOND INPUT STREAM (= FEED STREAM)
NUMBER OF OUTPUT, STREAMS (= $5.$ $6 \bullet$ $7.$ $\frac{8}{11}$. STREAM NUMBER $\overline{12}$. OUTPUT NOTE EN(16.-32.) /
LENGTH OF REACTOR
WIDTH OF REACTOR
DEPTH OF REACTOR ARE USED BY CSTR2
R (FEET) $15:17$ (FFFT) $\overline{1}8.$ $(FEFT)$ _NOT`USING`A
IF USING A F
IE_ (K2_ZHR) A RÉSIDENCE TIME DISTRIBUTION MODEL
[RTD MODEL $\frac{1}{2}$ ⁰ $\frac{0}{34}$ $19.$ \equiv \equiv LYSIS RATE $20.$; RATE (K2 /HR)
- METAB. RATE (K1 /HR)
OF SOLUBILIZATION OF DEGRADABLE SOLIDS (
PING RATE CF VOLATILE ORGANICS (K5 /HR)
PING RATE CF VOLATILE ORGANICS (K5 /HR)
CRGANISMS YIELD FACTOR (Y DIMENSIONLESS)
FION OF LYSIS PRODUCT BASAL $21.$ **RATE** $(K3 / HR)$ STRIPPING RATE FRACTION OF
FRACTION OF
RATE OF AIR
UNIT_GROWTH DEGRADARLE CARBON
NONDEGRADABLE CARBON $29.$ FRACTION OF NZC RATIO IN RAW FEED
PZC PATIO IN RAW FEED
ABSORPTION NUMBER (FOR OXYGEN TRNSFER) $\overline{30}$. $\overline{31}$ $32-$ NOT PRESENTLY USED
NUMBER OF CSTR,S IN SERIES - EITHER AN RTD MODEL OR THE
ACTUAL NUMBER OF TANKS IN A STEP AERATION PROCESS $33.$ $34.$ MAXIMUM OF TEN TANKS IN SERIES TANK
FRACTION OF FEED STREAM TO FIRST TANK
FRACTION OF FEED STREAM TO SECOND TANK $\frac{35}{36}$ $37.$ ETC FRACTION OF FEED STREAM TO TENTH TANK (IF ANY) $44.$ $45.$ CAPACITY OF FACH BLOWER SCF/HR $46.$ $EN(27) = EN(45) * EN(46)/EN(4)/EN(34)$ $SIN=SI(1,11)+SI(2,11)$ STORE FEED STREAM IN SI(4,J)
DO 600 J=2,JJ
SI(4,J)=SI(2,J) CALCULATING THE VARIABLES FOR USE BY THE SECONDARY CLARIFIER
EM IS THE FOOD TO MICOORGANISM RATIO AND TAER IS THE AERATION TIME TANKS=FN(34)
IF(EN(19).GT_1.0) TANKS=1.0 TAER=FN(16)*EN(17)*EN(18)*TANKS *6.24/(SI(1,3)+SI(2,3))*EN(4)
TM=TAER/EN(34) FM=SI(2,10)/SI(1,12)/TAER*24.0

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 $\frac{c}{c}$ $NCSTR=FN(34)+0.01$ \overline{C} DO 610 IJ=1, NCSTR
IF(EN(34+IJ).LE. 0.0) GO TO 20 C SI(2,3)=SI(4,3)*EN(34+IJ)
DO 900 KB=6,JJ
SI(2,KB)=SI(4,KB)*EN(34+IJ) 900 CALL MIXERI EN(29)=EN(29)*SI(1,11)/SO(1,11)
DO 910 KB=2,JJ
SI(1,KB)=SO(1,KB) $\begin{array}{c} 910 \\ 20 \\ C \end{array}$ CONTINUE CALL CSTR2 C DQ 620 J=2, JJ
SI(1, J)=50(1, J) $\begin{array}{c} 620 \\ 610 \\ C \\ C \end{array}$ CONTINUE $SO(1,5) = S_1(2,5) - EN(5)$ $DELS = SO(1, 11) - S1N$

RETURN
END

SUBROUTINE CSTR2

 $\begin{matrix} 1 \\ 2 \end{matrix}$

 $rac{c}{c}$

LLST(50),NS(100),EN(100),SI(4,30),SO(4,30),KPRNT(10)
IS,NE,JJ,LOOP,NIN,NOUT,MSN,ISP,NC,III,NCALC,NOCOMP,NSR COMMON **COMMON** MODE, NPLNT, LOOPC COMMON SN(25,30), EEN(600), NPOINT(25,2) THIS IS THE BASIC CSTR ROUTINE TO DESCRIBE
BIOLOGICAL PROCESSES AND IS CALLED BY ACTSL
WRITTEN BY P. TAN THE AEROBIC EN VECTOR ********* ***

NOTE-CSTR2 USES ONLY EN(16.-32.)

EN(I.-15.) AND EN(33.-) ARE USED BY ACTSL1

NOTE-CSTR2 HAS ONLY ONE INPUT STREAM, THE MIXED LIQUOR FLOW

LENGTH OF REACTOR (FEET)

DEPTH OF REACTOR (FEET)

DEPTH OF REACTOR (FEET)

= $\frac{16}{17}$.
18. $\overline{1}\overline{9}$. 2222222222223332
223456789012 FRATE OF AIR ADDITION TO EACH REACTOR (SCF/HR)

RATE OF AIR ADDITION TO EACH REACTOR (SCF/HR)

INIT RATE GROWTH CONSTANT (K4 /HR)

FRACTION OF MLSS THAT IS NITRIFIERS

N/C RATIO IN RAW FEED

P/C RATIO IN RAW FEED

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 $\frac{d}{dt} = \frac{1}{2} \frac{d^2 \phi}{dt^2}$

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LISTING OF THE SECONDARY SETTLING TANK MODEL

SUBROUTINE SECLAR1

COMMON
COMMON LLST(50), NS(100), EN(100), SI(4, 30), SO(4, 30), KPRNT(10)
IS, NE, JJ, LOOP, NIN, NOUT, MSN, ISP, NC, III, NCALC, NOCOMP, NSR MODE , NPL NT , LOOPC
SN(25 , 30) , EEN(600) , NPOINT(25 , 2)
/A/ TAER , TM , FM , DELS COMMON **COMMON** WRITTEN BY P. TAN
FOR WWTP SIMULATION
FOR REVISED STREAM LIST 2/9/71 SIMULATES A SECONDARY CLARIFIER EN VECTOR
********** STANDARD FORMAT
NUMBER OF CLARIFIERS IN PARALLEL
HEAD LOSS THROUGH CLARIFIER (FEET
FIRST OUT<u>PUT SIREAM,</u> (FFELUENT) $1 - 15$. -4513 OF WATER) FIRST OUTPUT STREAM (EFFLUENI)
SECOND OUTPUT STREAM (UNDERFLOW)
THIRD OUTPUT STREAM (SCUM TROUGH FLOW)
MODE OF OPERATION
= 1.0 FOR FIXED FRACTION OF RECYCLE
= 2.0 FOR MAXIMUM CONCENTRATION OF UNDERFLOW
FRACTION OF RECYCLE $16.$ $\frac{17}{19}$: - SQ.FT. $\frac{20}{21}$
 $\frac{21}{22}$ NUMBER OF UNDERFLOW SLUDGE PUMPS
CAPACITY OF PUMPS IGPH EACH
FRACTION OF CAPACITY USED $EN(17) = EN(2U)*EN(21)*EN(22)/SI(1,3)$ $SO(1, 2) = S[(1, 2)$
 $SO(2, 2) = S[(1, 2)$
 $SO(3, 2) = S[(1, 2)$
 $SO(1, 4) = S[(1, 4)$
 $SO(2, 4) = S[(1, 4)$
 $SO(3, 4) = S[(1, 4)]$ CALCULATE EXIT HEADS
SO(1,5)=SI(1,5)-EN(5)
SO(2,5)=SI(1,5)-EN(5)
SO(3,5)=SI(1,5)-EN(5) CALCULATING THE SLUDGE VOLUME INDEX
SVI=56.1+113.J*FM*(1.05**(SI(1,4)-20.0)) CALCULATING THE MAXIMUM UNDERFLOW CONCENTRATION CU $CU=1.0E+06/SVI$ CALCULATING THE MIXED LIQUOR SUSPENDED SOLIDS XMLSS
XMLSS=SI(1,11)/SI(1,6)*1.0E6 METHOD=EN(16)+0.001
GO TO (1,2),METHOD CONTINUE

FOR FIXED FRACTION OF RECYCLE

SO(2,6)=SI(1,6)*EN(17)

CALCULATING THE OVERFLOW RATE GSS (USGPD/SQFT)

CALCULATING THE OVERFLOW RATE GSS (USGPD/SQFT)

GSS=SO(1,6)*2,879/EN(18)/EN(4)

THE EXTRAPOLATION OF BATCH SE CONTINUE **TANKS**

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 267 SO(2,11)=SI(1,11)-SO(1,11)
UNDERFLOW CONCENTRATION CANNOT BE GREATER THAN CU
IF((SO(2,11)/SO(2,6)*1.0E6) .LT. CU) GO TO 10
SO(2,11)=CU*1.0E-6*SO(2,6)
SO(1,11)=SI(1,11)-SO(2,11) C C
CC
C CONTINUE
FOR MAXIMUM CONCENTRATION OF UNDERFLOW
ROUGH ESTIMATE OF UNDERFLOW FLOW
SO(2,6)=SI(1,1)*1,0E6/CU SO(1,6)=SI(1,6)-SO(2,6)

