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RBC PERFORMANCE UNDER TRANSIENT LOADING CONDITIONS

Performance of Rotating Biological Contactors
under Transient Loading Conditions

By

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Abstract

This report examines the dynamic response of a 0.5 metre pilot scale rotating biological contactor when operated under transient influent conditions. Experimental data are presented for two modes of operation: carbon oxidation and carbon oxidation plus nitrification. During the carbon oxidation experimental runs, a 2.0 metre RBC was operated in parallel with the 0.5 metre RBC, thus allowing comparisons in the performance of the units.

When the 0.5 metre RBC was operated under nitrifying conditions, it was found that the effluent filterable TKN responded positively to influent variations in filterable TKN loading, TKN concentration and hydraulic loading. Transfer function - noise models were developed which successfully predicted the time varying effluent TKN response. The response in effluent filterable TKN was predicted most precisely by influent TKN loading. Models based on influent TKN concentration and flow were not as precise in predicting effluent response. The effluent response of the 0.5 metre RBC was found to be greater than the response of activated sludge pilot units when operated at similar levels of removal.

When the 0.5 metre RBC was operated in the carbon oxidation mode, significant effluent responses were observed for carbon loading and concentration. Little correlation was found between influent flow and effluent carbon concentration. Operating under identical conditions, the 2.0 m RBC showed significant responses to carbon loading, carbon concentration and flow.

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TABLE OF CONTENTS

	<u>PAGE</u>
Abstract	ii
Acknowledgments	iv
Table of Contents	v
List of Figures	vii
List of Tables	ix
1. Introduction	1-1
2. Literature Review	2-1
2.1 Process Description	2-1
2.2 Historical Development	2-2
2.3 Carbon Oxidation	2-5
2.3.1 Design	2-5
2.3.2 Performance	2-7
2.3.3 Diurnal Variability, Hydraulic Pulses and Intermittent Flow	2-7
2.4 Nitrification	2-9
2.4.1 Design	2-10
2.4.2 Performance	2-10
2.5 Denitrification	2-11
2.6 Steady State Modelling	2-12
2.7 Dynamic Modelling	2-16
2.7.1 Linear Dynamic Stochastic Modelling	2-18
2.7.2 The Box and Jenkins Method of Developing Linear Dynamic Stochastic Models	2-20
3. Experimental Equipment and Procedures	3-1
3.1 Pilot Plant Design and Operation	3-1
3.2 Experimental Plan and Start-Up	3-1
3.3 Diurnal Flow Experiments	3-5
3.4 Dynamic or Non "Steady State" Experimental Design	3-5
3.5 Feed Characteristics	3-7
3.6 Sample Preparation and Analyses	3-15
4. Results and Discussion	4-1
4.1 Dynamic Time Series Data	4-1
4.2 System Response to Hydraulic, Organic and Inorganic Loading Variations	4-5
4.2.1 General	4-5
4.2.2 Impulse Response Functions - Carbon Oxidation Mode	4-8
4.2.3 Impulse Response Functions - Nitrification Mode	4-11
4.2.4 Discussion	4-14
4.3 Dynamic Transfer Function - Noise (TF-N) Model Development	4-14
4.3.1 Introduction	4-14
4.3.2 Model Development	4-15
4.3.3 Effluent Filterable TKN Models	4-15
4.3.4 TF-N model Analysis to Determine Response Times	4-21

	<u>PAGE</u>
4.4 System Response to Natural Diurnal Variations in Flow and Concentration	4-21
4.4.1 TF-N Model Forecast Results	4-22
4.5 Comparison of RBC TF-N Models with Activated Sludge Models	4-24
4.6 Steady State Design	4-24
4.6.1 Carbon Oxidation Mode	4-26
4.6.2 Carbon Oxidation plus Nitrification Mode	4-31
4.6.3 Use of Mass Loading Plots in Design	4-35
4.7 Scale-Up	4-35
5. Conclusions	5-1
6. Recommendations for Future Work	6-1
7. References	7-1
Appendix A - <u>Data Listing</u>	A-1
Appendix B - <u>Analytical Procedures</u>	B-1
Total Kjeldahl Nitrogen	B-1
Ammonia	B-1
Nitrate	B-1
Nitrate plus Nitrite	B-2
Chemical Oxygen Demand (COD)	B-2
Biochemical Oxygen Demand (BOD)	B-3
Total Organic Carbon (TOC)	B-3
Suspended Solids	B-3
Dissolved Oxygen	B-3
Temperature	B-4
pH	B-4
Alkalinity	B-4
Appendix C - <u>Calculation Procedures</u>	C-1
Identification of the Transfer Function Model:	
TKN Load vs TKN Concentration	C-1
Identification of the Noise Model	C-2
Fitting and Applying Residual Checks to the Combined Transfer Function - Noise (TF-N) Model	C-2
Forecasting System Response to Impulse Forcings	C-6
Transfer Function Calculation Procedures to Determine Response Times	C-7
Appendix D - <u>Reactor Retention Times and Mixing Characteristics</u>	D-1
Tanks in Series Model Theory	D-2
Dispersion Model Theory	D-2
Appendix E - <u>Abbreviations and Symbols</u>	E-1

LIST OF FIGURES

<u>Figure No.</u>	<u>Description</u>	<u>Page</u>
3.1	Process Schematic	3-2
3.2	Planned Diurnal Flow Input to 0.5 m RBC for Peak/Average/Minimum Flow Ratio of 2/1/0.5	3-6
3.3	Experiment C1 Design, Carbon Oxidation Plus Nitrification	3-9
3.4	Experiment C2 and E2 Design, Carbon Oxidation Mode	3-9
3.5	Filterable COD Raw Wastewater Characteristics	3-10
3.6	Filterable TOC Raw Wastewater Characteristics	3-11
3.7	Filterable TKN Raw Wastewater Characteristics	3-12
3.8	Suspended Solids Raw Wastewater Characteristics	3-13
3.9	Correlation of Filterable BOD ₅ with Filterable TOC, Summer of 1976	3-14
4.1	Input and Response of 0.5 m RBC, Carbon Oxidation Plus Nitrification Mode	4-2
4.2	Input and Response of 0.5 m RBC, Carbon Oxidation Mode	4-3
4.3	Input and Response of 2.0 m RBC, Carbon Oxidation Mode	4-4
4.4	Effluent Filterable TOC Impulse Response Weights, 0.5 m RBC, Carbon Oxidation Mode, First Difference	4-9
4.5	Effluent Filterable TOC Impulse Response Weights, 2.0 m RBC, Carbon Oxidation Mode, First Difference	4-10
4.6	Effluent Filterable TOC Impulse Response Weights, 0.5 m RBC, Carbon Oxidation Plus Nitrification Mode, First Difference	4-12
4.7	Effluent Filterable NO ₂ -N plus NO ₃ -N Impulse Response Weights, 0.5 m RBC, Carbon Oxidation Plus Nitrification Mode, First Difference	4-13

LIST OF FIGURES (Cont'd)

<u>Figure No.</u>	<u>Description</u>	<u>Page</u>
4.8	Comparison of Effluent Filterable TKN Response and TKN Load Model (A2) Fit	4-20
4.9	Effluent Filterable TKN Response and Forecast for Diurnal Loading	4-23
4.10	Steady State Design Data for Filterable TOC Removal, 0.5 m RBC, Carbon Oxidation Mode	4-27
4.11	Steady State Design Data for Filterable COD Removal, 0.5 m RBC, Carbon Oxidation Mode	4-28
4.12	Steady State Design Data for Filterable TOC Removal, 2.0 m RBC, Carbon Oxidation Mode	4-29
4.13	Steady State Design Data for Filterable COD Removal, 2.0 m RBC, Carbon Oxidation Mode	4-30
4.14	Steady State Design Data for Filterable TOC Removal, 0.5 m RBC, Carbon Oxidation plus Nitrification Mode	4-32
4.15	Steady State Design Data for Filterable COD Removal, 0.5 m RBC, Carbon Oxidation plus Nitrification Mode	4-33
4.16	Steady State Design Data for Filterable TKN Removal, 0.5 m RBC, Carbon Oxidation plus Nitrification Mode	4-34
4.17	Comparison of TOC Mass Removal, 0.5 m RBC versus 2.0 m RBC	4-36
4.18	Comparison of COD Mass Removal, 0.5 m RBC versus 2.0 m RBC	4-37
C-1	Autocorrelation and Partial Autocorrelation Functions of TKN Load Model Noise Sequence	C-3
C-2	Autocorrelation of the Fitted TKN Load TF-N Model Residuals	C-5
C-3	Cross correlation Model Check, ∇X_t vs. TF-N Model Residuals	C-5

LIST OF TABLES

<u>Table No.</u>	<u>Description</u>	<u>Page</u>
3.1	Summary of Experimental Programme	3-4
3.2	Design Levels for Experiments C1, C2 and E2 Carbon Oxidation Mode	3-8
4.1	Summary of Cross Correlation Results, First Difference	4-6
4.2	Transfer Function - Noise Models Describing Effluent Filterable TKN Concentration	4-16
4.3	Diagnostic Checks Applied to the Residuals	4-19
4.4	Substrate Loading Models for RBC and Activated Sludge Processes	4-25
A-1	General Operating Data, Experiments A-1, A-2, A-3, A-4, B-1, B-2, C-1, C-2 and E-2	A-2
A-2	Bio-Surf Pilot Plant Testing, Analytical Results, Experiments A-1, A-2, B-1, B-2, C-1, C-2 and E-2	A-21
C-1A	Sample Calculation Model Estimates	C-3
C-1	Computer Program Used for Cross Correlations and Matrix Inversion, with Sample Output for Model A-1	C-9
C-2	Main Program Used for Estimation of the Noise Model, with Sample Output for Model A-1.	C-22
C-3	Computer Program used for Simultaneous TF-N Model Parameter Estimation and Residual Checks, with Sample Output for Model A-1	C-32
C-4	Subroutine TSHAUS	C-45
C-5	Forecast Program Used in Predicting Effluent Response	C-53
D-1	Tracer Response Analysis, 0.5 metre RBC, at 0.82 L/min	D-5
D-2	Tracer Response Analysis, 0.5 metre RBC, at 2.33 L/min	D-9
D-3	Tracer Response Analysis, 0.5 metre RBC, at 3.46 L/min	D-14

LIST OF TABLES (Cont'd)

<u>Table No.</u>	<u>Description</u>	<u>Page</u>
D-4	Tracer Response Analysis, 2.0 m RBC, at 37.2 L/min	D-16
D-5	Tracer Response Analysis, 2.0 m RBC, at 100.6 L/min	D-20
D-6	Tracer Response Analysis Computer Program	D-24

Performance of Rotating Biological Contactors
under Transient Loading Conditions

1. INTRODUCTION

The rotating biological contactor (RBC) is an aerobic biological treatment system based on the biosorption principle. It employs captive biological slimes to remove substrate from the liquid wastewater by physical and biochemical means.

RBC treatment design criteria are still in the development stage and have generally been based on hydraulic loading. Early investigations (Antonie, 1970) indicated that BOD₅ and ammonia removal followed first order kinetics. This led to designs based on percentage removal and hydraulic loadings. Further refinement of this approach provided designs based on effluent concentrations and areal hydraulic loadings, with no consideration provided for influent substrate concentrations or mass loadings. Recent publications (Antonie, 1976; Poon et al, 1977) suggest that influent substrate concentrations also have an influence on the design relationships.

Capital costs for the RBC have been shown to decrease linearly with hydraulic capacity (Antonie, 1976), potentially providing relatively inexpensive biological treatment for small communities, work camps, and summer camps. However, wastewaters from these sources tend to have large diurnal flow and substrate concentration fluctuations (Randtke et al, 1977) and if the RBC is to be used in these applications its dynamic behaviour must be evaluated and quantified.

Attempts to date to model the performance of the RBC have been based upon steady-state equations. Of the few kinetic studies reported, the deterministic models which have been developed are based on two general concepts (Friedman et al, 1976; Kornegay, 1969; Famolaro et al, 1976; Hansford et al, 1978):

1. kinetic models for growth, or
2. empirical models based on mass transport concepts.

To describe the removal capability of an RBC using these concepts, deterministic models must attempt to incorporate RBC mixing characteristics, retention time, active biofilm thickness, rotational speed, available surface area, system geometry, and diffusion coefficients for oxygen and substrate transfer. The parameter values describing these characteristics are estimates at best, and introduce potential weaknesses to the models. Further, model parameter values obtained in this fashion will not describe the dynamic relations between influent and effluent when the input to the RBC is highly variable.

An estimate of the stability for a RBC can only be obtained using techniques which elucidate the response to non-steady operation. Incorporation of the process dynamics into a deterministic model yields a complex system of linear differential equations (Kornegay, 1969). Time series analysis is an alternative to be considered as a method to describe the process dynamics. Using this method, insight into the significant influent variables affecting effluent quality, and forecasting of effluent quality for various influent values can be obtained to provide a basis for process assessment and control. Time series analysis as developed by Box and Jenkins (1970) is one approach that can be used to relate influent and effluent variables for actual experimental or operational data. The resulting model may be updated as more information becomes available. Berthoeux et al. (1976) and Murphy et al. (1977) have previously demonstrated the potential of time series model building for describing wastewater treatment systems.

This study has examined the performance of pilot scale RBCs operating under variable influent loadings of organic carbon and ammonia. The experimental design allowed the use of

time series analysis to examine the response of RBCs under non-steady influent conditions and provided a means for assessing design criteria for the RBC. Comparisons of the performance of the pilot scale RBC with a full scale RBC and conventional activated sludge units have been made.

2. LITERATURE REVIEW

2.1 Process Description

The Rotating Biological Contactor (RBC) is an aerobic biological waste treatment system which is based on the "biosorption" principle. By using the adsorptive and absorptive properties of captive biological slimes, organic material is removed from liquid wastes by physical and biochemical means. Each unit consists of a large number of lightweight polyethylene discs which are bonded together in a stack, mounted on a horizontal shaft and placed in a semicircular tank. The discs, which are vacuum-formed in sizes up to four (4) metres in diameter, are slowly rotated while approximately one half of their surface area is submerged in the wastewater. Rotational speeds are generally in the range of 2 to 20 rpm.

Immediately after startup, organisms naturally present in a wastewater begin to adhere to the disc surfaces, and multiply. After a period of 2 to 3 weeks, the unit normally has acclimatized and produces a relatively stable, treated effluent. As the discs rotate, the growth is alternately exposed to the wastewater and atmosphere. The rotating action aerates the wastewater, provides oxygen necessary for organism growth and controls the biomass population. Shearing forces exerted on the biomass cause any excess biomass to slough from the discs into the mixed liquor. The solids remain in suspension until subsequent removal by settling.

RBCs are relatively new to North America. The acceptance and application of new and different methods of wastewater treatment systems has historically taken many years. As a result, the literature available on RBCs is general and tends to concentrate on their many apparent advantages. These may be summarized as follows.

1. The system is not subject to washout conditions because of the large captive biomass.
2. The biomass population for the treatment of domestic wastewaters has been estimated to be the equivalent of 18000 to 30000 mg/L MLSS (Joost, 1969; Welch, 1968). Manufacturers suggest that these large numbers of captive microorganisms will provide a low F/M ratio and enable the system to absorb organic shock loads.
3. Capital investment costs are lower than activated sludge (Antonie, 1977) or trickling filter systems (Winkler, 1974). However, these costs increase linearly with treatment capacity and thus lose their competitive edge for large facilities.
4. Operating and maintenance costs are low, as little power is required to turn the discs, and head loss throughout the plant is minimal.

Little operator attention is required.

2.2 Historical Development

According to Hartmann (1960) the original concept of the RBC should be credited to Travis, who, in 1901, tried to increase the efficiency of his "Hydrolytic Tank" by hanging thin wooden strips, called colloid catchers, in the settling compartment of his unit. Solids accumulated on the slats, but did not always slough off, thus causing partial clogging of the unit.