CALCULATING THE OVERFLOW RATE GSS (USGPD/SQFT)

GSS=SO(1,6)*2,879/EN(18)/EN(4)

APPLY CORRELATION

XRSS=556,0*(GSS**0,494)/(XMLSS**1,82)/(TAER**0,439)*1.5

SO(1,11)=SI(1,11)*XRSS

SO(2,11)=SI(1,11) \overline{C} \overline{C} CALCULATING CORRECTED UNDERFLOW FLOW
SO(2,6)=SO(2,11)*1,0E6/CU
SO(1,6)=SI(1,6)-SO(2,6) \subset C
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c
 CONTINUE ASSUME THAT THE FLOCS ARE UNIFORM IN COMPOSITION
ALSO ASSUME THAT THE OVERFLOW AND UNDERFLOW SOLIDS HAVE THE
SAME COMPOSITION AS THE INCOMING FLOCS
PI=SO(1,111)
DO 600 I=13,19, ... SO (1, 1) =P1*S1(1, 1)
SO (2, 1) =S1(1, 1) –SO (1, 1)
CONTINUE $\frac{600}{5}$ SPLIT OVERFLOW INTO EFFLUENT AND SCUM FLOW
SO(3,18)=SI(1,18) SO(3,18)=SI(1,18)

SO(1,18)=0.0

SO(2,18)=0.0

SO(3,6)=SO(3,18)*(1.0–EN(19)*1.0E–6)/EN(19)*2.2E6

SO(1,6)=SO(1,6)–SO(3,6)

P1=SO(3,6)/(SO(1,6)+SO(3,6))

DO 610 1=13,17

SO(3,1)=SO(1,1)+P1

SO(1,1)=SO(1,1)-SO(3,1)

CONTINU CONTINUE
SO(3,19)=SO(1,19)*P1
SO(1,19)=SO(1,19)-SO(3,19) 610 $\frac{c}{c}$ SPLIT SOLUBLE COMPONENTS ACCORDING TO FLOW
P1=S0(1,6)/SI(1,6)
P2=S0(2,6)/SI(1,6) $\begin{array}{ll}\n\text{D0} & 620^\circ \text{I} = 20, \text{J} \text{J} \\
\text{S0}(1, 1) = \text{S1}(1, 1) * \text{P1} \\
\text{S0}(2, 1) = \text{S1}(1, 1) * \text{P2} \\
\text{S0}(3, 1) = \text{S1}(1, 1) - \text{S0}(1, 1) - \text{S0}(2, 1)\n\end{array}$.620
C
C CONTINUE SET ALL EXIT OXYGEN CONCENTRATIONS AT ZERO
SO(1,7)=0.0
SO(2,7)=0.0
SO(3,7)=0.0 C CALL BALNCE(2,1)
CALL BALNCE(2,2)
CALL BALNCE(2,3) $\frac{c}{c}$ IF(ISP) 30,30,31
WRITE(6,300) TAER
FORMAT(1HU,22HAERATION TIME
WRITE(6,301) FM
FORMAT(1X,22HFOOD TO MICRO RATIO =
WRITE(6,302) GSS \bullet $\frac{31}{300}$ $5F10 - 357H$ HOURS) 301 $, F10.3)$

 302 303
30
C
C

CONTINUE

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LISTING OF THE ANAEROBIC DIGESTER MODEL

FORMAT(1X,22HOVERFLOW RATE =
WRITE(6,303) SVI
FORMAT(1X,22HSLUDGE VOLUME INDEX =

SUBROUTINE ANDIG1

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COMMON LLST(50),NS(100),EN(100),SI(4,30),SO(4,30),KPRNT(10)
COMMON IS,NE,JJ,LOOP,NIN,NOUT,MSN,ISP,NC,III,NCALC,NOCOMP,NSR
COMMON MODE,NPLNT,LOOPC
COMMON SN(25,30),EEN(600),NPOINT(25,2) SIMULATES AN ANAEROBIC DIGESTER
WRITTEN BY P. TAN AUG/71 EN VECTOR
********* STANDARD FORMAT
NUMBER OF DIGESTERS IN PARALLEL
HEAD LOSS IN DIGESTERS (FT OF WATER)
DIAMETER OF DIGESTER (FEET)
DEPTH OF DIGESTER (FEET) $1 - -15$ $4\bullet$ $5.$ $\frac{16}{17}$. DEPTH OF DIGESTER (FEET)

= 1.0 FOR CONVENTIONAL DIGESTER OPERATION

= 2.0 FOR HIGH RATE DIGESTER OPERATION

TEMPERATURE OF OPERATION

FRACTION OF WATER TO FIRST OUTPUT STREAM (SUPERNATANT)

= 1.0 IF FIRST STAGE OF A TWO-S $18.$ $19.$ $\bar{2}0$. $21 22 \bullet$ $23 \cdot 24$ USING SO(4,JJ) AS TEMPORARY STORAGE OF INTERMEDIATE RESULTS DO 600 IK=2, JJ
SO(4, IK)=SI(1, IK)
CONTINUE $SO(1, 4) = EN(19)$
 $SO(2, 4) = EN(19)$ $SO(4,4) = EN(19)$
SO(1,5)=SI(1,5)-EN(5)
SO(2,5)=SI(1,5)-EN(5)
SO(4,5)=SI(1,5)-EN(5) CALCULATING VOLUME OF DIGESTER (CU FT) VOL=3.142*EN(16)*EN(16)*EN(17)/4.0*EN(4) CALCULATING DIGESTER RESIDENCE TIME (DAYS)
RESTIME=VOL*6.24/SI(1,3)/24.0 CALCULATING VOLATILE SOLIDS LOADING ON DIGESTER LB/CUFT/DAY
SSLOAD=SI(1,12)*24.0/VOL ALKALINITY CALCULATIONS
ALK=20.0*SI(1,23)/SI(1,6)

USGPD/SQFT)

 $5F10.3912H$

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 $\label{eq:1} \frac{1}{\sqrt{2}}\sum_{i=1}^{n-1}\frac{1}{\sqrt{2}}\sum_{i=1}^{n-1}\frac{1}{\sqrt{2}}\sum_{i=1}^{n-1}\frac{1}{\sqrt{2}}\sum_{i=1}^{n-1}\frac{1}{\sqrt{2}}\sum_{i=1}^{n-1}\frac{1}{\sqrt{2}}\sum_{i=1}^{n-1}\frac{1}{\sqrt{2}}\sum_{i=1}^{n-1}\frac{1}{\sqrt{2}}\sum_{i=1}^{n-1}\frac{1}{\sqrt{2}}\sum_{i=1}^{n-1}\frac{1}{\sqrt{2}}\sum_{i=1}^{n-1}\frac{$

 $\label{eq:2.1} \frac{1}{\sqrt{2}}\int_{0}^{\infty}\frac{1}{\sqrt{2}}\left(\frac{1}{\sqrt{2}}\right)^{2}d\mu\int_{0}^{\infty}\frac{1}{\sqrt{2}}\left(\frac{1}{\sqrt{2}}\right)^{2}d\mu\int_{0}^{\infty}\frac{1}{\sqrt{2}}\left(\frac{1}{\sqrt{2}}\right)^{2}d\mu\int_{0}^{\infty}\frac{1}{\sqrt{2}}\left(\frac{1}{\sqrt{2}}\right)^{2}d\mu\int_{0}^{\infty}\frac{1}{\sqrt{2}}\left(\frac{1}{\sqrt{2}}\right)^{2}d\mu\$

LISTING OF THE TRICKLING FILTER MODEL

SUBROUTINE TRICK1

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COMMON LLST(50),NS(100),EN(100),SI(4,30),SO(4,30),KPRNT(10)<br>COMMON IS,NE,JJ,LOOP,NIN,NOUT,MSN,ISP,NC,III,NCALC,NOCOMP,NSR<br>COMMON MODE,NPLNT,LOOPC<br>COMMON SN(25,30),EEN(600),NPOINT(25,2)
  SIMULATES A TRICKLING FILTER<br>UTILIZES THE EQUATION PROPOSED BY ECKENFELDER<br>WRITTEN BY P. TAN SEPT/71
  EN VECTOR<br>*********
  1.-15. STANDARD FORMAT<br>4. NUMBER OF TRICKLING FILTERS IN PARALLEL<br>5. HEAD LOSS IN FILTER (FT OF WATER - GREATER THAN HEIGHT)<br>16. DEPTH OF FILTER (FEET)<br>17. DIAMETER OF FILTER (FEET)
  \frac{5}{16}.
                  REACTION<br>EXPONENT<br>EXPONENT
                                       CONSTANT<br>ON DEPTH<br>ON LIQUID LOADING RATE
   \overline{18}.
  \frac{1}{2}.
  DO 600 J=2, JJ<br>SO(1, J)=SI(1, J)<br>SO(1, 5)=SI(1, 5)-EN(5)
  CALCULATE SURFACE AREA<br>AREA=3.142*EN(17)*EN(17)/4.0*EN(4)
  CALCULATE THE LIQUID LOADING RATE (USGPD/SQFT)<br>Q=SI(1,3)*28.79/AREA
  CALCULATE REMOVAL OF BOD ACCORDING TO EQUATION<br>REM = FRACTION OF DEGRADABLE CARBON REMAINING<br>REM=EXP(-EN(18)*EN(16)**EN(19)/Q**EN(20))
  ASSUME THAT THE REMOV.<br>SO(1,16)=SI(1,16)*REM<br>DO 610 J=18,22
                              THE REMOVAL APLLIES TO ALL DEGRADABLE COMPONENTS
  SO(I, J) = SI(I, J) * REMASSUME MICROORGANISM YIELD FACTOR IS 0.54<br>SO(1,15)=0.54*(SI(1,16)+SI(1,18)+SI(1,19)+SI(1,20)+SI(1,21)<br>1 +SI(1,22))*(1.0-REM)
  CALL BALNCE (2, 1)RETURN.
  END
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LISTING OF THE SCREEN MODEL 272

SUBROUTINE SCREEN1

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COMMON LLST!50l,NS(100),ENll00),SI(4,30),S0(4,30),KPRNT(l0). COMMON IS,NE,JJ,LOOP,NIN,NOUT,MSN,ISP,NC'III'NCALC,NOCOMP,NSR COMMON MODE, NPLNT, LOOPC COMMON SNC25,3U),EENC600},NPOINT!25,2) EN VECTOR
********* *********
1.-15. STANDARD FORMAT 4. = SCREENINGS, IF KNOWN FROM PLANT OPERATIONS <CUFT/MIG> = 0.0 IF USING BARMINUTER = ANY NEGATIVE NUMBER IF WANT TO USE INTERNAL SCREENINGS ESTIMATE
5. HEAD LOSS ACROSS SCREEN (FT OF WATER) 16. SCREEN SIZE, IF USING INTERNAL ESTIMATE OF SCREENINGS <IN) DO 600 J=2,<mark>Jl,J</mark>)
DO 600 J=2,JJ
DO 600 J=2,JJ $SO(1,5)=S_1(1,5)-EN(5)$ CALCULATE VOLUME OF SCREENINGS PER DAY $IF(EN(4))1, 2, 3$ SCR=19.0*10•0**1-0.8*EN(16)) GO_TO_4 RETURN $SCR=EN(4)$ VOL=SI(1,3l*SCR*24.0*1•0E-6 ESTIMATED DRY WEIGHT OF SCREENINGS CALCULATING MASS OF SCREENINGS PER WRAGS=30.U*VOL/24.0 CONVERT TO EOUIV CARBON WRAGS=WRAGS72.1
SO(1,16)=SI(1,16)-WRAGS CALL BALNCE $(2, 1)$ IF< ISP} 10,}0,11 WRITE(6,400) VOL
FORMAT!!IHU,13HSCREENINGS = **CONTINUE** RETURN END IS 30 LBS/CUFT HOUR 'F8.2,10H CUFT/DAY)

LISTING OF THE GRIT REMOVAL MODEL

SUBROUTINE GRIT1

COMMON LLST(50),NS(100),EN(100),SI(4,30),SO(4,30),KPRNT(10)
COMMON IS,NE,JJ,LOOP,NIN,NOUT,MSN,ISP,NC,III,NCALC,NOCOMP,NSR
COMMON MODE,NPLNT,LOOPC
COMMON SN(25,30),EEN(600),NPOINT(25,2)

WRITTEN BY P. TAN 27/8/71
DEGRITTER FOR SEWAGE TREATMENT
SIMPLE MODEL USING THE AVERAGE AMOUNT OF GRIT COLLECTED /MIG

EN VECTOR **********

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^STANDARD FORMAT
CU.FT. OF GRIT PER MILLION GALLONS OF RAW FEED
HEAD LOSS THROUGH CHAMBER (FT OF WATER) $1 - 15 \frac{4}{5}$.

DO 600 J=2, JJ
SO(1, J)=SI(1, J)
SO(1, 5)=SI(1, 5)-EN(5) 600 $CUFT = S1(1,3) * EN(4) * 1.0E - 6$ GRIT=CUFT*0.40*62.4*2.65
SO(1,11)=SI(1,11)-GRIT
SO(1,13)=SI(1,13)-GRIT IF (ISP) 1,1,2
CUFT=CUFT*24.0
WRITE(6,400) CUFT
FORMAT(1H0,25HVOLUME OF GRIT REMOVED =
CONTINUE 400 CU FT/DAY $5F8.2511H$ λ

RETURN
END
LISTING OF THE CHLORINATOR MODEL

SUBROUTINE CHLOR1

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COMMON LLST(50),NS(100),EN(100),SI(4,30),SO(4,30),KPRNT(10)<br>COMMON IS,NE,JJ,LOOP,NIN,NOUT,MSN,ISP,NC,III,NCALC,NOCOMP,NSR<br>COMMON MODE,NPLNT,LOOPC<br>COMMON SN(25,30),EEN(600),NPOINT(25,2)
 EN VECTOR
 *********
                   ***<br>STANDARD FORMAT<br>DISPOSITION OF WASTE<br>= 1.0 FOR RAW SEWAGE<br>= 3.0 FOR SETTLED RAW SEWAGE<br>= 4.0 FOR SETTLED RAW SEWAGE<br>= 4.0 FOR SEPTIC SETTLED RAW SEWAGE<br>= 5.0 FOR SAND FILTER EFFLUENT<br>= 6.0 FOR SAND FILTER EFFLUENT<br>USED
 1 - 15\bar{4}.
5.CL(1)=10.0<br>CL(2)=20.0<br>CL(3)=8.0<br>CL(4)=16.0<br>CL(5)=6.0
CL(6) = 3.0DO 600 J=2, JJ<br>SO(1, J)=SI(1, J)
I = EN(4)+0.01<br>CL2=SI(1,3)*24.0*10.0*CL(I)*1.0E-6<br>EN(5)=CL2
IF(ISP) 10,10,11<br>WRITE(6,400) CL2
FORMAT(1HO,17HCHLORINE USAGE =
                                                                                      5F8 - 2510HLBS/DAY
                                                                                                                                               \lambdaCONTINUE
RETURN<br>END
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LISTING OF THE MIXER-SPLITTER MODEL

SUBROUTINE MIXERl

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COMMON LLST(50),NS(100),EN(100),SI(4,30),SO(4,30),KPRNT(10)
COMMON IS,NE,JJ,LOOP,NIN,NOUT,MSN,ISP,NC,III,NCALC,NOCOMP,NSR
COMMON MODE,NPLNT,LOOPC
COMMON SN(25,30),EEN(600),NPOINT(25,2) EN VECTOR
1,-15. STANDARD FORMAT 16. FRACTION OF FLOW TO FIRST OUTPUT STREAM $\overline{17}$ $\overline{17}$ $\overline{17}$ SŌ(VONÍĂ))=21(J)
180 - Pan (J=3)
180 - Pan (J) GO 10 30
180 - Pan (Sili) 650 GO TO 40 CONTINUE CALL ALKMIX<PHl SUM OF INPUT STREAMS IS TEMPORARILY STORED IN SO(NOUT,J) DO 600 J=6,JJ SO<NOUT,Jl=O. DO 600 I=l,NIN SO<NOUT,Jl=SO(NOUT,Jl+SI(I,Jl 600 SO(NOUT,2)=PH
SO(NOUT,3)=0.0 SO(NOUT,4)=0.J SO(NOUT,5)=SI(1,5) \overline{D} ^O 61^C I=1, NIN $SO(NOUI-3)=SO(NOUI-3)+SI(1-3)$ $SO(NOUT,4)=SO(NOUT,4)+SU(1,3)*SU(1,4)$ IF(SI(I,5).LT.SO(NOUT,5)) SO(NOUT,5)=SI(I,5)
CONTINUE 610 SO(NOUT,4l=SO(NOUT,4l/SO(NOUT,3l CONTINUE C OUTPUT IF(NOUT.EO.ll RETURN DO 620 1=1,NOUT
SO(1,2)=SO(NOUT,2) $SO(1,3) = SO(NOUI,3) * EN(1+15)$ SO(I,4l=SO(NOUT,4) SO(I•5)=SO(NOUT•5) DO 620 J=6,JJ SO(J,J)=SO(NOUT,Jl*EN(I+l5) 620 c CONTINUE RETURN END

SUBROUTINE SEPA01

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COMMON LLST(50),NS(100),EN(100),SI(4,30),SO(4,30),KPRNT(10)<br>COMMON IS,NE,JJ,LOOP,NIN,NOUT,MSN,ISP,NC,III,NCALC,NOCOMP,NSR<br>COMMON MODE,NPLNT,LOOPC<br>COMMON SN(25,30),EEN(600),NPOINT(25,2)
                    EN VECTOR<br>*********
                    1.-15. STANDARD FORMAT<br>4. FRACTION OF FLOW TO FIRST OUTPUT STREAM (SUPERNATE)<br>5. FRACTION OF SOLIDS TO FIRST OUTPUT STREAM (SUPERNATE)
                   SO(1, 2) = S[(1, 2)]<br>
SO(2, 2) = S[(1, 2)]<br>
SO(1, 4) = S[(1, 4)]<br>
SO(2, 4) = S[(1, 4)]<br>
SO(1, 5) = S[(1, 5)]<br>
SO(2, 5) = S[(1, 5)]IF(SI(1,6).GT. 0.0) GO TO 10<br>DO 650 J=6,JJ<br>SO(1,J)=0.0<br>SO(2,J)=0.0<br>CONTINUE
 650
                   SO(1,3)=0.0<br>SO(2,3)=0.0<br>RETURN
                   CONTINUE
                   SO(1,6) = EN(4)*SI(1,3)*10.0<br>SO(2,6) = SI(1,6) - SO(1,6)SEPARATE SOLIDS<br>DO 600 J=13,19<br>SO(1,J)=SI(1,J)*EN(5)<br>SO(2,J)=SI(1,J)-SO(1,J)<br>CONTINUE
\frac{6}{5}^{00}SOLUBLE COMPONENTS ARE SPLIT ACCORDING TO FLOW<br>P1=SO(1,6)/SI(1,6)<br>DO 610 J=20,JJ<br>SO(2,J)=SI(1,J)*P1<br>SO(2,J)=SI(1,J)-SO(1,J)
6^{10}CONTINUE
                   CALL BALNCE(2,1)<br>CALL BALNCE(2,2)
                   RETURN<br>END
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LISTING OF THE PROGRAM TO CONTROL THE MLSS IN THE AERATION TANKS