Doman (1929) presented results which were obtained from the operation of what was probably the first RBC to resemble modern units. The RBC consisted of fourteen 20 gage galvanized iron plates (16 inches in diameter) which were

rotated in a 4.75 gallon, semi-circular tank. (At that time plastic materials were not available.) Doman obtained organic removal efficiencies characteristic of conventional aerobic treatment processes and identified research needs similar to those being investigated today. (i.e. optimum retention times, speed of rotation, surface area effects and temperature effects). Concurrently, Buswell (1929) reported on a system which he called the "Biologic Wheel" and cited as advantages that:

1. the area requirements were about 1/10 of that needed for a trickling filter,
2. the power costs were low as compared to the activated sludge process, and
3. nitrification was obtained.

The Great Depression and World War II delayed further research until Hartmann and Popel began investigating the process at the Technical University of Stuttgart, in 1955. Hartmann (1960) described the operation of two experimental immersion drip filter disc plants. The paper outlined performance characteristics and economics for their use as a sewage treatment system. Subsequent papers by Popel (1964) and Hartmann (1965) indicated that disc filters had become moderately well established on the continent. Lohr (1967) described a prototype RBC plant which used power from the sewage flow to rotate the discs. Reinisch (1969) gave theoretical reasons to support the claims that rotary disc plants were more economical to operate than conventional treatment methods.

In the U.S.A. and Canada, much of the early research was concerned with the use of RBC's for the treatment of wastewaters from small communities or industrial plants (Antonie et al, 1969, 1970). Using designs similar to those

used in Germany, these plants were constructed to treat either primary or septic tank effluents. Torpey et al (1972) experimented with multiple staged disc units which included a final stage exposed to light in order to grow algae and thereby remove inorganic nutrients from the sewage.

More recently, Reimer et al (1976) compared the performance of several pilot scale wastewater treatment units. Under nitrifying conditions he concluded that the RBC did not produce as stable an effluent as an activated sludge unit.

Commercial marketing of the RBC began in Europe in 1959 by J. C. Stengelin Ltd., Tuttingen, W. Germany, who now have licences in several countries. By 1973 there were more than 1,000 installations in Europe (primarily West Germany, France and Switzerland) treating domestic, industrial and mixed wastewaters. Treatment capacities varied from that generated by a single residence to 45.4 kg BOD₅ removed per day (Beak, 1973).

Allis - Chalmers Ltd. initially introduced the RBC system to North America in the mid - 1960's and in late 1970, Autotrol Corporation acquired the patents, inventories and contracts from Allis - Chalmers. Other companies now operating in North America include Bio Disc (Ames Crosta Mills Ltd), Rotordisk (CMS Ltd), Euromatic Bio Drum (European Plastic Manufacturing Company), the Rotating Biological Surface (Geo. A. Hormel & Co.), Enviroidisc (Enviroidisc Corp.), and the Rotating Disc Biological Reactor (Environmental Dynamics Corp.).

2.3 Carbon Oxidation

2.3.1 Design

Hartmann (1965) provided the first detailed account of RBC design practices used in Germany. These designs were introduced to South Africa by Pretorius (1973) and to North America by Steels (1974). The design methods were based on hydraulic loading, inlet BOD₅ concentration, and areal BOD₅ loading, for a town population of 10,000. Multiplication factors were provided for communities of different sizes.

Joost (1969) suggested a steady state design equation based on the assumption that the biochemical reaction taking place is concentration dependent, following a first order equation of the type

$$\% \frac{\text{BOD Reduction}}{\text{Stage}} = K \times C^a \times R^b \times T^c \times \Gamma^d \quad \dots \quad (1)$$

where: K is the treatability constant,

C is the concentration of the waste material,

R is the physical configuration constant,

T is the wastewater temperature,

Γ is the residence time, and

a, b, c, d are partial regression coefficients.

Unfortunately, data were not provided for the constants a, b, c, d or K. The above equation was solved using multiple regression analysis by Weng and Molof (1974), for a nitrifying system. The results indicated that influent loading,

concentration and flow had an effect on effluent quality but no indication was given as to the relative importance of each parameter.

In North America, Antonie (1970) presented data which suggested that BOD_5 removal was a "first order" reaction with respect to BOD_5 concentration. "First order" removal kinetics implies that a given percentage BOD_5 removal is possible regardless of inlet BOD_5 concentration or loading. Therefore, Antonie introduced RBC treatment facility designs based solely on hydraulic loadings and percentage removals. This approach is analagous to the volumetric loading rates once used for activated sludge design and hydraulic loading rates for trickling filter design.

Although many investigators believed hydraulic flow rate to be the determining factor in BOD_5 or COD removal in fixed film systems (i.e. trickling filters), Cook and Kincannon (1971) presented data which indicated that the important design parameter for trickling filters was the organic loading. They recommended that any comparison between trickling filter be made on a $g/d-m^3$ ($lbs/d-1000 ft^3$) basis. These results led to a great deal of controversy among researchers as to what the primary design criterion for RBCs should be and how experimental data should be reported.

Since the paper by Antonie (1970), the RBC manufacturers have all tended to use areal hydraulic loading as their primary design basis. It was not until 1976 that data were published indicating that areal mass loading was possibly a better design criterion. Stover and Kincannon (1976) operated a RBC pilot plant on slaughterhouse wastewater. They found that a plot of mass loading versus mass removal followed Monod type kinetics for COD removal. The plot showed less variability than a similar one using hy-

draulic loading. A limiting removal of 22.5 g COD/d-m² was obtained for loadings greater than 50 g COD/d-m².

2.3.2 Performance

Because the rotating biological contactor is a relatively new wastewater treatment process, many of the papers published on RBC research are devoted to describing only their effectiveness in treating industrial and municipal wastes, in terms of percentage removals. Pretorius (1971) found that a maximum removal rate of 0.49 g COD/d-g biomass was achievable in his 9 disc RBC system. At the same time Wells (1971) found that for an influent BOD₅ concentration of 250 mg/l, it was possible to improve BOD₅ removal efficiency from 50% to 83% simply by decreasing the flow rate from 325.9 l/d-m² (8.0 U.S. gal/d-ft²) to 163 l/d-m² (4.0 U.S. gal/d-ft²). The results of Pescod (1972) seemed to verify the "first order" kinetic results obtained by Antonie (1970). When viewed on a stage by stage basis, Pescod was able to obtain 95% COD removal at a loading of 4 kg COD/d-m² for organic wastes near 1000 mg/l.

Bruce et al (1973) recommended a maximum daily areal load of 6 g/d-m² (1.2 lb. BOD/d-1000 ft²) in order to obtain the standards set in the U. K. (30 ppm suspended solids, 20 ppm BOD₅). The MOE 1974 report recommended that a loading of 20 to 25 g/m²-d (1 lb BOD₅/d-1000 ft²) should be used to obtain 80 to 90% BOD₅ removal efficiency. EPA, on the other hand, simply suggested using an areal hydraulic loading of 61.1 l/d-m² (1.5 U.S.gal/d-ft²)) to obtain 87% BOD₅ removal or better.

2.3.3. Diurnal Variability, Hydraulic Pulses and Intermittent Flow

The problems peculiar to isolated sewage treatment plants serving small communities, summer camps and work camps are mainly related to highly variable flow conditions,

limited supervision and maintenance. Despite these problems there is normally a requirement that the plant will at all times produce a consistent effluent quality. All of the positive aspects of RBCs indicate that these units should be ideally suited for such applications. In addition, economic studies reveal that RBC construction costs increase linearly with size, suggesting that their use be limited to small treatment applications. (EPS Report No. 4-WP-73-4; Winkler, 1974; Antonie, 1976).

Popel (1964), Markii(1964) and Antonie (1970) investigated RBC effectiveness in situations where cyclic operations, shift work, holidays and weekends tend to create problems of operation. All three investigators reported an increase in BOD₅ or COD removal efficiency during cyclic operation. However, this was not confirmed by NCASI (1974) or Davis (1976) who found an immediate deterioration in organic removal efficiency during hydraulic pulses

Bruce et al (1973) subjected a RBC test unit to diurnal peak flow variations of 3/1/0.6 (maximum/average/minimum) in a rectangular wave pattern. The hydraulic pulse variation was considered to be a reasonable simulation of the flow pattern which would arise from a small community with synchronized activities such as schools and other institutions. The results of the study indicated that the RBC could not produce a stable effluent quality which would meet British effluent standards, although 90% BOD₅ removal was obtained. The report did not specify the influent organic loadings.

Three government institutions have recognized the potential of RBCs as a viable treatment alternative for small communities and initiated investigations into their use.

The EPA (1973) found the RBC process to be relatively well suited to summer camp application where sewage flow was low and fluctuated considerably. It should be noted however, that the hydraulic loadings were 4.9 to 48.9 l/d-m² (0.1 to 1.01 US gal/d-ft²) which are very conservative by manufacturers standards. The MOE (1974), EPS (1974), Bruce (1974) and Antonie (1970b) demonstrated that intermittent flow did not adversely affect organic removal efficiency, although they noted an initial increase in solids concentration where operations were resumed after a two-day stoppage in flow. Forgie (1974) operated a bio-disc unit at a five man workcamp in the Northwest Territories. The unit was subjected to loadings ranging from 4.9 to 19.5 g/d-m² BOD₅ (7.3 to 29.2 g/d-m² COD). Good percentage removals were obtained at all levels, although effluent concentrations varied considerably.

2.4 Nitrification

Nitrification of wastewaters may be achieved by using the activated sludge, trickling filter or RBC process operating under conditions favorable for the development of nitrifying bacteria. Nitrifying bacteria are strict autotrophs and are distinctly different from the heterotrophic bacteria responsible for the degradation of organic carbonaceous matter. In the RBC process, nitrification has been observed to occur only in the final RBC stages owing to differences in yield between the nitrifiers and heterotrophes (Torpey 1972). Antonie (1972a,1972b) noted that nitrifying bacteria can only begin to compete with the heterotrophes at BOD₅ concentrations of 30 mg/l or less in systems utilizing areal hydraulic loadings ranging from 4.9 to 49 l/d-m² (0.1 to 1 U.S. gal/d-ft²). Torpey (1972) and Weng and Molof (1974) found that the limiting BOD₅ concentration was closer to 15 mg/l and noted that this was equivalent to a COD of 50 mg/l.

2.4.1 Design

Antonie (1970) claimed that ammonia removal kinetics are a first order function of retention time in the RBC system. Design considerations similar to carbon oxidation systems were therefore developed, namely, to base the primary design criterion on the areal hydraulic loading to the unit.

First order removal kinetics were also demonstrated by Stover and Kincannon (1975). They investigated $\text{NH}_3\text{-N}$ removal at influent loadings of 0.56, 1.12 and 2.25 g $\text{NH}_3\text{-N}/\text{d}\cdot\text{m}^2$. Removal efficiency was shown to decrease from 100% to 51% in a semi-logarithmic fashion when the flow rate was increased four fold. The investigation did not distinguish between the effect of flow and mass loading. Weng and Molof (1974) demonstrated that loading and not flow rate should be the primary design criterion. Nitrification was shown to remain constant when the flow and concentration were varied while maintaining a constant mass loading. However, using designed experiments, Wilson (1975) could not determine if nitrification followed "zero order" or "first order" kinetics. Murphy et al. (1975) compared models based on "zero order", "half order" and "first order" kinetics. The model which was "zero order" with respect to TKN concentration was selected as the best able to describe the data.

2.4.2 Performance

As with carbon oxidation, most of the literature has been devoted to describing reactor performance in terms of percentage removal. Hao and Hendricks (1975) operated a pilot plant RBC in Columbus, Indiana and obtained 92% $\text{NH}_3\text{-N}$ removal of at an areal flow rate of 0.06 $\text{m}^3/\text{d}\cdot\text{m}^2$ (1.5 U.S. gal/d-ft²). An increase to 0.1 $\text{m}^3/\text{d}\cdot\text{m}^2$ (2.5 U.S. gal/d-ft²) significantly decreased nitrification. Influent $\text{NH}_3\text{-N}$

concentrations averaged 8.8 mg/l at the low flow rate and 10.8 mg/l at the high flow rate. Wilson (1975) obtained an average $\text{NH}_3\text{-N}$ removal rate of 20 mg/h- m^2 (0.10 lb/d-1000 ft^2) from his system with temperatures varying from 7 to 20°C. Variations in mass removal rate at different temperatures were related by an Arrhenius expression. Stover and Kincannon (1975) noted that $\text{NO}_3\text{-N}$ production decreased with a simultaneous increase in $\text{NH}_3\text{-N}$ and COD load. They concluded that the increased COD load must have caused the heterotrophs to multiply in the last RBC compartments, subsequently reducing overall nitrification rates.

2.5 Denitrification

In contrast to ammonia removal, complete nitrogen removal may be achieved by the biological nitrification-denitrification process.

Denitrification on a RBC has been demonstrated by Pretorius (1973) and Davis and Pretorius (1974). These papers described the use of an enclosed, partially submerged, anaerobic RBC unit to denitrify a wastewater. They overcame two problems associated with conventional submerged biological bed denitrification processes:

- a) clogging of bed voids by the active biomass, and
- b) adherence of nitrogen gas to the sloughed biomass.

Shearing of the excess biological growth prevented clogging and exchange of the nitrogen gas to the wastewater and subsequently to the atmosphere prevented adherence. The maximum rate achieved was 250 mg $\text{NO}_3\text{-N}$ reduced/ $\text{m}^2\text{-h}$. Below 10°C, a severe inhibition of denitrification occurred.

Soyupak (1976) operated a submerged 0.5 metre RBC to denitrify sewage. He found that denitrification could be

expressed as zero order reactions. Denitrification rates were higher than suspended growth or packed column systems. Temperature dependency of the denitrification reaction rates were independent of flow rate in the range of 2.5 to 4.0 l/min and was empirically described using an Arrhenius relationship.

2.6 Steady State Modelling

Success at steady state modelling of RBCs based on empirical, semi-empirical or completely theoretical models is limited. Removal characteristics of conventional treatment systems have been described by biological growth kinetics and the mixing characteristics of the reactor. Using such an approach, models for rotating biological contactors must also incorporate parameters to describe the active biomass thickness, disc rotational speed, available surface area, system geometry, and diffusion coefficients for oxygen and substrate transfer. The methods used to describe these characteristics are at best, estimates, and therefore introduce potential weakness in the models.

Kornegay (1972) developed a kinetic expression for RBC substrate removal assuming Monod kinetics and a completely mixed reactor.

$$F(S_0 - S_1) = 2 PN \pi (r_o^2 - r_\mu^2) \left(\frac{S_1}{K_g + S_1} \right) \quad \dots(2)$$

where: F is the flow rate,
 S_0 , S_1 are the influent and effluent substrate concentrations,
 N is the number of discs,
 r_o is the total disc radius,
 r_μ is the submerged disc radius,
 K_g is the saturation constant,

P is the area capacity constant equal

$$\text{to } \frac{1}{Y_g} \mu_{\max} X_f d,$$

Y_g is the apparent yield of fixed film organisms,

μ_{\max} is the maximum specific growth rate,

d is the active microorganism thickness, and

X_f is the unit mass of biological film.

Kornegay assumed values for the parameters in the model and predicted removal capabilities for a number of theoretical design situations. The information gained from this study was used to determine RBC treatment capacities and design requirements. However, without model calibration using experimental data, the results can only be used to demonstrate a methodology.

A more recent investigation into the kinetic response of RBC's was conducted by Friedman et al (1976). Experimental data were used to calibrate a conventional plug flow "first order" model and an empirical model based on mass transport concepts.