SUBROUTINE CONTO2

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COMMON LLST(50),NS(100),EN(100),SI(4,30),SO(4,30),KPRNT(10)<br>COMMON IS,NE,JJ,LOOP,NIN,NOUT,MSN,ISP,NC,III,NCALC,NOCOMP,NSR<br>COMMON MODE,NPLNT,LOOPC<br>COMMON SN(25,30),EEN(600),NPOINT(25,2)<br>COMMON /A/ TAER,TM,FM,DELS<br>DATA DELS/
   WRITTEN BY P. TAN<br>FOR WWTP SIMULATION
   CONTROLS THE MLSS IN THE AERATION TANKS
   EN VECTOR<br>*********
                        ***<br>STANDARD FORMAT<br>FEED STREAM TO AERATION TANKS (EXCLUDING RECYCLE FLOW)<br>MLSS REQUIRED IN AERATION TANKS<br>FIRST OUTPUT STREAM (RECYCLE STREAM)<br>SECOND RECYCLE STREAM (WASTE SLUDGE FLOW)
   1 - 154.\frac{5}{12}.
   S = EN(4)<br>CALL STREAM(S)
CALCULATE FRACTION OF INCOMING FLOW TO RECYCLE - A<br>A=(EN(5)*1.0E-6*SN(IS,6)-SN(IS,11)-DELS)/(SI(1,11)-EN(5)*SI(1,6)*<br>1 1.0E-6)
  IF(A.IT.0.0) A=0.0<br>IF(A.GT.1.0) A=1.0<br>DO 600 I=3, JJ<br>SO(1, I)=SI(1, I)*A<br>SO(2, I)=SI(1, I)-SO(1, I)
  SO(1, 2) = S[(1, 2)<br>
SO(2, 2) = S[(1, 2)<br>
SO(1, 4) = S[(1, 4)<br>
SO(2, 4) = S[(1, 4)<br>
SO(2, 5) = S[(1, 5)<br>
SO(2, 5) = S[(1, 5)<br>
SO(1, 7) = 0.0<br>
SO(2, 7) = 0.0RETURN
  END
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LISTING OF SUPPORTING MODELS TO HANDLE STREAM VARIABLES 278

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SUBROUTINE CONVERTIM, KIN, JIN, KOUT, JOUT)

630 c 640 COMMON LLST(50),NS(100),EN(100),SI(4,30),SO(4,30),KPRNT(10)
COMMON IS,NE,JJ,LOOP,NIN,NOUT,MSN,ISP,NC,III,NCALC,NOCOMP,NSR
COMMON MODE,NPLNT,LOOPC
COMMON SN(25,3G),EEN(600),NPOINT(25,2)
DIMENSION V(30) CONVERTS STREAM VARIABLES FROM POUNDS PER HOUR TO PPM OR VICE VERSA INPUT VARIABLES
M = 1 FOR CON
M = 2 LEOR CON M = 1 FOR CONVERTING M = 2 FOR CONVERTING KIN = VECTOR TO BE READ = 1 FOR SN VECTOR *=* 2 FOR SO VECTOR = 3 FOR SI VECTOR JIN = ROW NUMBER OF INPUT VECTOR KOUT = VECTOR TO BE WRITTEN ON TO = 1 FOR SN VECTOR = 2 FOR SO VECTOR
=3 FOR SI VECTOR JOUT = ROW NUMBER OF OUTPUT VECTOR VARIABLES 2. TO 6. ARE NOT CHANGED READ FROM INPUT VECTOR GO TO C1,2,3J,KIN !)() 600 J=2,JJ V(Jl=SN<JIN,Jl GO TO 10
DO 610 J=2,JJ
V(J)=SO(JIN,J) GO TO 10
DO 620 J=2,JJ
V(J)=SI(JIN,J) CONTINUE IFIV(6l.GT. u.O l GO TO 40 V(3)=0.0
DO 700 J=6,JJ
V(J<u>)</u>=U.O $V(J) = U$. \tilde{Q}
GO TO 11 **CONTINUE** CONVERSION GO TO $(4,5)$, M CONTINUE CONVERTING FROM LBS/HR TO PPM PPM TO LBS/HR FROM SOLUBLE COMPONENTS
V(7)=V(7)/V(6)*l•OE6 $V(9) = V(9)/V(6)*1.0E6$ 00 630 J=20,JJ
V(J)=V(J)/V(6)*1.0E6 PARTICULATE COMPONENTS V(8)=V(8)/V(3)*1.0E5
DO 640 J=10,19 VIJI=VIJJ*7*VI31*1.OE5 GO TO 11 CONTINUE CONVERTING-FROM PPM TO LBS/HR SOLUBLE COMPONENTS Vl7l=V(7l*V(6l*l·OE-6 $V(9) = V(9) * V(6) * 1$. $0E-6$ LBS/HR TO PPM

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DO 650 J=20,JJ

 $V(J)=V(J)*V(6)*1.0E-6$ $\begin{matrix} 650 \\ C \end{matrix}$ C PARTICULATE COMPONENTS V(8l=V(8l*V(3l*1.0E-5 $0.660, J=10, 19$ 660 V(J)=V(J)*V(3)*1.0E-5 č. **CONTINUE** WRITING ON TO OUTPUT VECTOR c GO TO (6,7,8),KOUT c 6 DO 670 J=2,JJ
670 SN(JOUT,J)=V(J) GO TO 12 7 DO 680 J=2,JJ 680 SO(JOUT,Jl=V(J) GO TO 12

8 DO 690 J=2,JJ

690 SI(JOUT,J)=V(J)
C **CONTINUE** 8
690
2
2
2 RETURN END

SUBROUTINE ALKMIX(PH) COMMON LLST(50),NS(100),EN(100),SI(4,30),SO(4,30),KPRNT(10)
COMMON IS,NE,JJ,LOOP,NIN,NOUT,MSN,ISP,NC,III,NCALC,NOCOMP,NSR
COMMON MODE,NPLNT,LOOPC
COMMON SN(25,30),EEN(600),NPOINT(25,2) SUBROUTINE TO CALCULATE THE RESULTING PH WHEN STREAMS ARE MIXED $SUMALK=0.0$ SUMCN=0.0 SUMCI=0.0

DO 600 I=1,NIN

IF(SI(I,6).LE.0.0) GO TO 600

IF(SI(I,2).LE.0.0) SI(I,2)=7.0

ALK=20.0*SI(I,23)/SI(I,6)

CN=71.43*SI(I,27)/SI(I,6)

PH=SI(I,2) CALL ALKPH(1,ALK,CT,CN,PH) SUMCT=SUMCT+CT
SUMCN=SUMCN+CN CONTINUE CALL ALKPH (2, SUMALK, SUMCT, SUMCN, APH) $PH=APH$ RETURN END
SUBROUT INE ALKPH(NZ,ALK,CT,CN,PH) DIMENSION XCOF(4) SUBROUTINE TO CALCULATE CT OR PH GIVEN THE OTHER THREE VARIABLES $PK1=6.3$ PK2=10.3
PKN=9.3
GO TO (1,2), NZ CALCULATE CT CONTINUE
CONTINUE
CT=(ALK-CN/(1.0+10.0**(PKN-PH)))*(10.0**(PK1-PH)+1.0+10.0**
1(PH-PK2))/(1.0+10.0**(PH-PK2)*2.0) **RETURN** CALCULATE PH
CONTINUE CONTINUE

Al=ALK-CN

A2=ALK-CN-CT

A3=ALK-CN-CT

A5=ALK-CN-2.0*CT

A5=ALK-CN-2.0*CT

XCOF(1)=10.0**(-PK1-PK2-PKN)*A5

XCOF(2)=(A3*10.0**(-PK1-PKN) + A4*10.0**(-PK1-PK2))

XCOF(2)=(A3*10.0**(-PKN) + A2*10.0**(-PK1))

XCOF(4 $XCOF(4) = ALK$

CALL BISECT(XCOF,H)

 $PH=-ALOGIO(H)$

RETURN END

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SUBROUTINE BISECT(XCOF, H)

DIMENSION XCOF(4)

```
F(XCOF,H) = XCOF(1) + H*(XCOF(2) + H*(XCOF(3) + H*XCOF(4)))
```
 $HL = 1.0E - 5$
 $HR = 1.0E - 9$

IF(F(XCOF,HL)*F(XCOF,HR)) 10,20,30
H=SQRT(HL*HR)
IF(F(XCOF,HL)*F(XCOF,H).LT. 0.0) GO TO 2
HL=H
GO TO 4
HR=Has((H) HD)(H) CT 0.01) CO TO TO

- IF(ABS((HL-HR)/HL).GT.0.01) GO TO 10
- H=HL
RETURN

H=HR
IF(F(XCOF,HL).EQ. 0.0) H=HL
RETURN

$\frac{c}{30}$

 $\frac{c}{c}$

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 $\begin{array}{c} 2 \\ 4 \end{array}$

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H=HR
IF(ABS(F(XCOF,HL)).LT. ABS(F(XCOF,HR))) H=HL

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RETURN<br>END
```
LISTING OF THE REPORT GENERATOR

SUBROUTINE REPTO1 LLST(50)\$NS(100),EN(100),SI(4,30),SO(4,30),KPRNT(10)
IS,NE,JJ,LOOP,NIN,NOUT,MSN,ISP,NC,III,NCALC,NOCOMP,NSR
MODE,NPLNT,LOOPC
SN(25,30),EEN(600),NPOINT(25,2) COMMON
COMMON
COMMON COMMON WRITTEN BY P. TAN
FOR WWTP SIMULATION REPORTS ON THE VARIOUS STREAMS IN THE PLANT FORMAT
LENGTH 35 $=$ $=0.0$ RAW FEED TO PLANT (STREAM NUMBER)
INPUT TO PRIMARY CLARIFIER
EFFLUENT OF PRIMARY CLARIFIER
UNDERFLOW OF PRIMARY CLARIFIER $EN(17)$ \equiv EN(18)
EN(19)
EN(20)
EN(22)
EN(22) \equiv \equiv INPUT TO AERATION TANKS
OUTPUT OF AERATION TANKS
INPUT TO SECONDARY CLARIFIER \Rightarrow \equiv INPUT TO SECONDARY CLARIFIER
EFFLUENT OF SECONDARY CLARIFIER
UNDERFLOW OF SECONDARY CLARIFIER
RETURN ACTIVATED SLUDGE
WASTE ACTIVATED SLUDGE
INPUT TO DIGESTERS
DIGESTER SUPERNATANT
DIGESTED SLUDGE
INPUT TO TRICKENER
THICKE \equiv $=$ $=$ \equiv \equiv \equiv $=$ \equiv \equiv \equiv \equiv \equiv \equiv $EN(35)$ \equiv SET STREAM NUMBER TO 0.0 IF DO NOT WISH TO REPORT ON STREAM IF (MODE.EQ. 0) RETURN WRITE(6,79)
WRITE(6,80)
WRITE(6,82)
WRITE(6,80)
WRITE(6,85) LOOPC ÷ DO 50 I=1,20
S=EN(I+15)
IF(S.LE.0.0)
CALL STREAM(S)
IT=15 GO TO 50 CALL CONVERT (1,1,17,3,1)
SI(1,1)=S
IW=29+I WRITE(6,IW) SI(1,1),SI(1,3),(SI(1,J),J=7,12) CONTINUE
WRITE(6,79) $MODE = 0$, F5.0, 7F10.2)
, F5.0, 7F10.2)
, F5.0, 7F10.2)
, F5.0, 7F10.2)
, F5.0, 7F10.2)
, F5.0, 7F10.2) FORMAT(1H0,20HRAW FEED
FORMAT(1H0,20HINPUT TO PRI. CLAR.
FORMAT(1H0,20HEFF. OF PRI. CLAR.
FORMAT(1H0,20HUNDERFLOW PRI CLAR
FORMAT(1H0,20HINPUT TO AER TANKS
FORMAT(1H0,20HOUTPUT OF AER TANKS

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FORMAT(1H0,20HINPUT TO SEC. CLAR.
FORMAT(1H0,20HEFF. OF SEC. CLAR.
FORMAT(1H0,20HEFF. OF SEC. CLAR.
FORMAT(1H0,20HUNDERFLOW SEC CLAR.
FORMAT(1H0,20HWASTE ACT. SLUDGE
FORMAT(1H0,20HINPUT TO DIGESTER
FORMAT(1H0,20HDIGESTER S , F5.0, 7F10.2)
, F5.0, 7F10.2

RETURN
END

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LISTING OF THE COST ESTIMATION PROGRAM

SUBROUTINE COST1

LLST(50),NS(100),EN(100),SI(4,30),SO(4,30),KPRNT(10)
IS,NE,JJ,LOOP,NIN,NOUT,MSN,ISP,NC,III,NCALC,NOCOMP,NSR
MODE,NPLNT,LOOPC
SN(25,30),EEN(600),NPOINT(25,2)
ION.TITLE (10),ELECT(3) COMMON
COMMON
COMMON
COMMON COMMON SN

CALCULATES THE FIXED CAPITAL INVESTMENT, THE OPERATING AND
MAINTENANCE COSTS OF A WASTEWATER TREATMENT PLANT.
PLACES A SURTAX ON EFFLUENT IF THE EFFLUENT QUALITY IS POO
COSTS TAKEN FROM ECKENFELDER AND BARNHARDT, AND FROM POOR. SMITH.

EN VECTOR

- $1.$ MODULE **NUMBER** 1. MODULE TYPE
3. MODULE LENGTH
4.-15. = TWELVE ZEROES UNIT PROCESS

PRETREATMENT

NO. OF PRETREATMENT UNITS

PRIMARY SEDIMENTATION

NO. OF PRI. SETTLING TANKS

ACTIVATED SLUDGE TANKS

NO. OF AERATION TANKS

NO. DE ALERATION TANKS
- 111122222222222334567890 NO. OF AERATION TANKS
AIR BLOWERS
FINAL SEDIMENTATION
FINAL SEDIMENTATION
NO. OF FINAL TANKS
SLUDGE RETURN PUMPS
NO. OF SLUDGE RETURN PUMPS
ANAEROBIC DIGESTION
NO. OF ANAEROBIC DIGESTERS
CHLORINATION
NO. OF CHLORINATION UN CHLORINATION UNITS
FILTRATION
VACUUM FILTERS
INCINERATION
SLUDGE INCINERATERS
DRYING BEDS
DRYING BEDS
THICK BEDS NO. OF VĂCUUM OF NO. SLUDGE
NO. OF
NO. OF THICKENING
THICKENERS
HAULAGE SLUDGE NO. OF
SLUDGE
- SLOUGE HAULAGE
TRICKLING FILTERS
NO. OF TRICKLING FILTERS
AERATED LAGOONS
NO. OF LAGOONS
TYPE OF PLANT
TYPE OF PLANT
(FOR UTILITIES COST) 44 . CHE LAGOONS
E OF PLANT
R UTILITIES COST) $46.$

 $47.$ USED
USED NOT
NOT PRESENTLY
PRESENTLY $48.$ NOT PRESENTER USED
DESIGNED FLOW MIGPD
LABOUR COST S/MAN-H
ENR INDEX FOR YEAR
STREAM NUMBER OF FF
ACTUAL PLANT FLOW M -55523 S/MAN-HOUR S/YR)
EFFLUENT $\frac{54}{55}$. FLOW MIGPD

SURFACE AREA PER UNIT /1000 SQFT VOLUME PER UNIT MIGPD CAPACITY PER UNIT CPM/1000 SURFACE AREA/1000 SQFT PER UNIT CAPACITY MIGPD VOLUME PFR UNIT /1000 CUFT DESIGNED FLOW MIGPD SURFACE AREA PER UNIT/100 SQFT LBS SOLIDS/DAY PER UNIT SURFACE AREA SQFT SURFACE AREA PER UNIT SQFT SLUDGE VOLUME MIG/YR VOLUME PER UNIT/1000 CUFT SURFACE AREA PER UNIT/1000 SQFT

PARAMETER
DESIGNED FLOW MIGPD

 $\frac{1}{2}$: 0
3: 0 PLANT FOR **PRIMARY** \equiv TRICKLING FILTER PLANT
ACTIVATED SLUDGE PLANT \equiv FOR FOR \equiv

A = 1H*
B = 1H -
E LECT(1) = 101000.0
E LECT(2) = 162000.0
E LECT(3) = 378000.0

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287
           WRITE(6,404) PPC
           CALCULATE THE ENGINEERING COST FACTOR<br>CENG=0.08*(1.0E6/PPC)**0.146<br>CALCULATE THE ENGINEERING COSTS<br>COST=CENG*PPC<br>WRITE(6.405) COST<br>DPC=PPC+COST
           WRITE(6,402) (B,I=1,14)<br>WRITE(6,406) DPC
           CALCULATING THE CONTRACTOR'S FEE<br>COST=DPC*0.1
           WRITE(6,407)
                                    COST
           WRITENOST<br>CALCULATING THE CONTINGENCY COSTS<br>CALCULATING THE CONTINGENCY COSTS<br>WRITE(6,408) COST<br>FCI=FCI+COST
           WRITE(6,402) (B, I=1,14)<br>WRITE(6,409) FCI<br>WRITE(6,402) (B, I=1,14)
            OPERATING AND MAINTENANCE COST ESTIMATE
           WRITE(6,410)<br>WRITE(6,401)
                                    (A, I=1, 21)READ(5,200) TITLE
           WRITE (6,550)<br>WRITE (6,411)