Plug Flow Model

$$C_e = C_b e^{-K_e t} \quad \dots \quad (3)$$

Mass Transport Model

$$\begin{aligned} K_m \left(\frac{1}{C_b} - \frac{1}{C_{bi}} \right) + \text{Ln} \left(\frac{C_{bi}}{C_b} \right) \\ = \frac{fh K_o^* A_s}{V_1} \theta = \frac{K'' \theta}{V_1} \quad \dots \quad (4) \end{aligned}$$

where:

C_{bi} , C_b , C_e are the influent, soluble approach, and effluent substrate concentrations,
 K_e is the reaction rate constant,
 t is the overall time in the system,
 K_m is the half saturation constant,
 f is the proportionality constant,
 h is the effective biomass depth,
 K_o^* is the maximum areal removal rate,
 A_s is the submerged surface area per disc face or stage,
 K'' equals $f \cdot h \cdot K_o^* \cdot A_s$,
 θ is the average hydraulic retention time per disc face, and
 V_1 is the tank liquid volume per disc face.

A general method was presented for obtaining K'' from the experimental data. The models were then compared to data collected from independent studies. The mass transport model was shown to be superior in predicting effluent quality on a stage by stage basis.

Grievess (1972) incorporated a mass transport approach and Monod kinetics to develop a steady state model for rotating disc systems. The model is:

$$\frac{C_b}{C_o} = \frac{1}{1 + \frac{N}{F} [P_1 \cdot A_s + P_2 \cdot (\text{rpm}) \cdot (1 - \bar{e}^{P_1(A_a)/(P_2)(\text{rpm})})]} \dots (5)$$

where: C_b , C_o are the concentration of the limiting substrate in the bulk liquid, and in the raw feed,

N is the number of discs,

F is the flow rate,

A_s is the area of one disc which is exposed to the atmosphere,

A_a is the area of one disc submerged in the bulk liquid,

P_2 is the quantity of liquid film attached to the biological film which enters the reactor per unit time,

P_1 is defined as
$$\frac{K_2 K_1}{1 + K_1}$$

K_2 is the liquid film coefficient,

K_1 is defined as
$$\frac{(\hat{\mu})(X)(\Delta z)}{(y)(K_c)(n)(K_L)}$$
,

$\hat{\mu}$ is the maximum specific growth rate,

X is the organism concentration,

Z is the active depth of biological film,

y is the yield coefficient,

K_c is the saturation constant in the Monod equation, and

n is the efficiency factor.

The model effectively described effluent concentrations obtained by a number of independent investigations for influent substrate concentrations less than 50 mg/l TOC.

2.7 Dynamic Modelling

The steady state models discussed previously have been used to predict the steady state behaviour for either RBC design or operation. Use of steady state models is inconsistent with the variable loading of most biological waste treatment systems. Parameter values which have been obtained for these models will not describe the temporal relationships between influent and effluent when the input to the RBC is highly variable. An estimate of the stability of RBCs can only be obtained by using techniques which elucidate the responses to non-steady operation. This fact was recognized by Grieves (1972) who developed a dynamic mathematical representation of fixed film reactors. The dynamic model was developed from theoretical and empirical considerations of a fixed biological film attached to a rotating surface, and is represented by the following differential equations:

$$\begin{aligned} \frac{dC_{1,1}}{dt} = & \frac{K_L}{\Delta Z_1} [C^* - C_{1,1}] - D \frac{C_{1,1} - C_{2,1}}{\frac{1}{2}(\Delta Z_1 + \Delta Z_2)} \\ & - \frac{(\hat{\mu})(X)(C_{1,1})}{(Y)[K_C + C_{1,1}]} \end{aligned} \quad \dots (6)$$

For L = 2 through 5, M = 1

$$\begin{aligned} \frac{dC_{L,1}}{dt} = & \frac{D}{\frac{1}{2}\Delta Z_L} \frac{C_{L-1,1} - C_{L,1}}{\Delta Z_{L-1} + \Delta Z_L} - \frac{C_{L,1} - C_{L+1,1}}{\Delta Z_L + \Delta Z_{L+1}} \\ & - \frac{(\hat{\mu})(X)(C_{L,1})}{(Y)[K_C + C_{L,1}]} \end{aligned} \quad \dots (7)$$

For $L = 6, M = 1$

$$\frac{d C_{6,1}}{dt} = \frac{D}{\frac{1}{2}\Delta Z_6} \frac{C_{6,1} - C_{5,1}}{\Delta Z_5 + \Delta Z_6} - \frac{(\hat{\mu})(X)(C_{6,1})}{(Y)[K_C + C_{6,1}]} \quad \dots (8)$$

For the liquid film,

$$\frac{d C_{LF,1}}{dt} = - \frac{K_L}{\delta_L} [C_{LF,1} - C_{1,1}] \quad \dots (9)$$

For the bulk liquid,

$$\frac{d C_b}{dt} = F [C_o - C_b]/V_b - F_f [C_b - C_{LF,Q-1}]/V_b - (KL)(A)[C_b^* - C_{1,1}]/V_b \quad \dots (10)$$

- where:
- L is an element at any depth in the active biological film of thickness ΔZ_L ,
 - M is the angular position of the liquid film on the biological disc,
 - $C_b, C_o, F, \hat{\mu}, X, Y, K_C$ have been previously defined in equation 5,
 - D is the substrate diffusivity,
 - $C_{L,M}$ is the limiting substrate concentration in element L, M,
 - K_L is the liquid film coefficient,
 - C^* equals the $C_{LF,M}$ if the element is in the reactor atmosphere,

- C^* equals C_b if the element is submerged in the bulk liquid,
- A is the area in the plane perpendicular to the direction of diffusing limiting substrate,
- δ_L is the thickness of an element LF in the liquid film,
- V_b is the volume of bulk liquid in the reactor, and
- $C_{LF,1}$ is the concentration of the limiting substrate in the liquid film.

The equations were solved using an analog computer. Values for the parameters used in the model were obtained from literature sources and laboratory data. The adequacy of the model was substantiated with dynamic data generated from two laboratory rotating disc reactors.

2.7.1 Linear Dynamic Stochastic Modelling

The solution of the dynamic model presented by Grieves (1972) is both tedious and time consuming. A simpler mathematical model that would describe the system is desirable. The need for a simple dynamic model for fixed-film systems is even more evident when it is considered that future treatment facilities may be automatically controlled using computers. For successful automatic control, predictive knowledge of the system response to dynamic inputs is necessary.

Time series analysis is a viable alternative to be considered as a method of describing the dynamics of a system, providing insight into the significant influent

parameters affecting effluent quality, and forecasting future effluent quality for various influent situations. In addition, once the dynamic transfer function-noise models are developed, they are ideally suited for control purposes. Because time series analysis is simply a statistical approach relating influent and effluent parameters of actual experimental or operational data gathered on a discrete time basis, the model may be updated as more information about the system becomes available.

MacGregor (1975) and Berthoeux et al (1976) successfully developed time series models relating input BOD₅ data to effluent BOD₅. Tan (1975) did not observe any significant dynamic effects in effluent TOC or suspended solids concentration, but did observe significant relationships for effluent NO₂-N + NO₃-N concentration for an activated sludge plant. Up to date literature reviews of dynamic modelling of suspended growth systems were provided by Tan (1975) and Sutton (1976). The state of the art of sewage treatment plant control was reviewed by Olsson (1976).

MacGregor (1975) recommended the use of designed input sequences in order to eliminate the high degree of correlation naturally present in municipal sewage (i.e. flow, organic carbon concentration and ammonia concentration). Sutton (1976) successfully used this design suggestion and was able to describe the dynamic nature of combined and separate nitrifying suspended growth systems. Dynamic models relating total Kjeldahl nitrogen (TKN) concentration to TKN effluent concentration and variations in effluent NO₂-N + NO₃-N concentration to TKN input concentration plus input organic loading (TOC) were developed.

2.7.2 The Box and Jenkins Method of Developing Linear Dynamic Stochastic Models

Data analysis for building dynamic transfer function-noise models involves an iterative procedure to identify tentative models, to estimate model parameters, and to test residuals to determine the adequacy of the fit. These models have the form:

$$Y_t = \frac{(w_0 - w_1\beta - w_2\beta^2 - \dots - w_s\beta^s)\beta^b}{(1 - \delta_1\beta - \delta_2\beta^2 - \dots - \delta_r\beta^r)} X_t + N_t \quad \dots (11)$$

where: X_t is the input deviation from the mean at time t ,
 Y_t is the output deviation from the mean at time t ,
 δ, w are the transfer function model parameters,
 r, s are the transfer function model orders,
 b is the system delay period,
 β is the backward shift operator, and

$$N_t = \frac{(1 - \theta_1\beta - \dots - \theta_q\beta^q)}{(1-\beta)^d (1 - \phi_1\beta - \dots - \phi_p\beta^p)} a_t \quad \dots (12)$$

where: N_t is the sequence of disturbances in the output which is not explained by X_t ,
 θ, ϕ are the noise model parameters,
 p, d, q are the noise model orders, and
 a_t is a white noise sequence of independent random variables with zero mean and constant variance

To identify potential transfer function models, the cross correlation function between influent and effluent variables is used. The cross correlation function between two input-output series (X and Y) separated by a constant lag, k , is given by:

$$\rho_{xy}(k) = \gamma_{xy}(k) / \sigma_x \cdot \sigma_y, \quad k = 0, \pm 1, \pm 2, \dots \pm k \quad \dots (13)$$

where: $\gamma_{xy}(k)$ is the cross covariance function between X and Y, and
 σ_x, σ_y are the standard deviations of the X and Y series.

The impulse response function, an equivalent form of the cross correlation function, can also be used to interpret the data. Analysis of the cross correlation function or the impulse response function allows the identification of potential transfer function models, the model orders (r,s), the delay period (b), and the initial estimate of the parameter values (ω, δ). The autocorrelation functions of the residuals of the fitted transfer function models are used to identify potential noise models, N_t . Once the parameters of the combined transfer function-noise models are estimated efficiently by using a non-linear least squares technique, model adequacies are verified through diagnostic checks.

Diagnostic checks are made by calculating and examining the results of cross correlations between input and residuals, and by calculating the "S" statistic, which is then compared to the chi-square distribution (X^2) with $k - r - s$ degrees of freedom:

$$S = m \sum_{k=0}^k r^2_{x'\hat{a}}(k) \quad \dots (14)$$

Noise model verification is obtained by autocorrelating the residuals of the transfer function noise model. A general lack of fit test is obtained by calculating the "Q" statistic which is compared to the chi-square distribution (X^2) with $k - p - q$ degrees of freedom:

$$Q = m \sum_{k=1}^k r^2 \hat{\hat{a}}(k) \quad \dots (15)$$

A more detailed description of the methods used are provided in Box and Jenkins (1976).

3. EXPERIMENTAL EQUIPMENT AND PROCEDURES

3.1 Pilot Plant Design and Operation

One 0.5 m RBC and one 2.0 m full scale RBC were used during this study. The units were obtained from Autotrol Inc., Milwaukee, Wisconsin. The 0.5 m unit provided 23.23 m² (250 ft²) of available disc surface area, while the 2.0 m unit had 733.93 m² (7900 ft²). Rotational speeds of the large and small unit were fixed at 3 RPM and 13 RPM respectively, thereby providing a peripheral tip velocity of 0.34 m/s (1.12 ft/s).

Feed to the half metre unit was provided by feed scoops or by a variable speed positive displacement pump, depending on the flow rates required. Feed to the 2 m unit was provided by variable speed positive displacement pumps. The influent raw feed used during the course of the experiments was degrittied raw sewage from the Burlington Sewage Treatment Plant.

Figure 3.1 presents a schematic diagram of the system used and the identification codes for the units. The 0.5 m unit (0.5 M1) was operated indoors for a period of 2-1/2 months and for the remaining time, 0.5 M1 was run in parallel with the 2 m unit (2.0 M). The biodiscs were covered with hoods to provide insulation and protection from the natural environment (wind, rain, sunlight). All experiments were conducted with the hoods in place.

3.2 Experimental Plan and Start-Up

As previously discussed, the RBC has many inherent positive features which makes its use attractive in small community, workcamp and summercamp applications. Sewage generated by such sources is highly variable in both flow and concentration, and it is desirable to assess the per-

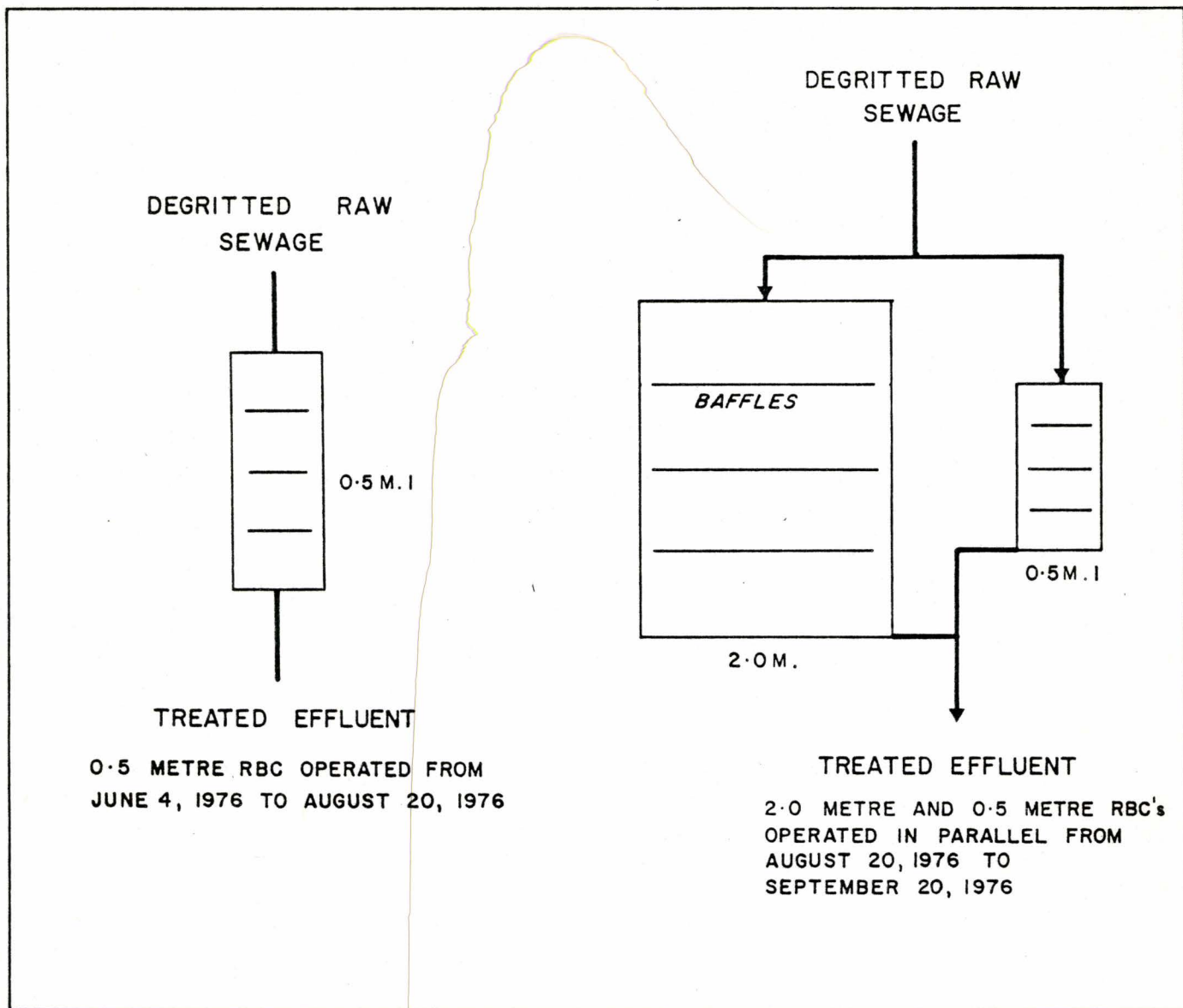


FIGURE 3.1
PROCESS SCHEMATIC

formance of an RBC under such conditions. Further, there is also a need to examine whether hydraulic or organic loading is the primary design criterion. To satisfy these goals, experiments were designed to permit the use of time series analysis to evaluate the dynamic response of the RBC. The experimental programme was developed to provide data in two modes of operation; carbon oxidation and carbon oxidation plus nitrification.