                                    TITLE
            SUMHRS = 0.
           UPOM=Q_0Q=EN(55)DO 620 I=1,15
           IF(EN(14+2*I).LE.0.0) GO TO 620<br>GO TO (21,22,23,24,25,26,27,28,29,30,31,32,33,34,35),I
\frac{c}{21}HOUR=133.0*Q+510.0*Q**0.37<br>COST=HOUR*EN(51)
           WRITE(6,501) HOUR, COST
           GO TO 630<br>HOUR=(222.V*EN(18)+555.0*EN(18)**0.5)*EN(19)<br>COST=HOUR*EN(51)
22WRITE(6,502) HOUR, COST<br>GO TO 630<br>HOUR=(720.0*Q+590.0*Q/EN(20)**0.67)
23
           COST=HOUR*EN(51)<br>WRITE(6,503) HOUR, COST<br>GO TO 630<br>GO TO 620
24GO TO 620<br>GO TO 620<br>HOUR=(13.0*EN(28)+144.0*EN(28)**0.5)<br>COST=HOUR*EN(51)
\frac{25}{26}WRITE(6,507) HOUR, COST<br>GO TO 630<br>HOUR=(30,0*0*0+170,0*0**0.37)
28COST=HOUR*EN(S1)WRITE(6,508) HOUR, COST<br>GO TO 630<br>HOUR=(0.27*Q+154.0*Q**0.37)<br>COST=HOUR*EN(51)
29
           WRITE(6,509) HOUR, COST<br>GO TO 630<br>HOUR=(400.0*0+1600.0*0**0.37)
30COST=HOUR*EN(51)<br>
WRITE(6,510) HOUR, COST<br>
GO TO 630<br>
HOUR=(0,014*EN(36)+20,0*EN(36)**0,37)
31
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288 COST=HOUR*EN(51)
WRITE(6,511) HOUR,COST
GO TO 630 GO TO 620
GO TO 620
HOUR=(10.0*EN(42)+120.0*EN(42)**0.5)
COST=HOUR*EN(51) $\frac{32}{33}$ $\tilde{34}$ WRITE(6,514) HOUR, COST GO TO 650
HOUR=(50.0*EN(44)+100.0*EN(44)**0.5)
COST=HOUR*EN(51)
WRITE(6,515) HOUR,COST 35 $\frac{c}{630}$ CONTINUE
SUMHRS=SUMHRS+HOUR UPOM=UPOM+COST 620 **CONTINUE** $WRITE(6,415)$ $(B, I=1,28)$ WRITE(6,413) SUMHRS, UPOM $\frac{c}{c}$ CALCULATE THE SUPERVISION COST
COST=0.1*UPOM
WRITE(6,414) COST
DOM=UPOM+COST
CALCULATE THE UTILITIES COST \subset CALCULATE THE UTILITIES COST

KI=EN(46)+0.01

COST=ELECT(KI)*Q**0.65*EN(47)

WRITE(6,416) COST

DOM=DOM+COST

CALCULATE SLUDGE HAULAGE COSTS

CALCULATE SLUDGE HAULAGE COSTS

COST=8000.0*EN(40)**0.5

WRITE(6,513) COST

DOM= \subset C WRIIE(6,417) COSI

DOM=DOM+COSI

CALCULATE PLANT SUPPLIES

COST=0.05*UPOM

WRITE(6,418) COSI

DOM=DOM+COSI

WRITE(6,419) (B,I=1,14)

WRITE(6,420) DOM C CALCULATING LABORATORY COSTS \subset WRITE(6,421) COST
OMNET=DOM+COST
CALCULATING SURCHARGE ON EFFLUENT (IF EFFLUENT QUALITY IS POOR)
IF(EN(54).LE. 0.0) GO TO 50 \sim \subset S= EN(54).LE. 0.0, 50 10 50
CALL STREAM(S)
LBS=SN(IS,3)*3.0E-4
IF(SN(IS,10).LT.LBS) 60 TO
SURBOD=(SN(IS,10)-LBS)*0.05
GO TO 52
SURBOD=0.0
SURBOD=0.0 51 51 JE(SN(IS, 11). LT. LBS) GO TO
SURSS=(SN(IS, 11)-LBS)*0.05
GO TO 54
SURSS=0.0 52 53 53 SORTSUR=SURBOD+SURSS

WRITE(6,422) TOTSUR

OMNET=OMNET+TOTSUR

CONTINUE 54 50 WRITE(6,419)
WRITE(6,423) $(B, I=1, 14)$
OMNET CCCC CALCULATING THE COST OF THE FIXED CAPITAL INVESTMENT.
BASED ON AN INTEREST RATE OF 8.0 PERCENT AND A PLANT LIFE OF 25
YEARS. NOTE THAT THE ANNUITY PAYMENTS REPRESENT DEBT RETIREMENT
AND INTEREST CHARGED IN THE CASE OF BORR

RETURN.
END

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PROGRAM WWTP(INPUT, OUTPUT, WWRUN, TAPF2=INPUT, TAPF6=OUTPUT, ITAPE5=WWRUN) LLST(50),NS(100),EN(100),SI(4,30),SO(4,30),KPRNT(10)
IS,NE,JJ,LOOP,NIN,NOUT,MSN,ISP,NC,III,NCALC,NOCOMP,NSR
MODE,NPLNT,LOOPC
SN(25,30),EEN(600),NPOINT(25,2)
ON TITLE(10) COMMON COMMON **DIMENSION** DIMENSION NAME(20) **GEMCS/INTERCOM** ***** PRINTING CONTROL CHARACTERS ***** ***** PRINTING CONTROL CHARACTERS *****

KPRNT[1]=1 SUPPRESSES ABOVE PRINTING

KPRNT[2]=1 SUPPRESSES ABOVE PRINTING

KPRNT[2]=1 SUPPRESSES ABOVE PRINTING

KPRNT[3]=1 SUPPRESSES ABOVE PRINTING

KPRNT[4]=1 SUPPRESSES ABOVE P REWIND5 NAME(1)=8H MIXER1
NAME(2)=8H CONTL1 CONTL1 NAME $(3) = 8H$ SETST1 NAME (4) = 8H
NAME (5) = 8H SETSPI
SEPAO1 NAME(5) = 8H

NAME(6) = 8H

NAME(7) = 8H

NAME(7) = 8H

NAME(7) = 8H

SH

NAME(10) = 8H

NAME(11) = 8H

NAME(11) = 8H

NAME(15) = 8H

NAME(15) = 8H

NAME(15) = 8H

NAME(17) = 8H

NAME(17) = 8H

NAME(17) = 8H

NAME(17) = 8H PRISET1
ACISLI DPTRI SECLARI ϵ TRICK1 GRITI
ANDIG1 CHLORI
COSTI
SCREENI
REPIOI CONT₀₂ $NAME(18)=8H$ CONVO1 NAME (19) = 8H
NAME (20) = 8H CĂSET WRITE(6,234)
FORMAT(1HU,5UHTO ANSWER QUESTIONS BELOW TYPE 1.0/YES OR 0.0/NO λ $I I I = 25$ CALL DLOAD1 IK IS AN INDICATOR AS TO WHETHER A STRAIGHT THROUGH CALCULATION IS USED (IK=1) OR WHETHER A RECYCLE IS PRESENT (IK=0) $K=0$
IF(LOOP.EQ.999) IK=1

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 $\begin{matrix} 1 \\ 1 \end{matrix}$

WRITE(6,240)
FORMAT(1H0,30HWANT TO SET UP A NEW PLANT -
READ(2,221) FLAG
IE(FLAG,LE.0.0) GO TO 92 240 λ CONTINUE 90 CALL FLW 92 CONTINUE
WRITE(6,238)
FORMAT(1X,45HWANT TO SET A RANDOM FLOW AND CASE STUDY -
READ(2,221) FLAG
IF(FLAG.LE.U.U) GO TO 93 96 238 λ خ
236ع
5
5 CONTINUE
SETS A CASE RUN AROUND INPUT FEED
CASET MUST HAVE EQUIPMENT NUMBER 25 AND SHOULD NOT BE INCLUDED
IN THE CALCULATION ORDER.
CALL DISKIO(1,25)
CALL CASET
GO TO 1000 $\frac{C}{93}$ CONTINUE
CALL STREAM(1.0)
IN=IS CALL STREAM(2.0)
IF(IS.LE.III) GO
CALL STREAM(0.0)
SN(IS.1)=2.0 TO 94 CONTINUE 94 DO 600 J=2, JJ
SN(IS, J)=SN(IN, J) 600 5000 CONTINUE
LOOPC=1
WRITE (6,196) LOOPC $\begin{matrix} 0 \\ 1 \\ 0 \\ 0 \end{matrix}$ CONTINUE CALCULATING EQUIPMENT IN CALCULATION ORDER LIST $N\overline{C} = \overline{1}$ $MODE = 0$ KTEST=0
LOOP=1 IF(IK.EQ.0) GO TO 98 $MODE = 1$ $KTEST=1$ $LOOP = 999$ 98 CONTINUE C_{C}°
 C_{102}° READ EN VECTOR IF (KPRNT(5)) 108,108,104
CONTINUE $\overline{1}0\overline{4}$ WRITE (6,182) LOOP
DO 106 IKE=1,III
IF (SN(IKE,1).LE.0.0) GO TO 106
WRITE (6,194) (SN(IKE,J),J=1,JJ)
CONTINUE 106 $\bar{\zeta}$ 08. NE=LLST(NC)
MM=NE CALL DISKIO (1,MM)
NN=EN(3)+.001
NIN=EN(6)+.001
NOUT=EN(11)+.001 NTYPE=ABS(EN(2))+.001 $KSW = 0$ IF (EN(2).LE.O.) KSW=1
ISP=U SUPPRESSES ALL PRINTING OF INPUT AND OUTPUT STREAMS \subset $ISP = 0$ C
C
C FINDING INPUT STREAMS

IF(NIN.EQ.0) GO TO 128
DO 122 I=1,NIN
S=EN(I+6) S=EN(1+6)
CALL STREAM (S)
IF (IS-III) 114,114,110
CONTINUE
SI(I,1) = S
DO 112 J=2,JJ
SI(I,J) = 0.
CONTINUE 110 112 GO TO 122

GO TO 122

SI(1, J)=SN(1S, J)

CONTINUE

IF (INT(S+. 001)) 114 116 $(i\bar{j}\bar{N}\bar{T}\bar{S} + \bullet \cup 01)) 122.122.118$ 118 CONTINUE $M = S + 001$ IF (NS(M)-6) 122,120,120 CONTINUE
SN(IS,1)=0.
CONTINUE
LES (ISP) 12 120 122 TE (15P) 128,128,124

WRITE (6,184) NE ,NAME(NTYPE)

DO 126 IKE=1,NIN

WRITE (6,194) (SI(IKE,J),J=1, 124 $(SI(IKE, J), J=1, JJ)$ CONTINUE 126 $\begin{array}{c} 1 \\ 1 \\ 2 \\ 3 \\ 4 \end{array}$ CONTINUE CALL MODULE (NTYPE) CCCC STORING OUTPUT STREAMS AND PRINTING IF(NOUT) 166,166,136 CONTINUE
DO 150 1=1,NOUT 136 IF(SO(I,3).LE.0.0) GO TO 150 $S = EN(I+11)$ S=EN(I+11)
SO(I,1)=S
M=S+.001
IF(NS(M)-6) 140,150,140
CALL STREAM(S)
CALL STREAM(0.)
IF(IS-III) 146,146,144
IF(IS-III) 146,146,144
WRITE (6,190)
GO TO 150
DO 148 J=1,J
DO 148 J=1,JJ
SN(IS,J)=SO(I,J)
CONTINUE 140 142 144 146 $\begin{array}{c}\n148 \\
150\n\end{array}$ IF (ISP) 158,158,154 WRITE (6,186) NE , NAME(NTYPE)
DO 156 IKE=1,NOUT 154 WRITE(6,194)
WRITE(6,194)
WRITE (6,194) (SO(IKE,J),J=1,JJ)
CONTINUE 256 IF (LOOP-999) 166,160,160
IF (KSW.NE.0) GO TO 166
IF(KPRNT(6).LT.1) GO TO 166
WRITE(6,188) NE, NAME(NTYPE) 158 WRITE(6,194)

MRITE(6,194)

WRITE(6,194)

WRITE(6,195) (SO(IKE,J),J=1,JJ)

FORMAT(1X,5F12.3) 195
164
C
166 $NC = NC + 1$ IF (NC-NCALC) 102,102,168

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 $\frac{C}{168}$ CONTINUE
IF (LOOP-999) 170,172,172
CONTINUE
NC=0
GO TO 166
IF (KTEST) 174,174,176
NC=0
MODE-1 170 172 $\overline{1}\overline{7}\overline{4}$ $MODE = 1$ GO TO 166 176 CONTINUE CONTINUE
CALL ICLOAD
WRITE(6,220)
FORMAT(1X,35HWANT TO MAKE ANY PLANT CHANGES -
READ(2,221) FLAG
JF(FLAG.LE.0.0) GO TO 230
CALL ENSET
CALL ENSET
LOOPC=LOOPC+1
WRITE(6,196) LOOPC
GO TO 100
CONTINUE
WRITE(6,238) 220 λ $\mathcal{L}_{\mathcal{A}}$ CONTINÛĔ
WRITE(6,238)
FORMAT(1X,45HWANT TO GENERATE A NEW RANDOM CASE STUDY –
READ(2,221) FLAG
IF(FLAG.LE.U.U) GO TO 95
GO TO 236
CONTINUE
WRITE(6,240)
WRITE(6,240)
READ(2,221) FLAG
IF(FLAG.GE.1.0) GO TO 90
STOP 230 232 Ń 95 $\frac{6}{182}$ $(1HU, 32H5555$
 $(1HU, 31H5555$ SN TABLE ON ENTERING LOOP , 15,7
INPUT STREAMS FOR MODULE , 15,4H $515.7H$ \$\$\$\$\$) FORMAT FORMAT
1A8,7H
FORMAT 184 $$5555$
(1H0,32H55555 OUTPUT STREAMS FOR MODULE , 15, 4H 186 $-$, A8, 188 FINAL OUTPUT STREAMS FOR MODULE , 15,4H ERROR IN SN *****)
*** END OF EXECUTION 190 $\frac{1}{1}$ 92 ********** λ $\overline{1}96$ **** **RUN, 14, 26H** 198 FORMAT (10A8) $\frac{200}{221}$ FORMAT (1HO, 10A8)
FORMAT (F12.3)

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END

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 C NCALC=NCALC+1
LLST(NCALC)=9 NCALC=NCALC+1 $LIST(NCALC) = 10$ $\tilde{\epsilon}$ ASKING FOR DETAILS OF THE WASTE SLUDGE POLICY MO=NPOINT(10,1) WRITE(6,535) $FORMATIIX, 30HFRACTION OF SLUDGE WASTED =$ 535 ¹ READ(2,200) F $EEN(MQ+17)=F$ $EEN(MQ + 16) = 1.0 - F$ NCALC=NCALC+1 C WRITE(6,420)
FORMAT(1X,20HIE WASTE ACT SL TO 420 ,/,5X,2CHPRI CLAR -TYPE 1.0 1,/,5X,31HDIRECTLY TO DIGESTER -TYPE 2.0 $, 50, 50, 21H$ THICKENER - TYPE 23.0

READ(2,200) F

JF=F+0.01

MQ=NPQINT(1),1) $EEN(MQ+16)=0.0$ EEN(MQ+17)=0.0
EEN(MQ+18)=0.0
EEN(MQ+18)=0.0
EEN(MQ+15+JF)=1.0 EENTMQ+15+JF)=1.0

IF(F.LT.3.0) GO TO 40

NCALC=NCALC+1

LLST(NCALC)=12

NCALC=NCALC+1

LLST(NCALC)=19

WRITE(6,430)

FORMAT(1.300) 430 Y $READ(2,200) F$ MQ=NPOINT(19,1) $EEN(MQ+17)=F$ $EEN(MQ+16) = 1.0-F$ $\frac{C}{40}$ NCALC=NCALC+1
LLST(NCALC)=20 $\mathsf C$ WRITE(6,440)
FORMAI(1HV,18HANY DIGESTERS - 440 \mathbf{I} READ(2,200) F IF(F.LT.1.0) GO TO 50
LP2=1 NCALC=NCALC+1
LLST(NCALC)=13 ASKING FOR DETAILS OF THE DIGESTERS

WRITE(6,450)

FORMAT(1X,30HONE OR TWO STAGE DIGESTION -

READ(2,200) F

MG=NPOINT(13,1)

EEN(MQ+16)=F-1.0

EEN(MQ+17)=1.0-EEN(MQ+16)

IF(F.LT.2.0) GO TO 60

IF(F.LT.2.0) GO TO 60

LLST($\frac{c}{c}$ 450 λ 460 $EEN(MQ+4)=F$ WRITE(6,515)
FORMAT(5X,30HDIAMETER OF TANK (IN FT) =
READ(2,200) F
EEN(MO+16)=F λ 515 \overline{W} RITE(6,540) READ(2,200) -F $EEN(MQ+17)=F$ WRITE(6,525)

298 WRITE(6,710)
FORMAT(1X,31HAVERAGE BOD5 TO YOUR PLANT = |)
READ(2,200) F
SN(IS,10)=FLOW*10.0*F*1.0E-6
WRITE(6,720)
FORMAT(1X,43HAVERAGE SUSPENDED SOLIDS TO YOUR PLANT =
READ(2,200) F
SN(IS,11)=FLOW*10.C*F*1.0E-6
WRITE(6,73 710 720 $\overline{}$ WRITE(6,730)
FORMAT(1X,36HAVERAGE DISSOLVED ORGANIC CARBON =
READ(2,200) F
SN(IS,9)=FLOW*10.0*F*1.0E-6 730 λ SN(15,92)=FLOW*10.0*F*1.0E-6

SN(15,22)=0.2*SN(15,9)

SN(15,20)=0.2*SN(15,9)

SN(15,21)=SN(15,9)-SN(15,20)-SN(15,22)

BODSOL=1.9*(SN(15,20)+SN(15,21))

BODP=SN(15,10)-BODSOL

VSS=BODP/1.2 VSS=BODP/1.2

SN(IS,15)=FLOW*10.0*5.0E-6

SN(IS,16)=0.5*VSS

SN(IS,17)=0.0

SN(IS,18)=0.2*VSS

SN(IS,19)=0.3*VSS

SN(IS,19)=0.3*VSS

SN(IS,12)=2.1*(VSS+SN(IS,15))

SN(IS,12)=0.6*(SN(IS,11)-SN(IS,12))

SN(IS,14)=0.4*(SN(IS $SN(I5, 8) = SN(I5, 9) + VSS + SN(I5, 15)$ $IV = \overline{I} S$ CALL STREAM(2.0)
DO 620 J=2,JJ
SN(IS,I)=SN(IV,I) 6^{20} CALL STREAM(16.0) $DO 640 J=6.00$ $SN(TS, J) = SN(TS, J) * FRAC$ CONTINUE
SN(IS,3)=SN(IS,3)*FRAC 640 PURGE SN FILE
DO 630 I=1, II
IS=SN(I,1)+0.01 C IF((IS.EQ.1).OR.(IS.EQ.16).OR.(IS.EQ.2)) GO TO 630 SN(1,1)=0.0
CONTINUE 630 **RETURN** END

SURROUTINE CASET

LLST(50),NS(100),EN(100),SI(4,30),S0(4,30),KPRNT(10)