A summary of the experimental programme is provided in Table 3.1. The experimental designs are discussed in Sections 3.3 and 3.4.

Prior to start-up of each unit, dye studies were performed to characterize the mixing regime of the RBCs. (Series A1, A3, A5). This data also provided an indication of peak residence times which was used in the development of the discrete sampling programme. Results from the dye studies are presented in Appendix D.

Acclimation took place during June, 1976 (series A2) with start-up of 0.5 M1 on June 1. The feed rate was set at approximately 0.8 l/min to allow for the growth of nitrifiers. Almost complete nitrification was achieved in two weeks with the high influent temperatures (28°C) obtained by passing the degrittied raw sewage through a heater. The heater was bypassed after two weeks and influent temperatures dropped to normal (18 to 20°C).

The 2.0 metre biodisc (2.0 M) was started up on July 16, 1976 in the carbon oxidation mode, at flow rates of approximately 100 l/min. Acclimation lasted until August 5, 1976. The 2.0 metre unit was not used in this study until September 7, 1976.

Runs in series B1 and C1 extended from June 29 to July 22, 1976. They were followed by an acclimation period

Table 3.1Summary of Experimental Programme

<u>Operational Mode</u>	<u>Experiment</u>	<u>Run No</u>	<u>Type of Sampling</u>
Carbon Oxidation Plus Nitrification	Hydraulic Characterization 0.5 Ml	A1	Grab
Carbon Oxidation Plus Nitrification	Acclimation, 0.5 Ml	A2	Grab and 24 hour composites
Carbon Oxidation	Hydraulic Charac- terization, 2.0 M	A3	Grab
Carbon Oxidation	Acclimation, 2.0 M	A4	Grab and 24 hour composites
Carbon Oxidation Plus Nitrification	Diurnal Influent Variations, 0.5 Ml	B1	24 hour composites and hourly grab samples
Carbon Oxidation	Diurnal Influent Variations, 0.5 Ml	B2	24 hour composites and hourly grab samples
Carbon Oxidation Plus Nitrification	Dynamic Perfor- mance of 0.5 Ml	C1	hourly grab samples
Carbon Oxidation	Dynamic Perfor- mance of 0.5 Ml operated in parallel with 2.0 m	C2	30 minute grab samples
Carbon Oxidation	Dynamic Perfor- mance of 2.0 M Operated in Parallel with 0.5 Ml	E2	30 minute grab samples

of 6 days before beginning series B2 (July 28, 1976 to August 9, 1976). Series C2 and E2 were conducted from September 7 to September 10, 1976 and concluded the experimental plan for this study.

3.3 Diurnal Flow Experiments (Series B1 + B2)

As the performance of rotating biological contactors was to be assessed for small community applications, the Series B experiments were designed to simulate the approximate diurnal flow and concentration variation typical of sewage from a small community. The 0.5 Ml was subjected to a sinusoidal flow input which attained max/avg/min peak flow ratios of 2/1/0.5, synchronous with the natural sewage concentration fluctuations of Burlington STP (Figure 3.2). Two average flow rates were employed, 65.1 l/d-m² (1.6 U.S. gal/d-ft²) for nitrification (Series B1) and 208.3 l/d-m² (5.1 U.S. gal/d-ft²) for carbon removal (series B2).

During the course of these runs, 24-hour flow weighted composites and hourly grab samples were obtained from both the influent and effluent of 0.5 Ml. The data was used to verify the adequacy of the dynamic transfer function-noise models which were to be developed from the Series C experiments, and also, to assess the performance of the RBC.

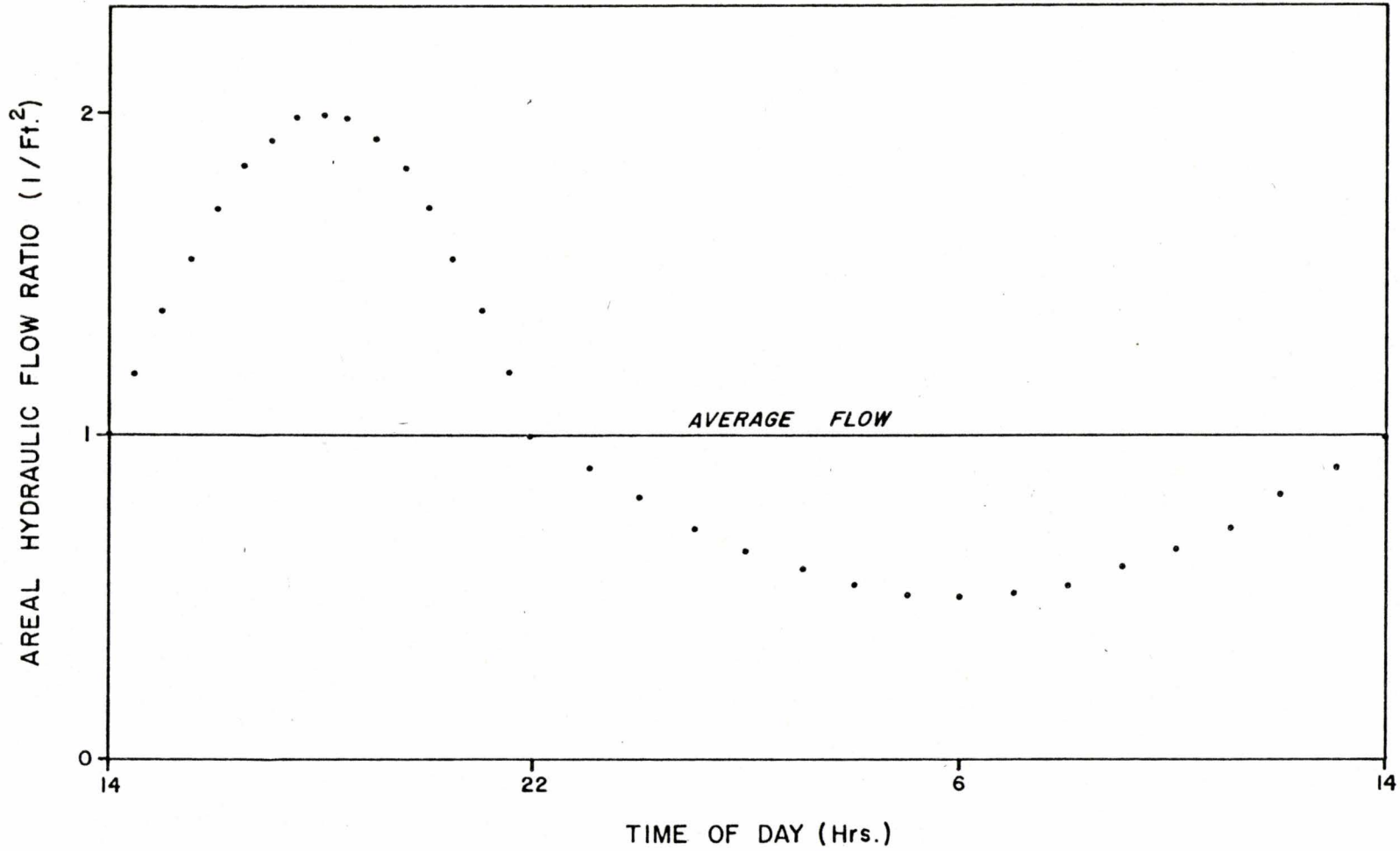
3.4 Dynamic or Non "Steady State" Experimental Design (Series C1, C2, E2)

The purpose of the dynamic experiments was to examine the performance of the 0.5 metre and 2.0 metre RBCs under variable influent conditions and to develop transfer function models which describe the dynamic behaviour of the system. In order to do so, experiments were designed and samples taken according to the following:

- a factorial design was used to separate the effects of the influent variables chosen;

FIGURE 3.2

PLANNED DIURNAL FLOW INPUT TO 0.5m RBC FOR PEAK/
AVERAGE/MINIMUM FLOW RATIO OF
2/1/0.5



- step changes in the influent variables were used to obtain the complete response spectrum;
- large numbers of paired samples (>100) were collected at discrete, equispaced time intervals; and,
- the sample interval was chosen consistent with the system response, which required prior knowledge of the residence time at various flow rates.

A two level, three parameter factorial design, presented in Table 3.2 and Figures 3.3 and 3.4, was selected. Three influent variables were chosen: hydraulic loading, filterable organic carbon concentration (TOC) and filterable total Kjeldahl nitrogen concentration (TKN). Experimental design levels are reported in Table 3.2. Loading boundaries (Table 3.2) were chosen to provide adequate system response while operating about the mean design level recommended for these units at municipal sewage treatment plants.

The design, using domestic sewage, required interference with the influent variables. This was accomplished by adding dextrose as an organic carbon spike, and ammonium chloride as a nitrogen spike. Two experiments were run to provide data for dynamic model building at average hydraulic loadings of 224 l/d-m² for carbon oxidation and 80 l/d-m² for nitrification. The units were run at the centre values of the parameters for two days prior to the start of experiments.

3.5 Feed Characteristics

The RBCs were continuously fed with normal degrittled wastewater from Burlington STP. Probability distributions of filtered influent COD, TOC, TKN and suspended solids are given in Figures 3.5, 3.6, 3.7 and 3.8. A correlation of TOC with filtered BOD₅ is given in Figure 3.9.

Table 3.2

Design Levels for Experiments C1, C2 and E2Carbon Oxidation Mode

<u>Influent Variables</u>	C1	C2 and E2	
		<u>Operating Value</u>	<u>Design Level</u>
Hydraulic Flow Rate L/Day/m ²	120	305	+
	40	143	-
Filtrable OC mg/l	50 to 60	80 to 90	+
	20 to 30	50 to 60	-
Filtrable TKN mg/l	50 to 60	25	+
	20 to 30	10	-
TOC Loading G/Day/m ²	6.6	26	+
	1	8	-
TKN Loading G/Day/m ²	6.6	7.6	+
	1	1.4	-

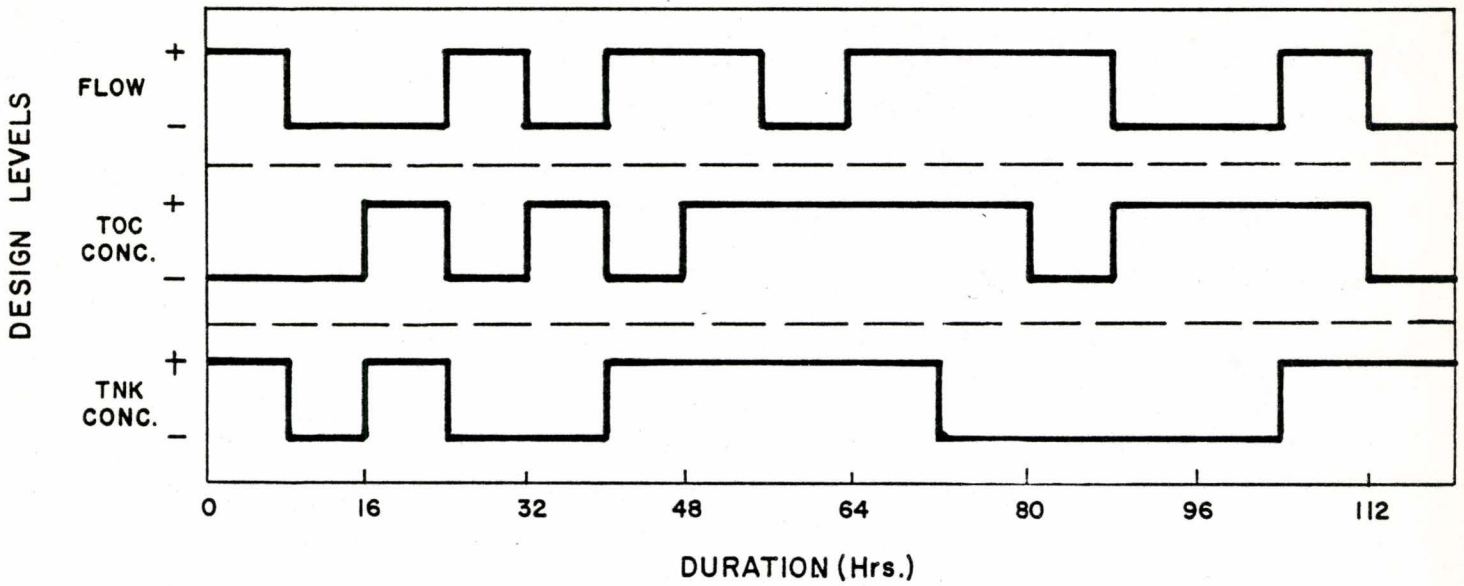


FIGURE 3.3
 EXPERIMENT C1 DESIGN, CARBON OXIDATION
 PLUS NITRIFICATION MODE

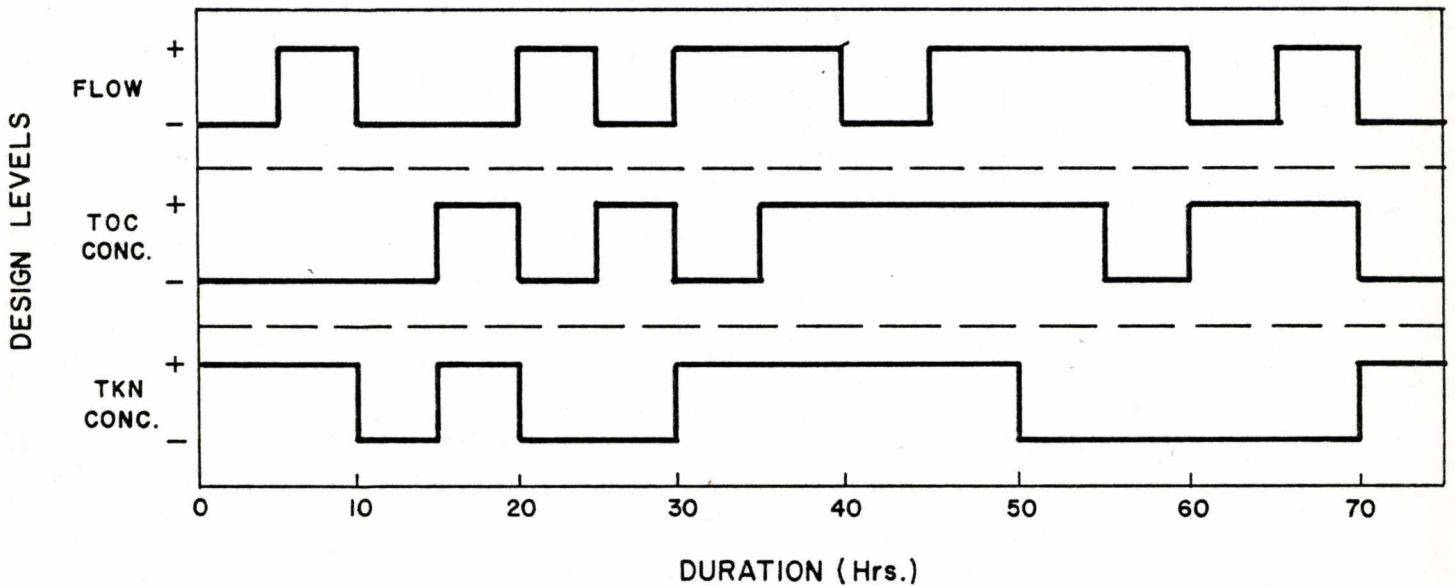


FIGURE 3.4
 EXPERIMENT C2 & E2 DESIGN
 CARBON OXIDATION MODE

FIGURE 3-5

FILTERABLE COD RAW WASTEWATER CHARACTERISTICS

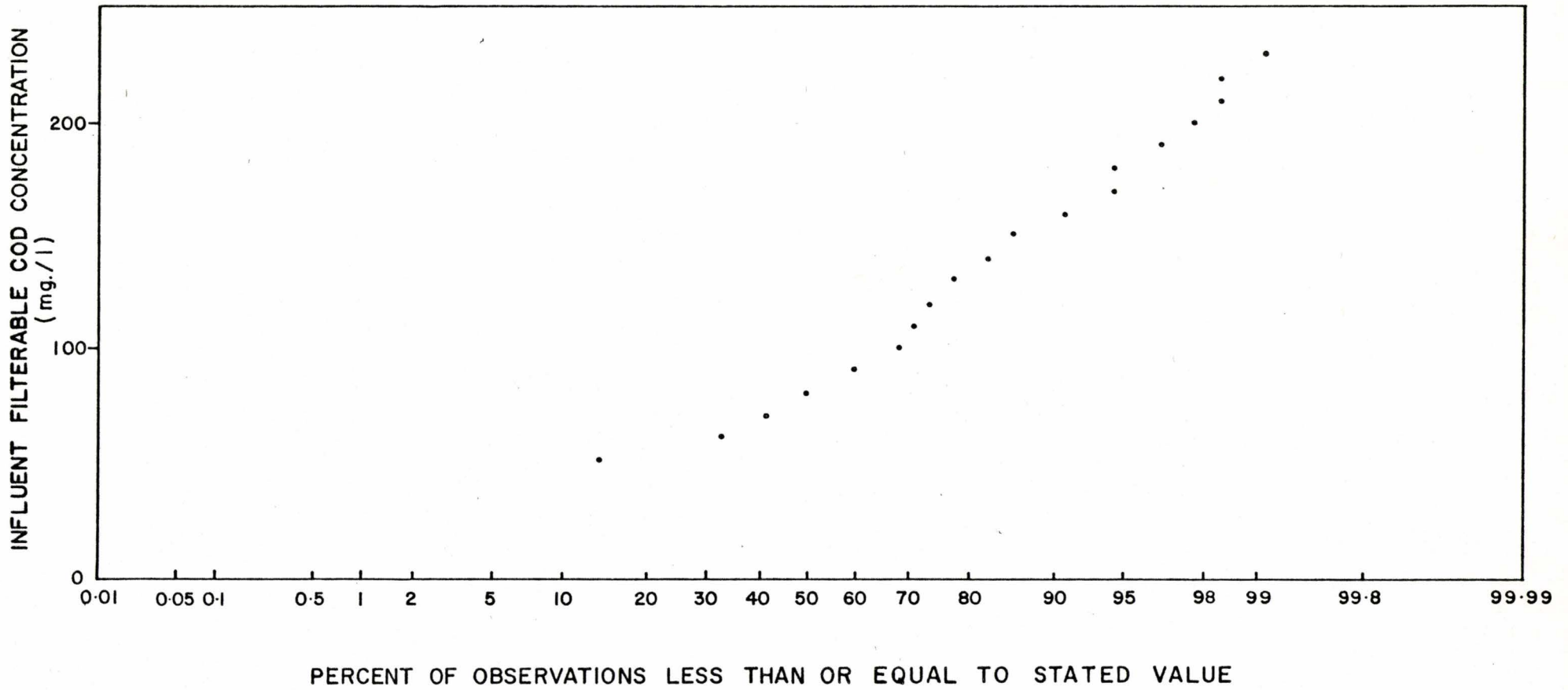


FIGURE 3-6

FILTERABLE TOC RAW WASTEWATER CHARACTERISTICS

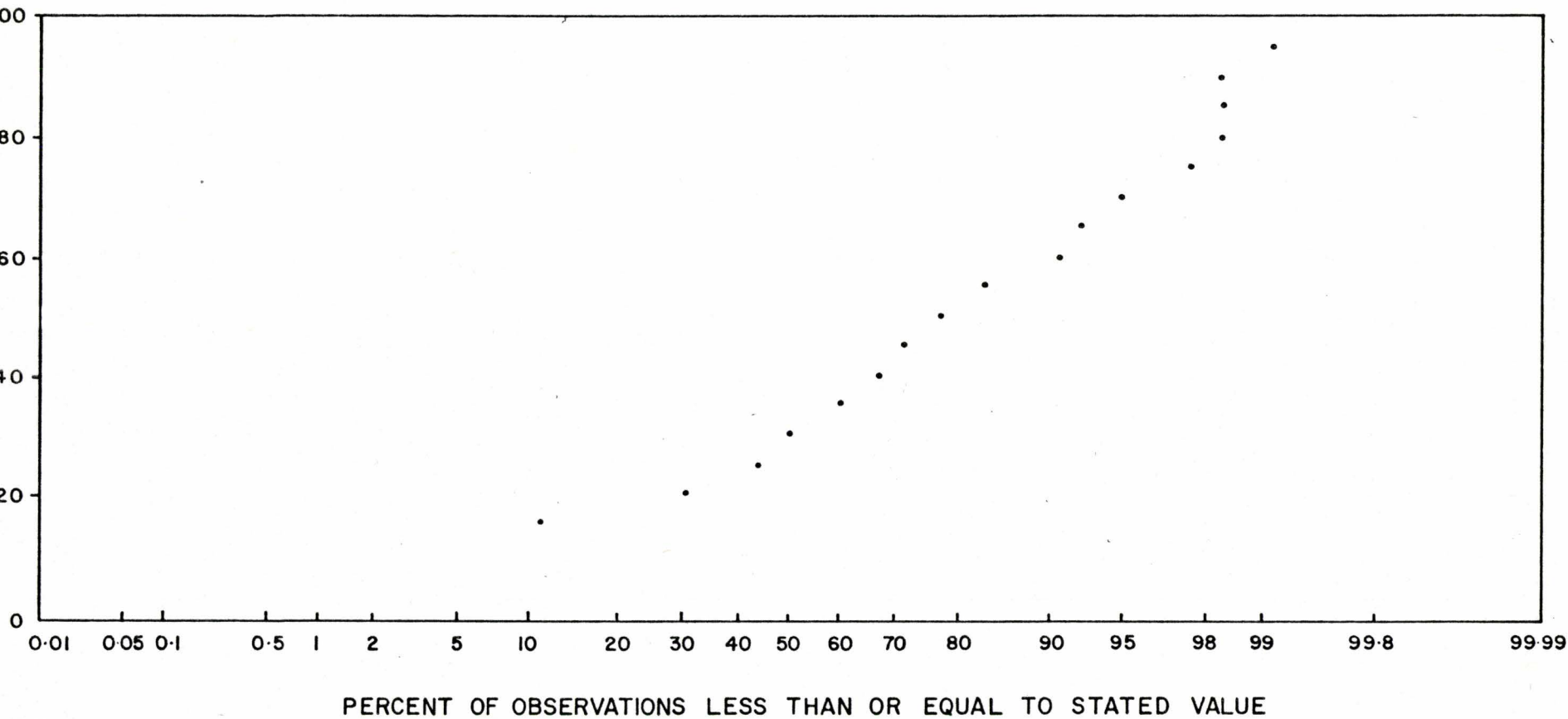


FIGURE 3-7

FILTERABLE TKN RAW WASTEWATER CHARACTERISTICS

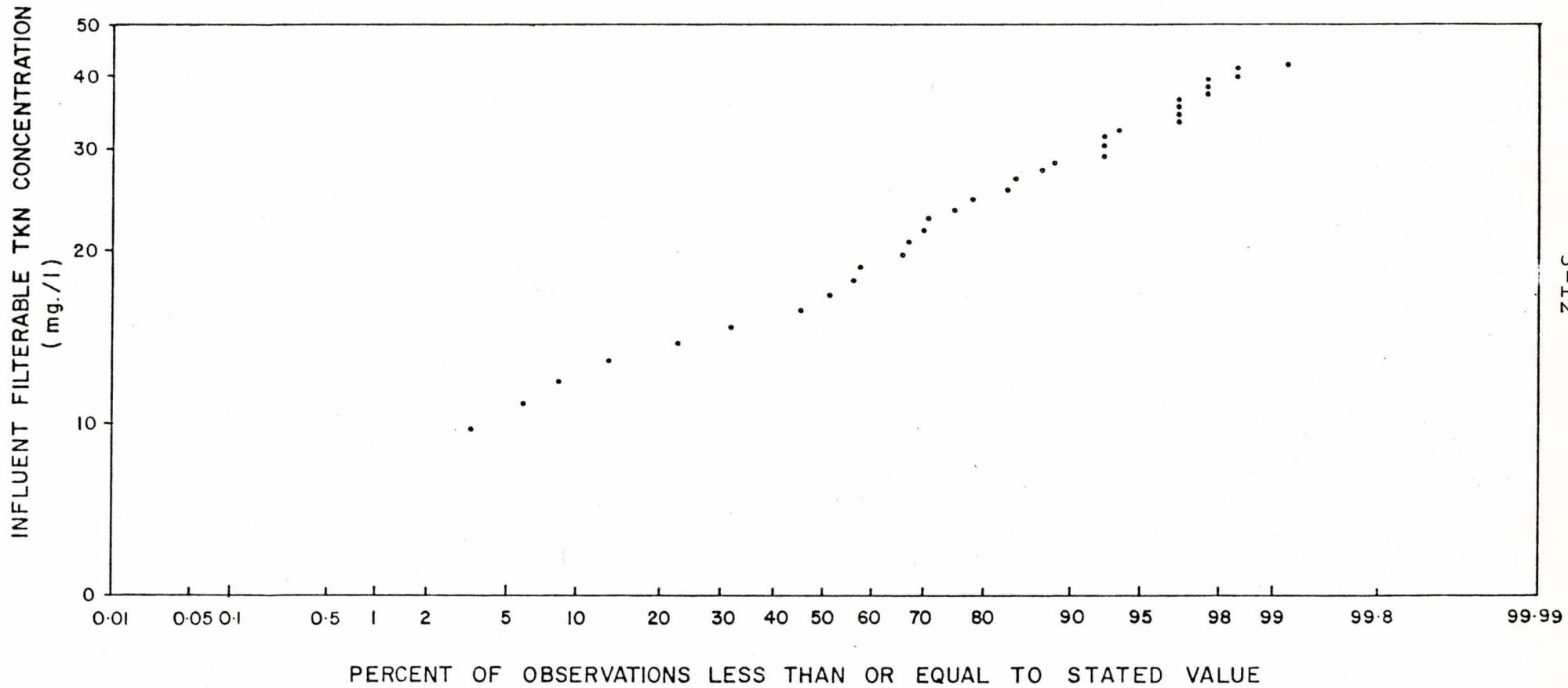


FIGURE 3-8

SUSPENDED SOLIDS RAW WASTEWATER CHARACTERISTICS

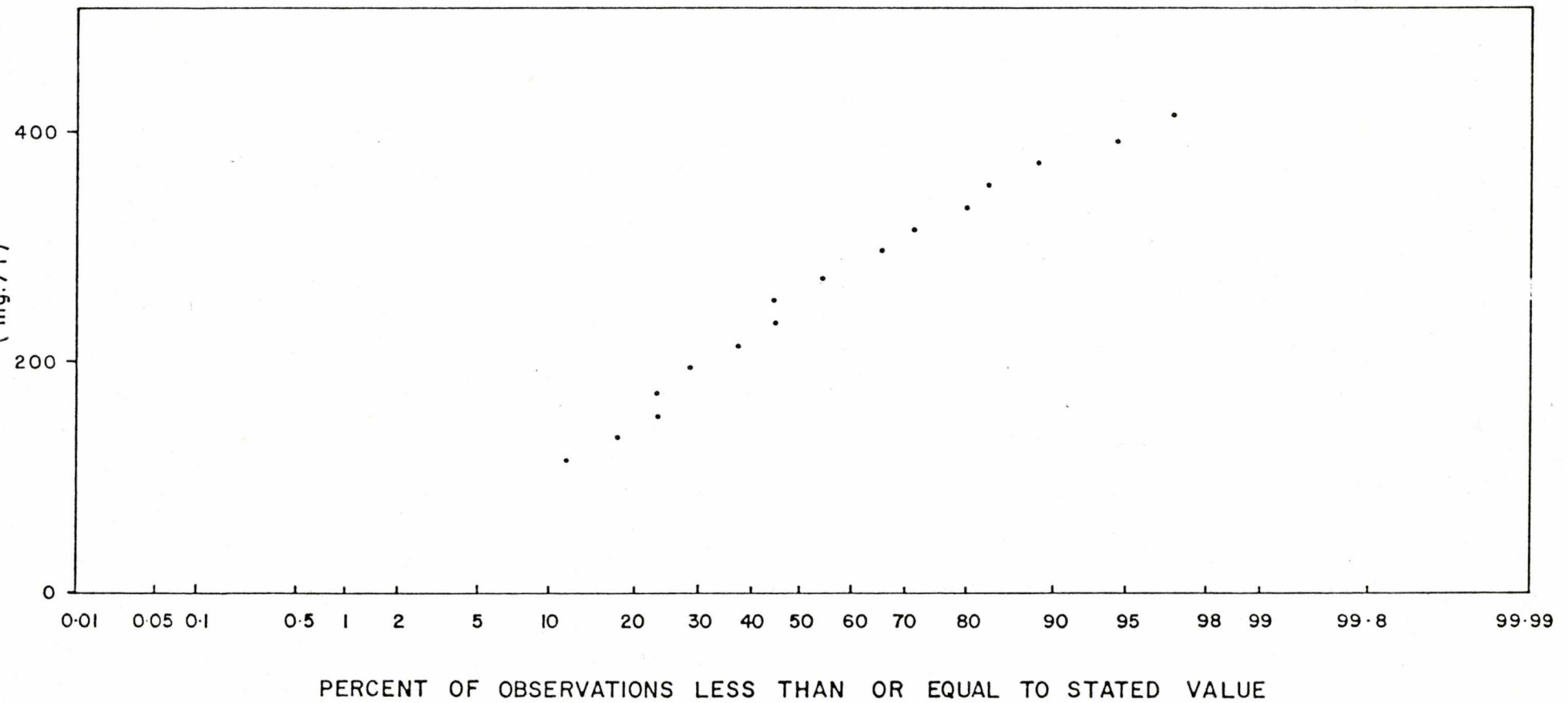
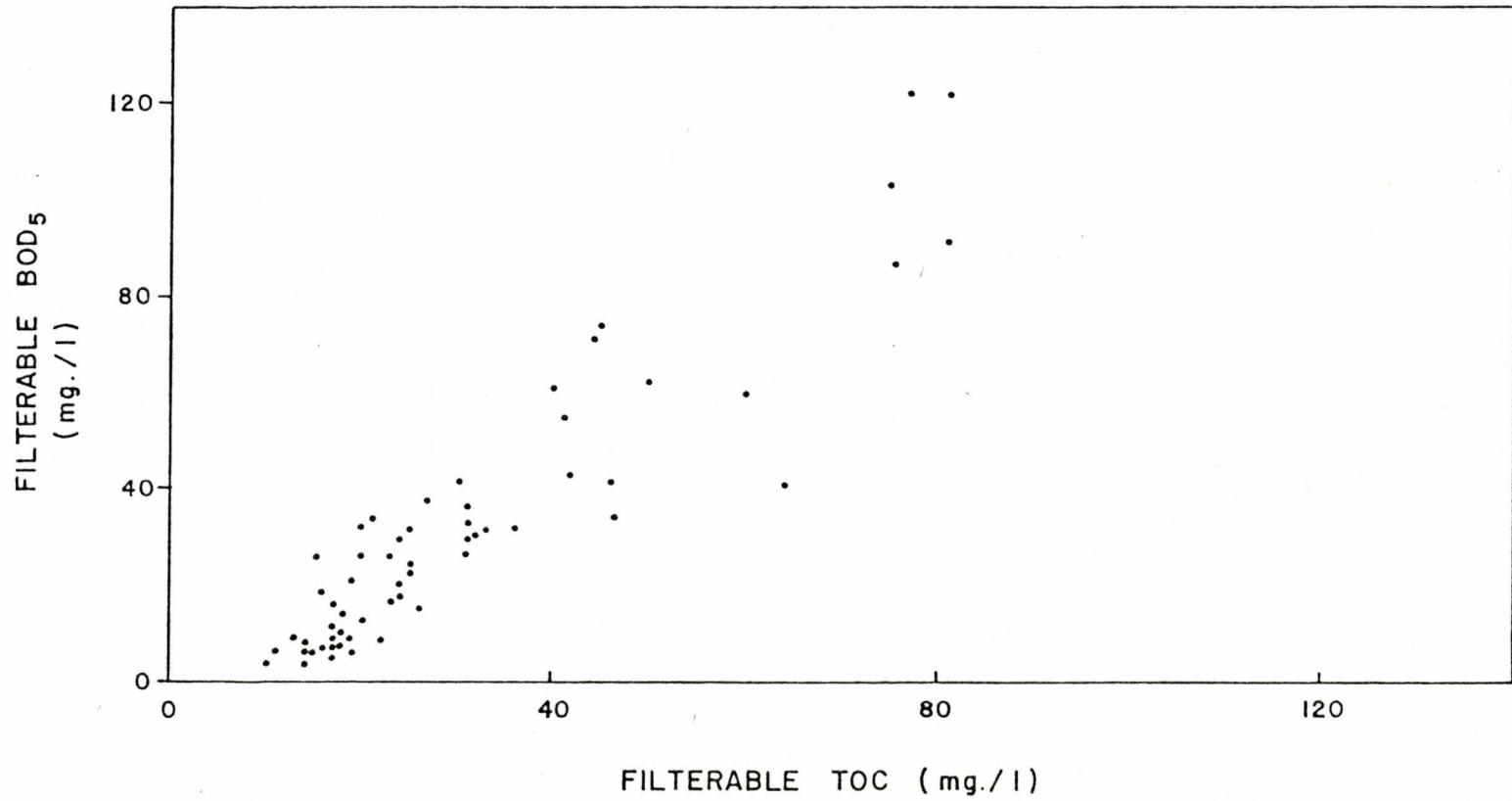