IS,NE,JJ,LOOP,NIN,NOUT,MSN,ISP,NC,III,NCALC,NOCOMP,NSR COMMON MODE, NPLNT, LOOPC **COMMON** SN(25,30), EEN(600), NPOINT(25,2) **COMMON** COMMON /C/ IND GENERATES A CASE STUDY AROUND THE AVERAGE RAW FLOW EN VECTOR 1.-15. STANDARD FORMAT $\frac{6}{11}$. STREAM NUMBER OF USUAL RAW WASTE FLOW (=1.0) 1.0 $=$ STREAM NUMBER
EQUIPMENT NUM
EQUIPMENT NUM GENERATED INPUT STREAM (=2.0)
OF PRIMARY SETTLER (=5.0)
OF THE ACTIVATED SLUDGE TANKS
OF THE SECONDARY CLARIFIERS $\frac{12}{16}$. ABER OF
NUMBER
NUMBER REAM (2.0)
R (=5.0)
SLUDGE TANKS $\frac{17}{18}$. $(= 7 \cdot 0)$ **NUMBER** EQUIPMENT $(=8.0)$ $\frac{1}{2}8$: EQUIPMENT NUMBER OI $\begin{bmatrix} 1.1 \end{bmatrix}$ DIGESTERS
5 TO 1.0E8 $\begin{matrix} 15.0 \\ 15.0 \\ 15.0 \\ 15.0 \end{matrix}$ OF \leq THE STARTER IN THE RANDOM NUMBER GENERATOR IF(EN(20).GE.1.0E6) GO TO 9
WRITE(6,470) WRITE(63470)
FORMAT(1X,5UHTYPE ANY NUMBER BETWEEN 1 MILLION TO 100 MILLION
READ(2,200) F
FORMAT(F12.3)
MQ=NPOINT(25,1)
EEN(MQ+20)=F 470 200 $EN(20) = \overline{F}$ $\overline{IND} = \overline{EN(20)}$
 $IND = IND*2+1$ CONTINUE WRITE(6,500)
FORMAT(1X,2CHREADY TO PLAY GAMES $\frac{500}{c}$ **>/>20H++...CLUES.....** $S = EN(7)$
CALL STREAM(S)
IIN=IS S=EN(12)
CALL STREAM(S) IF(IS.LE.III) GO TO 22
CALL STREAM(0.0) CONTINUE $IO=IS$ $SN(10,1) = EN(12)$ GENERATING A RAW WASTE FLOW
SET WATER FLOW
CONTINUE CALL RANDOM(Z)
IF(Z.LT.0.25) GO TO 10
SN(IO,6)=SN(IIN,6)*Z*2.0 SET COMPONENT FLOWS
DO 600 J=13,JJ
CONTINUE CALL RANDOM(Z)
IF(Z.LT.0.25) GO TO 20
SN(IO,J)=SN(IIN,J)*Z*2.0
CONTINUE $\frac{6}{5}^{00}$ SET TOXIC COMPONENT FLOW
CALL RANDOM(Z) $SN(IO, 30) = Z$ $SN(IO, 2) = SN(III, 2)$ $SN(10, 4) = SN(11N, 4)$
SN(10, 5)=SN(11N, 5)
SN(10, 7)=SN(11N, 7) CALL BALNCE(1,IO)

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 C ₂₄ GENERATING EQUIPMENT FAILURES CALL RANDOM(Z)
KO=Z*11.0+1.0
GO TO (1,1,2,2,3,3,4,4,4,4,5,5,5),KO $MM=EN(16)+U$. UI 1 DO 610 I=1, NCALC
IF(LLST(I).EQ.MM) GO TO 620 610 GO TO 24
CONTINUE
MQ=NPOINT(MM,1) 620 IF(KO.EQ.2) GO TO 11
EEN(MQ+4)=EEN(MQ+4)-1.0 WRITE(6,400)
WRITE(6,400)
FORMATIIX,25HONE PRIMARY TANK IS DOWN 400 J. GO - TO **C** EEN(MQ+23)=EEN(MQ+23)-1.0
WRITE(6,410)
FORMAT(1X,50HONE OF THE PRIMARY SLUDGE PUMPS IS NOT WORKING 11 -410 GO TO -5 $\frac{c}{2}$ $MN = EN(17)+0.01$ THE 230 I=1,NCALC
DO 630 I=1,NCALC
IF(LLST(I).EQ.MM) GO TO 640 DO 630 630 GO. $\overline{10}$ 24 640 CONTINUE $MQ = NPOINT(MM, 1)$ IF(KO.EQ.4) GO TO 12
EEN(MQ+4)=EEN(MQ+4)-1.0 TO WRITE(6,420)
FORMAT(1X,35HONE ACTIVATED SLUDGE TANK IS DOWN 420 λ GO TO 5 EEN(MQ+45)=EEN(MQ+45)-1.0
WRITE(6,430) 12 FORMATIIX,40HONE OF THE AIR BLOWERS IS DOWN 430 ١ $\frac{C}{3}$ $MM=EN(18)+0.01$ DO 650 I=1, NCALC
IF(LLST(I).EQ.MM) GO TO 660 650 $\overline{60}$ $\overline{10}$ 24 660 CONTINUE MO=NPOINT(MM,1)
IF(KO.EQ.6) GO TO 13
EEN(MQ+4)=EEN(MQ+4)-1.0 WRITEL6,440)
FORMATIIX,35HONE OF THE SECONDARY CLARIFIERS IS DOWN 440 ١ GO TO 5 $EEN(MQ+20) = EEN(MQ+20) - 1.0$ 13 WRITE(6,450) FORMAT(1X,45HONE OF THE SECONDARY SLUDGE PUMPS IS DOWN
GO TO 5 450 λ C 4 $MM = EN(19)+0.01$ NO 670 I=1,NCALC
IF(LLST(I).EQ.MM) GO TO 680
GO TO 24 670 CONTINUE 680 MQ=NPOINT(MM,1) MQ=NPOINT(MM,1)
KO=KO-6
GO TO (14,15,16,17),KO
IF(EEN(MQ+4),LT,2,0) GO TO 24
EEN(MQ+4)=EEN(MQ+4)-1.0
WRITE(6,460)
WRITE(6,460)
FORMAT(1X,25HONE DIGESTER IS DOWN
GO TO 5
CONTINUE
CO TO 5
CONTINUE 14 460 -1 15 CALL RANDOM(Z) 16 $EEN(MQ+24)=Z$ GO TO -5 $\frac{1}{5}$ $EEN(MQ+19)=25.0$ CONTINUE

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RETURN
END

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SUBROUTINE RANDOM(Z)

GENERATES RANDOM NUMBERS HAVING A UNIFORM DISTRIBUTION BETWEEN 0-1

COMMON /C/ IND

IF(IL.EQ.0) GO TO 1
IL=0
M=2**30 $FM=M$
 $N=2**15+3$
 $IX=IND$ CONTINUE
IX=MOD(N*IX,M)
FX=IX
Z=FX/FM

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RETURN

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               TCOST=TCOST+COST(IJ)<br>GO TO 120
\frac{c}{100}CONTINUE<br>WRITE(6,470) TCOST<br>FORMAT(1HU,3UH.....TOTAL ANALYTICAL COST = $
4705F8.21WRITE(6,520)<br>FORMAT(1HU,45HWANT TO SEE THE SUMMARY REPORT ON STREAMS -<br>READ(2,200) AN<br>IF(AN.LT.1.0) GO TO 190<br>CALL DISKIO(1,24)<br>CALL REPTO1<br>CONTINUE
520\lambda290RETURN<br>END
               SUBROUTINE ENSET
\frac{c}{c}COMMON<br>COMMON
                               LL ST(50) »NS(100) »EN(100) »SI(4,30) »SO(4,30) »KPRNT(10)<br>IS »NE »JJ»LOOP »NIN »NOUT »MSN »ISP »NC »III »NCALC »NOCOMP »NSR<br>MODE »NPLNT »LOOPC<br>SN(25 »30) »EEN(600) »NPOINT(25 »2)
               COMMON
CALLOWS PLAYER TO MAKE PLANT ADJUSTMENTS
               CONTINUE<br>WRITE(6,500)
\overline{1}40
500
               FORMATIIX, 22HTYPE EQUIPMENT NUMBER
              FORMAT(1X,22HTYPE EQUIPMENT NUMBER<br>READ(2,200) S<br>FORMAT(F12.3)<br>F(S.LE.0.01) GO TO 130<br>MM=S+0.01<br>MM=S+0.01<br>MM=S+0.01<br>MO=NPOINT(MM.1)<br>CONTINUE<br>VRITE(6,510)<br>VRITE(1X,44HTYPE ELEM. NUMBER AND VALUE CHANGED TO<br>READ(2,210) SJ,EL
                                                                                                                 \lambda200
150
510\lambda210
               I J=SJ+0.01<br>EEN(MQ+IJ)=EL<br>GO TO 150
\begin{array}{c} C \\ 130 \\ C \end{array}CONTINUE
               RETURN<br>END
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27:00 \circ 0 \overline{O} 0 $\frac{0}{0}$ 15.0
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0.7 -3.3 \circ $\mathbf 0$ \circ $\frac{16}{23}$
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