FIGURE 3.9

CORRELATION OF FILTERED BOD₅ WITH FILTERED TOC

SUMMER OF 1976



3.6 Sample Preparation and Analyses

Samples for total organic carbon (TOC) analyses were filtered through 0.45 micron Gelman glass fibre filters followed by acidification to pH 2 with concentrated hydrochloric acid. Filtered COD, $\text{NH}_3\text{-N}$, $\text{NO}_2\text{-N}$, $\text{NO}_3\text{-N}$ and TKN samples were frozen along with all BOD samples. Unfiltered COD and TKN samples were acidified to pH 2 with sulphuric acid before storage with the TOC samples at 0°C to 5°C. During the dynamic runs, the filtered COD samples were prepared in the same manner as the TOC samples. All samples were stored in polyethylene bottles.

Further details of the analytical procedures are given in Appendix B.

4. RESULTS AND DISCUSSION

4.1 Dynamic Time Series Data

The designed and observed influent concentration, effluent concentration, and flow rate for experiments C1, C2 and E2 are presented in Figures 4.1, 4.2 and 4.3. The flow rates were obtained by direct measurement during the experiment and were identical to the design levels. All experimental raw data is presented in Appendix A.

Good agreement is indicated between the designed input sequence and the measured influent concentrations, except during the first ten hours of the carbon removal dynamic experiments (Figures 4.2 and 4.3), when the effluent TKN concentrations frequently exceeded influent TKN concentrations. Dilution water was used during this period to reduce influent TOC concentration according to the design. Inadequate mixing was achieved before the influent sample was taken, causing an underestimation of the actual influent concentrations. These data were eliminated before further analysis was undertaken.

An effluent TOC response to the influent TOC and flow fluctuations is observed (Figures 4.2 and 4.3), but it is not apparent which influent variable caused the variation in effluent TOC concentration. The sensitivity of the nitrification process to input variations is apparent in Figure 4.1. Analysis of the data by time series methods should allow an assessment of the system response to hydraulic, organic and inorganic loading variations.

FIGURE 4-1
INPUT AND RESPONSE OF 0.5m RBC, CARBON OXIDATION
PLUS NITRIFICATION MODE

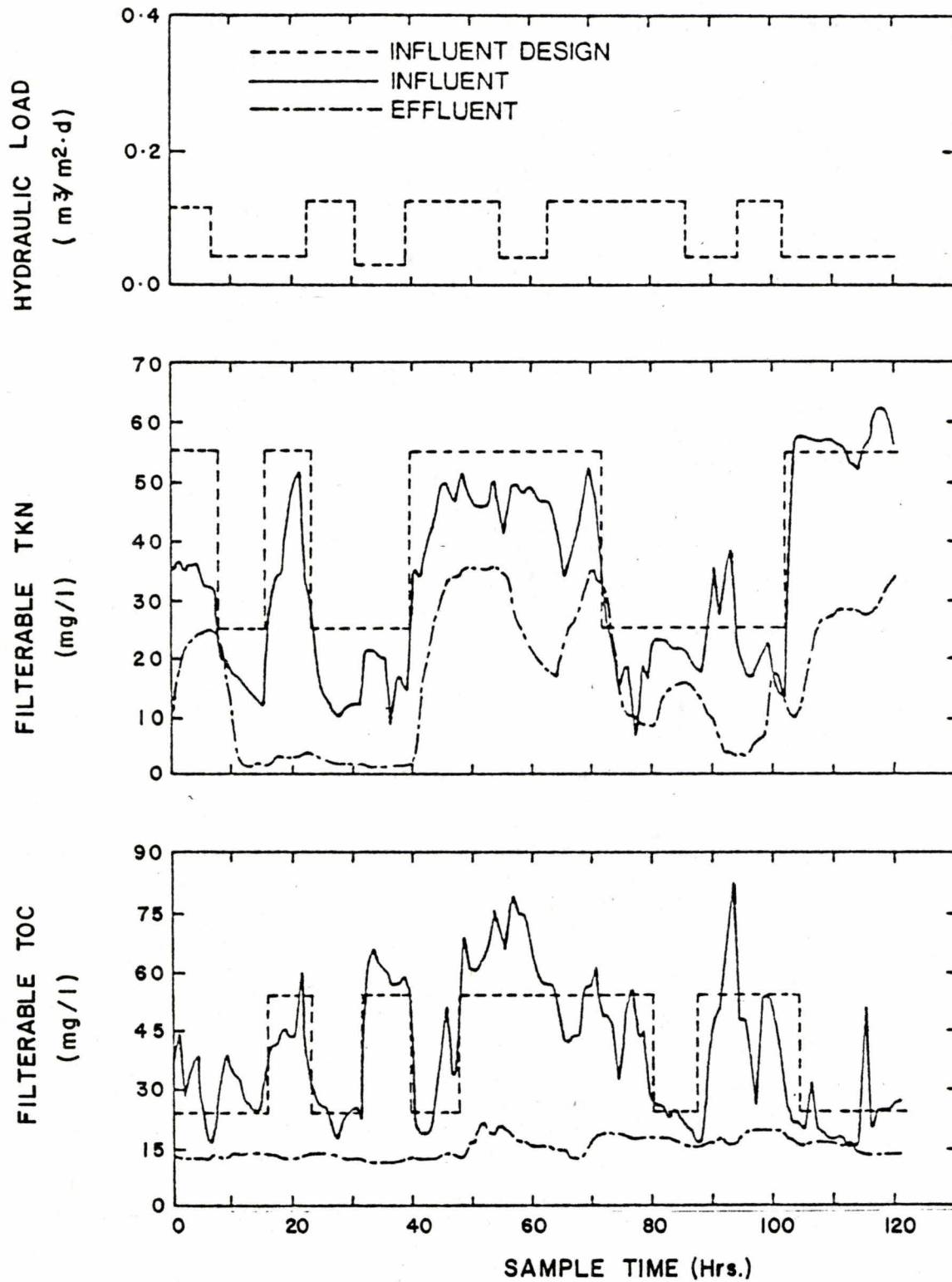


FIGURE 4.2
INPUT AND RESPONSE OF 0.5m RBC, CARBON OXIDATION MODE

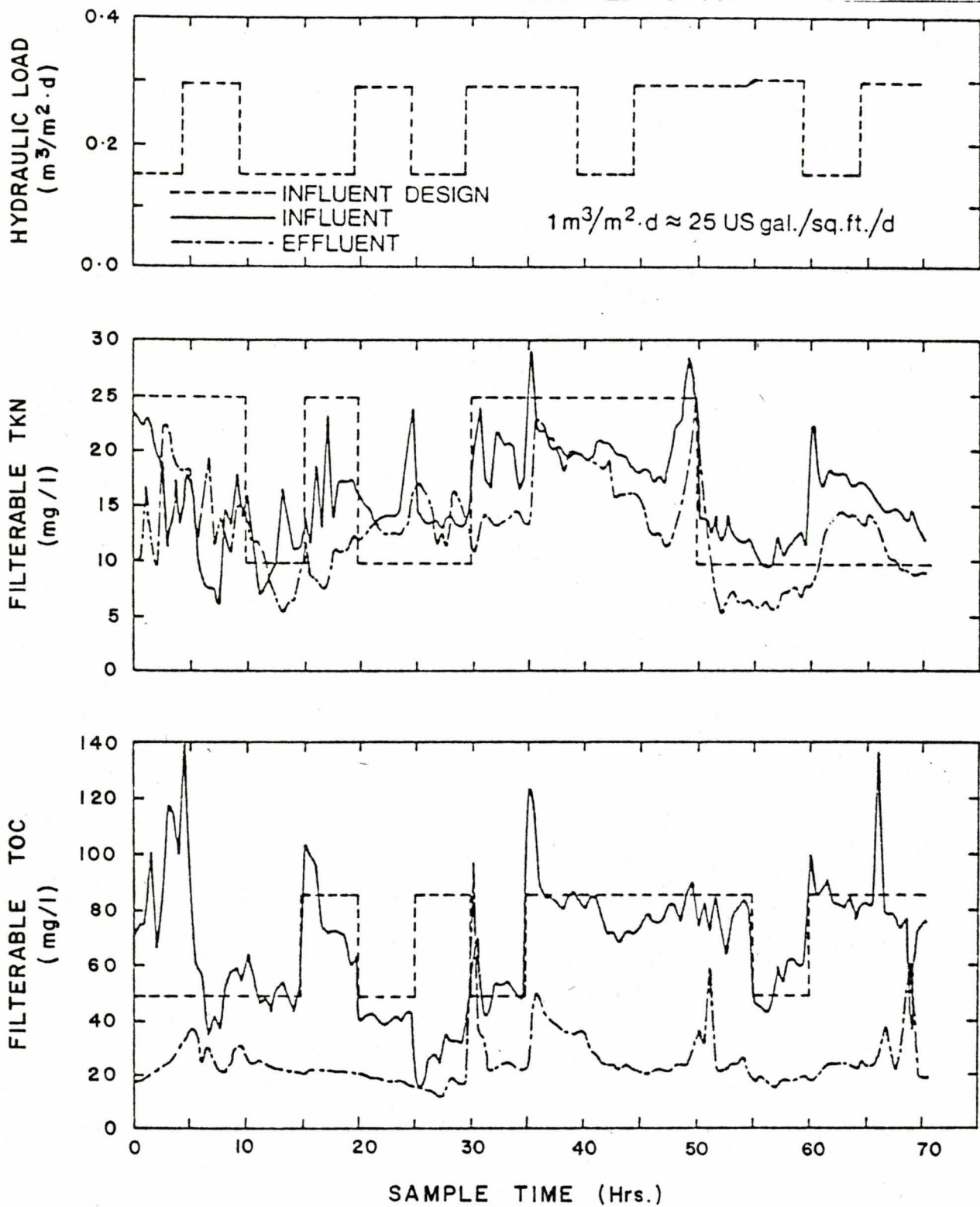
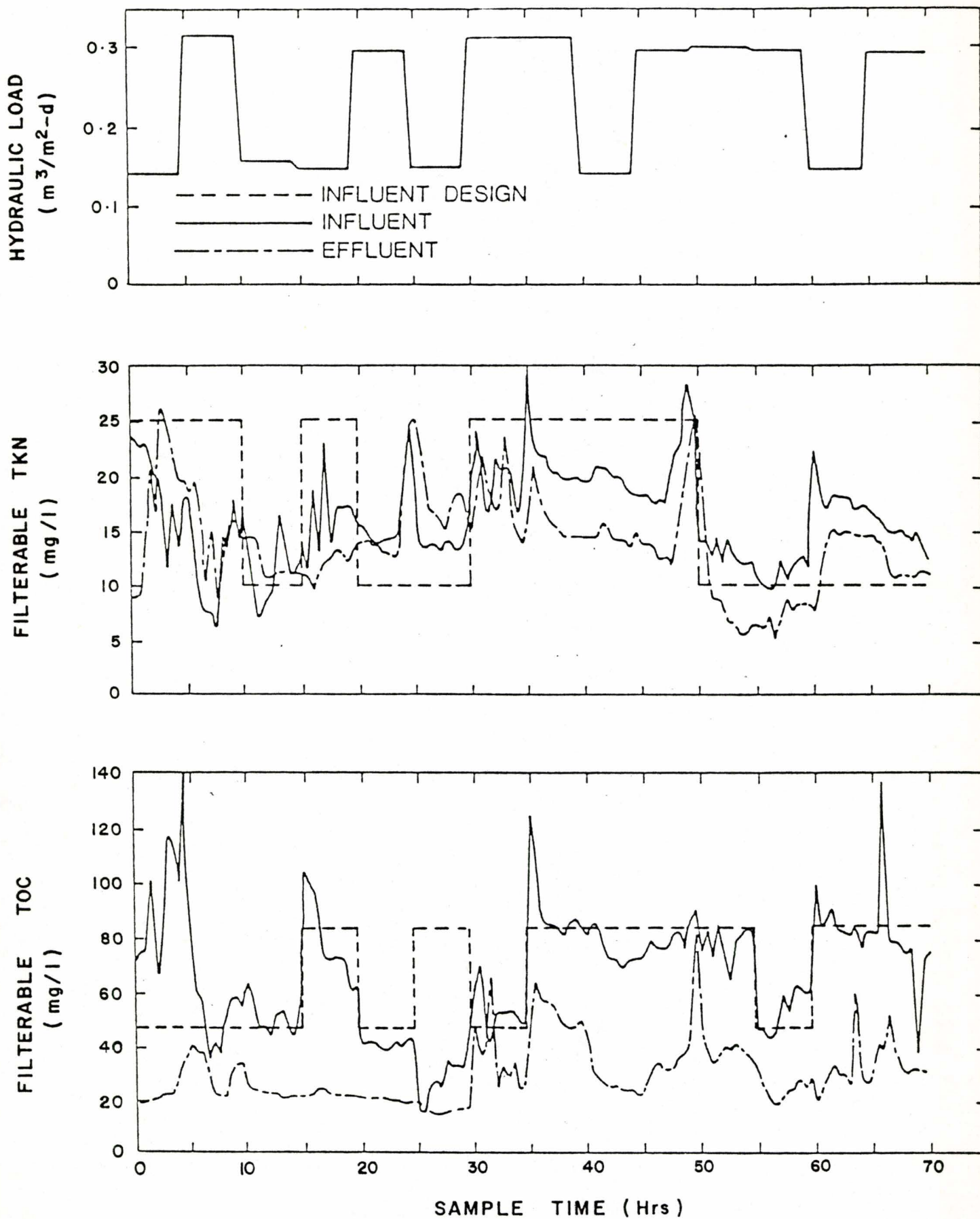


FIGURE 4-3

INPUT AND RESPONSE OF 2.0m RBC, CARBON OXIDATION MODE



4.2 System Response to Hydraulic, Organic and Inorganic Loading Variations

4.2.1 General

The impulse response function allows an assessment of the influent variables affecting effluent quality. It is also used to determine potential transfer function models which will describe the impulse response function and provide initial estimates of the potential transfer function model parameters (r , b , s , w , δ).

In order to verify suspected significant relationships between influent parameters and effluent response, the major relationships of interest were cross-correlated. The results of this initial screening are summarized in Table 4.1. Using Table 4.1, cause and effect relationships can be determined since a factorial design was used successfully in the experimental design.

The data presented in Table 4.1 provide several interesting relationships. During the carbon oxidation operational mode, influent carbon loading and concentration showed a significant correlation with effluent carbon concentration. However, significant effluent responses to dynamic flow conditions were not observed in the 0.5 metre RBC., while they were observed with the 2.0 metre RBC. There are two important ramifications from these observations:

1. The 0.5 metre RBC is used in many situations to obtain pilot scale performance data for the design of treatment plants. Since the performance of the two units have not been shown to be identical, care should be taken in extrapolating performance data.
2. As discussed in Chapter 2, the primary design criterion for RBCs has been based on areal hydraulic loading. However, a significant effluent

TABLE 4.1
SUMMARY OF CROSS CORRELATION RESULTS,
FIRST DIFFERENCE*

<u>Influent vs Effluent</u>		Observed Significant Correlations At Low Lags (95% Confidence)	
<u>Load</u>	<u>Conc.</u>	<u>Carbon Oxidation Mode</u>	<u>TKN Removal Mode</u>
COD Load	COD	Yes	Yes
TOC Load	COD	Yes	Yes
NH ₃ -N Load	COD	Yes	No
TKN Load	COD	Yes	No
Flow	COD	Yes/No	No
COD Load	TOC	Yes	Yes
TOC Load	TOC	Yes	Yes
NH ₃ -N Load	TOC	Yes	No
TKN Load	TOC	Yes	No
Flow	TOC	Yes/No	No
COD Load	NH ₃ -N	n/a	Yes
TOC Load	NH ₃ -N	n/a	Yes
NH ₃ -N Load	NH ₃ -N	n/a	Yes
TKN Load	NH ₃ -N	n/a	Yes
Flow	NH ₃ -N	n/a	Yes
COD Load	TKN	n/a	Yes
TOC Load	TKN	n/a	Yes
NH ₃ -N Load	TKN	n/a	Yes
TKN Load	TKN	n/a	Yes
Flow	TKN	n/a	Yes
COD Load	NO ₂ + NO ₃ -N	n/a	Yes
TOC Load	NO ₂ + NO ₃ -N	n/a	Yes
NH ₃ -N Load	NO ₂ + NO ₃ -N	n/a	No
TKN Load	NO ₂ + NO ₃ -N	n/a	No
Flow	NO ₂ + NO ₃ -N	n/a	Yes
<u>Conc.</u>	<u>Conc.</u>		
COD	COD	Yes	No
TOC	COD	Yes	No
NH ₃ -N	COD	No	No
TKN	COD	No	No
COD	TOC	Yes	No
TOC	TOC	Yes	No
NH ₃ -N	TOC	No	No
TKN	TOC	No	No
COD	NH ₃ -N	n/a	No
TOC	NH ₃ -N	n/a	No
NH ₃ -N	NH ₃ -N	n/a	Yes
TKN	NH ₃ -N	n/a	Yes
COD	TKN	n/a	No
TOC	TKN	n/a	No
NH ₃ -N	TKN	n/a	Yes
TKN	TKN	n/a	Yes
COD	NO ₂ + NO ₃ -N	n/a	No
TOC	NO ₂ + NO ₃ -N	n/a	No
NH ₃ -N	NO ₂ + NO ₃ -N	n/a	Yes
TKN	NO ₂ + NO ₃ -N	n/a	Yes

*This table applies for both 0.5 MI and 2.0M (2.0 M/0.5 MI) in the carbon oxidation mode, and only 0.5 MI in the nitrification mode.

response to hydraulic loading was not observed for carbon oxidation on the 0.5 metre RBC.

While the 0.5 metre RBC was operating under nitrifying conditions, significant effluent carbon responses were not observed to occur in relation to either hydraulic loading variations or carbon concentration (Table 4.1). However, the combined effect of these parameters (carbon loading) did show a significant relationship with effluent carbon concentration. Influent COD and TOC loadings were not excessive during the experiment and were within normal design limits. Such an observation would not be expected with an activated sludge plant operating under nitrifying conditions, and indeed such an effect was not observed by Sutton (1976). The observation indicates that the RBC may be overly sensitive to fluctuating influent loading conditions.

During the carbon oxidation plus nitrification mode of operation, influent ammonia loading, influent ammonia concentration and hydraulic loading showed significant correlations with effluent ammonia concentration (see Table 4.1). The effluent response to these influent conditions will be examined and quantified using time series modelling techniques.

An example of cause and effect is evident during the carbon oxidation plus nitrification operational mode. A significant correlation is observed between influent carbon loadings and effluent ammonia concentration (Table 4.1). However, significant correlations between influent carbon concentration and effluent ammonia concentration were not observed, while hydraulic loading versus effluent ammonia concentration was significant. Since hydraulic loading and substrate concentration were not confounded in the experimental design, it is reasonable to conclude that the significant effects observed must have been caused by hydraulic loading variations.

4.2.2 Impulse Response Functions - Carbon Oxidation Mode

The impulse response weights for influent filterable TOC loading and TOC concentration versus effluent filterable TOC concentration presented in Figure 4.4 indicate that a significant positive response was obtained in the effluent within one-half hour after the step changes occurred. Use of shorter sampling intervals might have defined the time to the maximum response. The response was directly proportional to the influent TOC loading and concentration. No significant effluent TOC response was observed for hydraulic loading to the 0.5 metre RBC.

Significant effluent filterable TOC responses are observed for variations in influent filterable TOC loading, TOC concentration, and flow rate to the 2.0 meter RBC (Figure 4.5). Figure 4.5 also indicates that the response was instantaneous, which is physically impossible. This result is observed because step changes were made 15 minutes before the hour, while discrete sampling was done at half-hour intervals, beginning on the hour. This lag time in sampling can be accounted for during model building, however, transfer function models were not developed for carbon removal as part of this work.

An important sidelight of Table 4.1 and more visually appreciated in Figures 4.4 and 4.5 is the fact that the variations in hydraulic loading did not significantly affect effluent TOC concentration for the 0.5 metre RBC, while it did for the 2.0 metre RBC. Hydraulic design levels were chosen from manufacturers specifications (which are based solely on areal hydraulic loading) to ensure that a response would be observed. These results reflect a difference in performance of the two RBC units and suggest that the design basis of flow per unit area is not comparable for scale-up purposes.

FIGURE 4.4

EFFLUENT TOC IMPULSE RESPONSE WEIGHTS, 0.5m RBC,
CARBON OXIDATION MODE, FIRST DIFFERENCE

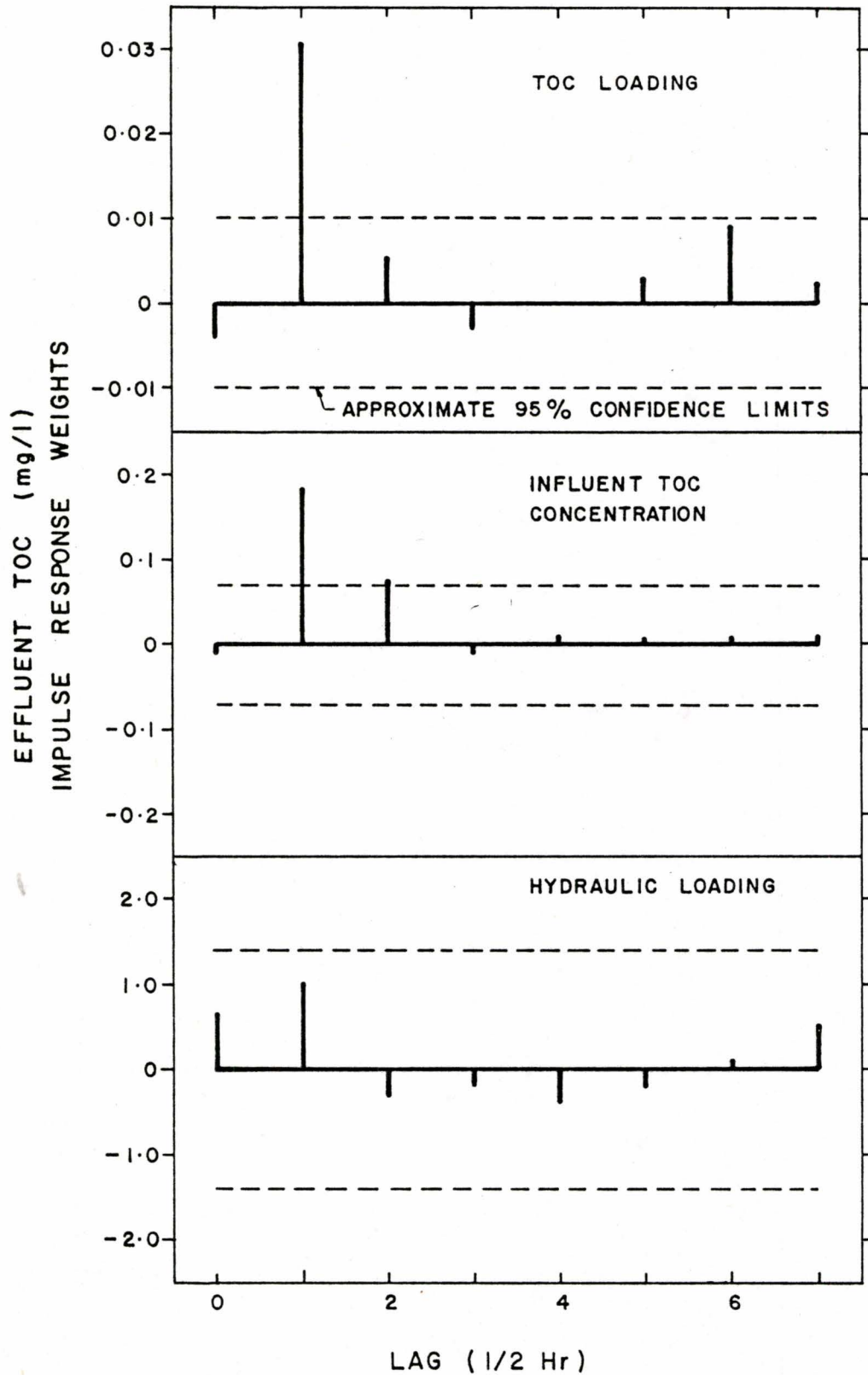
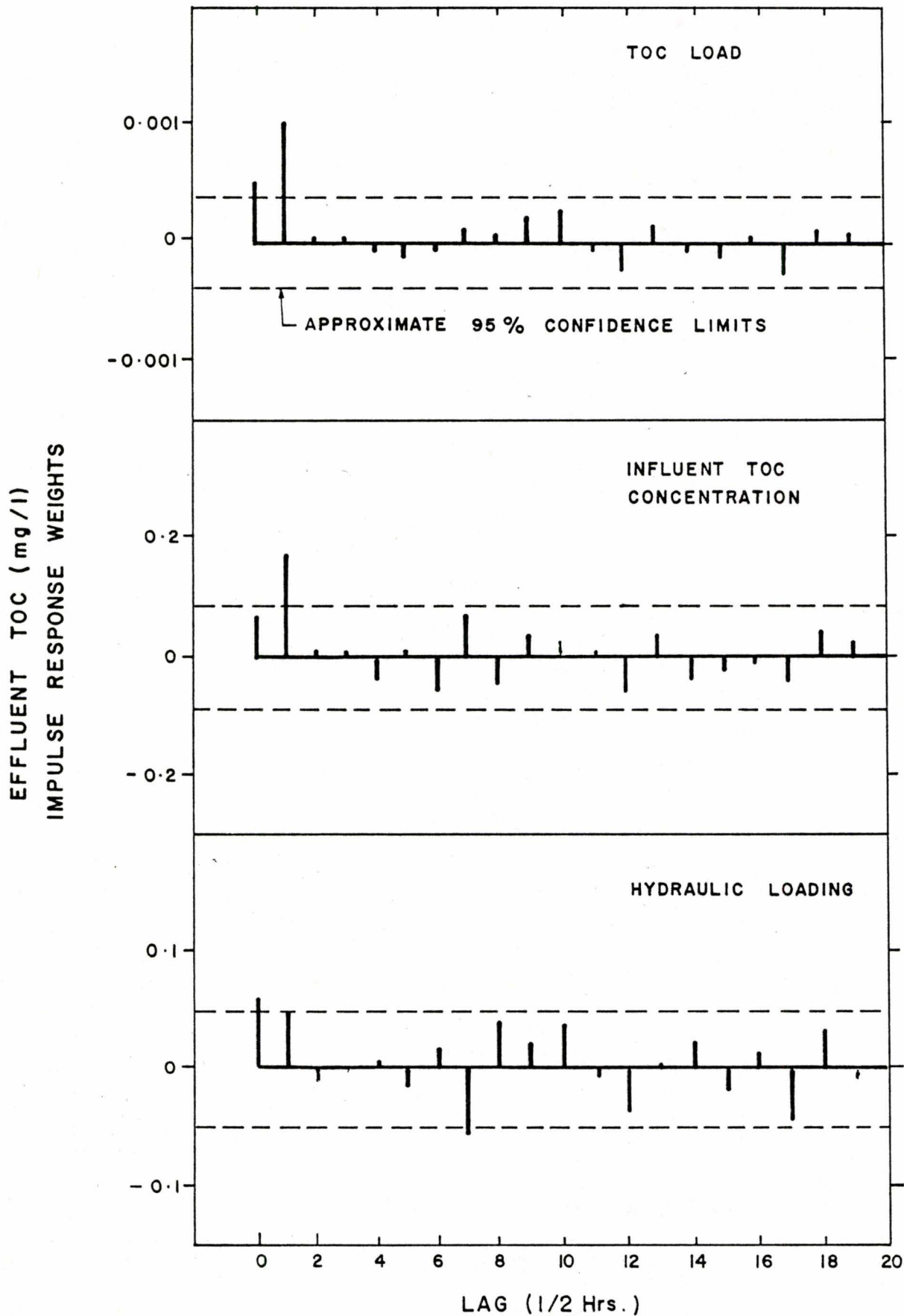


FIGURE 4.5
 EFFLUENT TOC IMPULSE RESPONSE WEIGHTS, 2.0m RBC,
 CARBON OXIDATION MODE, FIRST DIFFERENCE



4.2.3 Impulse Response Functions - Nitrification Mode

The significant positive correlations for filterable TKN loading, TKN concentration, and flow versus effluent filterable TKN concentration are presented in Figure 4.6 for the 0.5 metre RBC. The influent loading and concentration responses were observed to have a duration of about three hours. With flow, the significant TKN effluent response was observed only at a time delay of one hour. Shorter sampling intervals might have defined the time to maximum response.

The impulse response weights for filterable TKN loading, TKN concentration, and flow versus filterable effluent $\text{NO}_2\text{-N} + \text{NO}_3\text{-N}$ formation are given in Figure 4.7. A significant positive correlation is observed for influent TKN concentration, as would be expected. A significant negative correlation is observed for hydraulic loading. The combined relationship of TKN loading does not show any significant relationship due to the opposite effects of TKN concentration and hydraulic loading.

Investigations by Davies and Pretorius (1974) suggest that a decrease in $\text{NO}_2\text{-N} + \text{NO}_3\text{-N}$ formation coupled with an increase in flow can be explained because of the simultaneous increase in organic loading. It was explained that the increased organic load would enhance the growth of heterotrophs, which would displace nitrifier growth in the RBC system. As previously noted, the nitrifiers have been observed only in the final compartments of an RBC, while heterotrophs predominate in the initial RBC stages.

This explanation has merit in the long term. However, Table 4.1 indicates that there is no significant correlation at the 95 percent confidence level for TOC concentration versus $\text{NO}_2\text{-N} + \text{NO}_3\text{-N}$ formation. Figure 4.7 indicates a negative correlation for flow rate. Since flow

FIGURE 4.6

EFFLUENT TKN IMPULSE RESPONSE WEIGHTS, 0.5m RBC,
CARBON OXIDATION PLUS NITRIFICATION MODE, FIRST DIFFERENCE

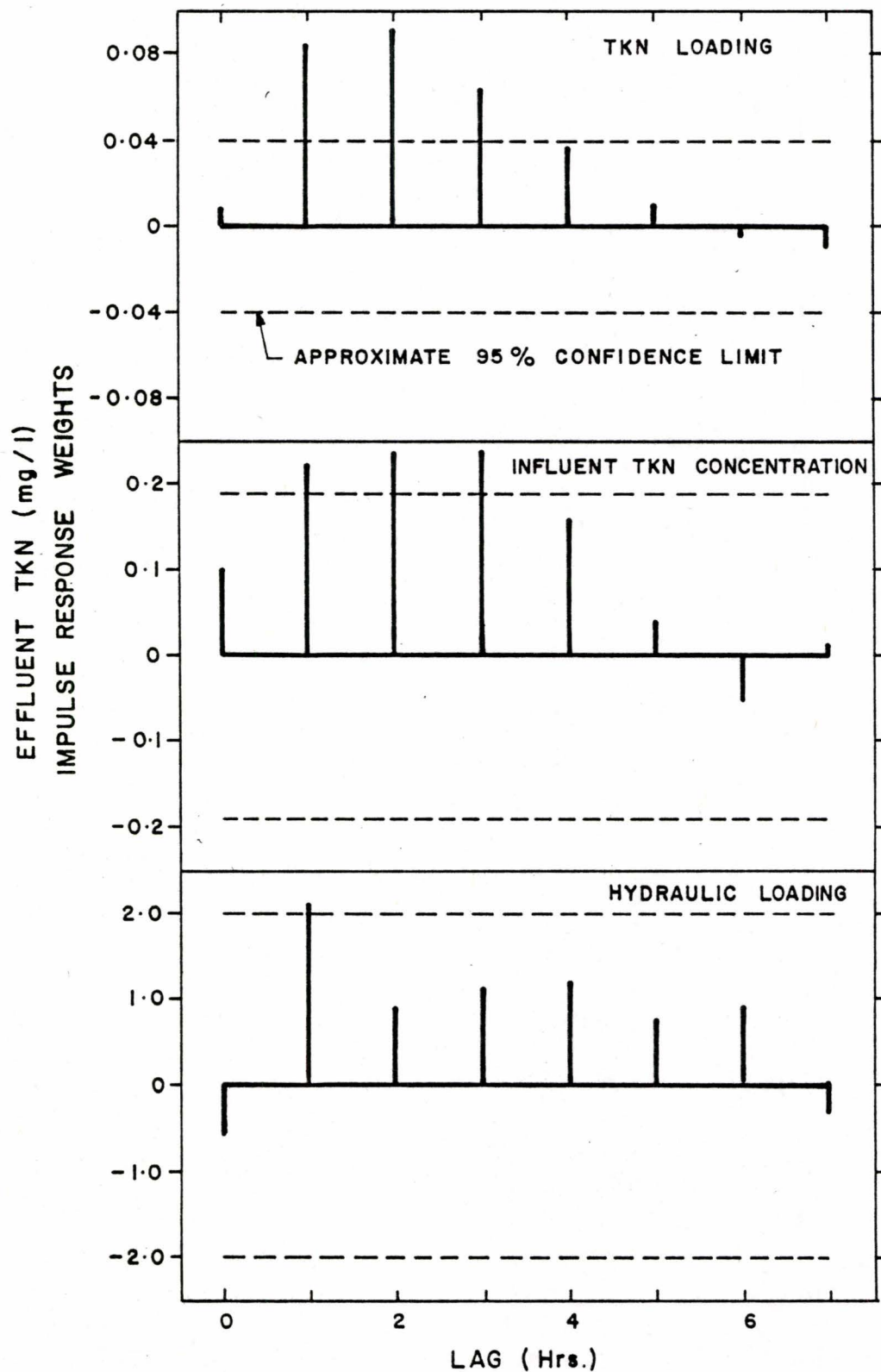
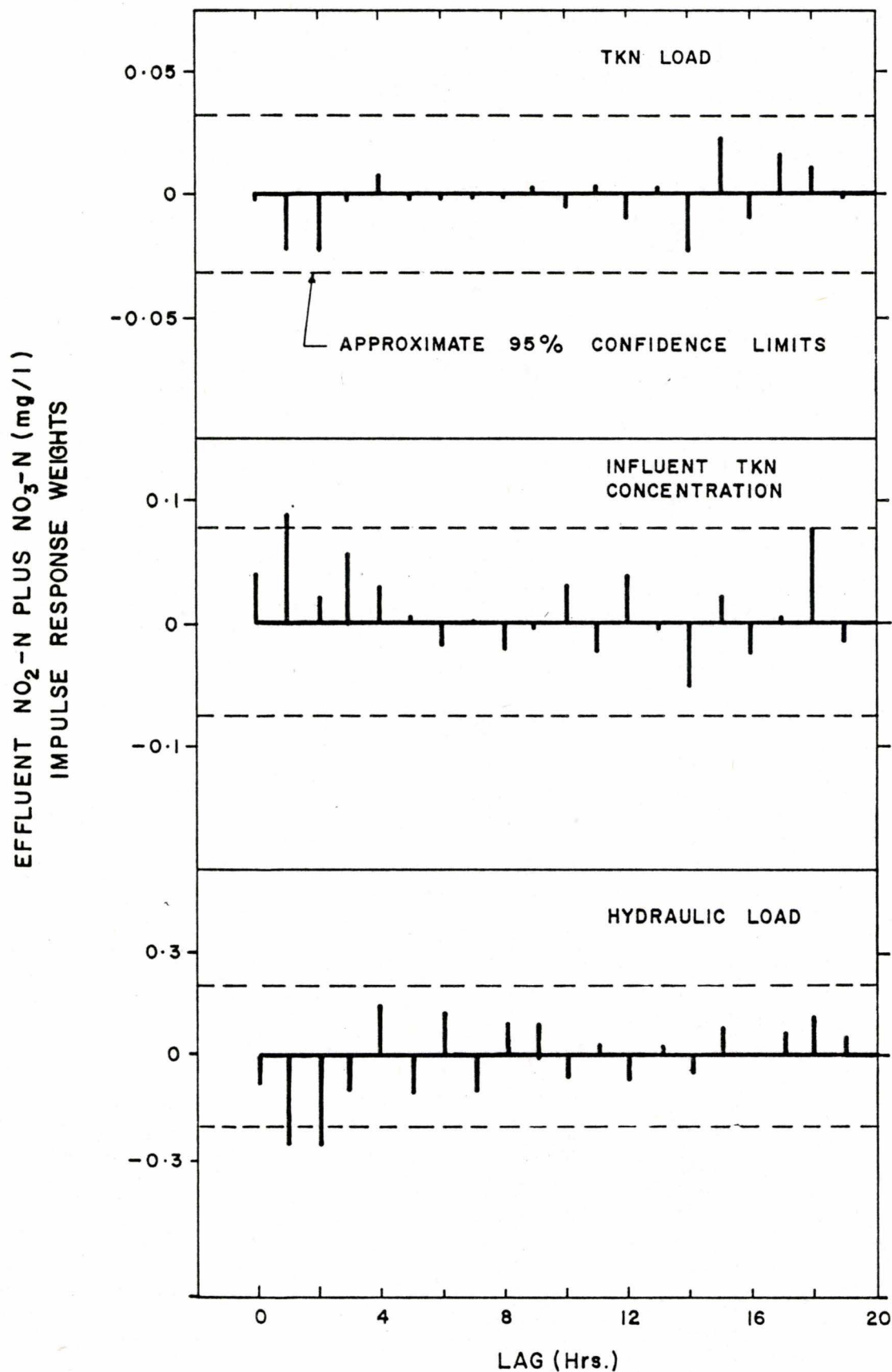


FIGURE 4.7

EFFLUENT $\text{NO}_2\text{-N}$ PLUS $\text{NO}_3\text{-N}$ IMPULSE RESPONSE FUNCTIONS
OF 0.5m RBC CARBON OXIDATION PLUS NITRIFICATION MODE,
FIRST DIFFERENCE



rate and TOC concentration were not confounded in the experimental design, it is reasonable to conclude that the decrease in $\text{NO}_2\text{-N} + \text{NO}_3\text{-N}$ formation may be attributed primarily to retention time or flow rate, in the short term.

4.2.4 Discussion

The impulse response functions (Figures 4.4, 4.5, 4.6 and 4.7) would indicate several potential disadvantages of the RBC system. Response to step inputs took place in less than one sampling period. Recovery to new steady-state values was about one hour for carbon oxidation and three hours of carbon oxidation plus nitrification. This indicates that the RBC would be sensitive to influent fluctuations and would provide little reserve capacity to minimize fluctuations in effluent quality. Although these slugs should not seriously affect the biomass, additional treatment capacity or equalization may be required to reduce these fluctuations and improve treatment.

4.3 Dynamic Transfer Function - Noise Model Development

4.3.1 Introduction

The cross correlation results presented in Section 4.2 indicate that there are a several parameters which affect effluent quality. Design parameters for RBCs have been based on areal hydraulic loading, while the effect of concentration and substrate loading have essentially been ignored. Only in the past two years has some emphasis been placed on concentration (Antonie 1976).

Quantification of RBC effluent response to various input parameters can be obtained through the use of the dynamic transfer function - noise (TF-N) models developed in this study. Model development should indicate which of the three influent parameters (flow, concentration, or loading)

best describes effluent quality, and therefore, is the best alternative on which to base design.

Transfer function - noise models were developed only for nitrifying conditions, using TKN data.

4.3.2 Model Development

The TF-N models listed in Table 4.2 describe effluent filterable TKN concentration. The models were developed according to the iterative procedure described from the impulse response weights and used as starting values in TSHAUS. The dynamic and noise model parameters were estimated simultaneously. Details of the calculation procedure are provided in Appendix C for a particular example. Diagnostic checks applied to the model residuals confirm both TF model and noise model adequacy at the 95% confidence level (Table 4.3).

4.3.3 Effluent Filterable TKN Models

Filterable influent TKN loading, TKN concentration and hydraulic loading models were found to adequately describe effluent TKN response (Table 4.2). Comparison of the dynamic transfer function-noise models was made through examination of the residual sums of squares. Extra sums of squares testing (Draper and Smith (1968)) was used to differentiate between models of different numbers of parameters. The TKN load model (Model A2) has the lowest residual mean square (Table 4.2) and, of the models considered, best describes effluent TKN concentration. Figure 4.8 demonstrates the good fit of the TF-N model with the experimental data. However, the TF model underestimates the peaks and valleys, while still adequately predicting response times. The sensitivity of the nitrifiers to varying influent loads, and the smooth response patterns as exhibited in Figure 4.8 are noted.

TABLE 4.2

TRANSFER FUNCTION-NOISE MODELS DESCRIBING
EFFLUENT FILTERABLE TKN CONCENTRATION

Input/Output Variables*	Model No.	Model	Residuals		
			Sum of Squares	Degrees of Freedom	Variance
TKN LOAD/TKN CONC.	A1	$Y_t = \frac{[(0.08312 \pm 0.01738) + (0.03450 \pm 0.02426)\beta] \beta X_T + a_t (1-\beta)^{-1}}{(1-0.5590 \pm 0.1365\beta) (1-0.2339 \pm 0.1808\beta)}$	306.6	116	2.64
TKN LOAD/TKN CONC.	A2	$Y_t = \frac{[(0.08110 \pm 0.01724) + (0.0460 \pm 0.02755)\beta + (0.02815 \pm 0.02813)\beta^2] \beta X_T}{(1-0.3994 \pm 0.2390\beta)}$ $+ \frac{a_t (1-\beta)^{-1}}{(1-0.2203 \pm 0.1823\beta)}$	294.4	115	2.56
Log ₁₀ TKN LOAD/TKN CONC.	A3	$Y_t = \frac{[(7.134 \pm 1.878) + (3.946 \pm 2.663)\beta + (2.894 \pm 2.667)\beta^2] \beta X_T}{(1-0.4476 \pm 0.2471\beta)}$ $+ \frac{a_t (1-\beta)^{-1}}{(1-0.3401 \pm 0.1766\beta)}$	374.0	115	3.25

Loading models of the form (0, 2, 1) and (1, 0, 1) were also developed, but inadequacies were found with the transfer function during cross-correlation residual checks.

* Loading units are in g/d, concentration units are in mg/l, flowrate is in l/d.

TABLE 4.2 (Cont'd)

Input/Output Variables*	Model No.	Model	Residuals		
			Sum of Squares	Degrees of Freedom	Variance
TKN CONC./TKN CONC.	B1	$Y_T = \frac{[(0.1066 \pm 0.0573) + (0.07413 \pm 0.07367)\beta + (0.0670 \pm 0.07640)\beta^2]}{(1 - 0.4148 \pm 0.4597\beta)} \beta X_T$ $+ \frac{a_t (1-\beta)^{-1}}{(1 - 0.5409 \pm 0.1570\beta)}$	498.5	115	4.33
TKN CONC./TKN CONC.	B2	$Y_T = \frac{[(0.1096 \pm 0.0578) + (0.04406 \pm 0.06974)\beta] \beta X_T}{(1 - 0.7078 \pm 0.2549\beta)} + \frac{a_t (1-\beta)^{-1}}{(1 - 0.5334 \pm 0.1553\beta)}$	509.7	116	4.39
TKN CONC./TKN CONC.	B3	$Y_T = \frac{[(0.1029 \pm 0.0578) + (0.1064 \pm 0.0624)\beta + (0.08904 \pm 0.05776)\beta^2]}{(1 - 0.4148 \pm 0.4597\beta)} \beta X_T$ $+ \frac{a_t (1-\beta)^{-1}}{(1 - 0.5407 \pm 0.1564\beta)}$	512.8	116	4.42

Some of the parameters in the above models pass through zero indicating models of the forms (1, 0, 1) and (0, 1, 1). However, models of these forms failed to pass the cross correlation checks between input and residuals and are therefore not reported.

* Loading units are in g/d, concentration units are in mg/l, flow rate in l/d.

TABLE 4.2 (Cont'd)

Input/Output Variables*	Model No.	Model	Residuals		
			Sum of Squares	Degrees of Freedom	Variance
Flow Rate x 10 ⁻³ /TKN CONC. C1		$Y_T = \frac{(1.514 \pm 0.745)\beta X_T}{(1 - 0.6723 \pm 0.3327\beta)} + \frac{a_t(1-\beta)^{-1}}{(1 - 0.5693 \pm 0.1531\beta)}$	530.5	117	4.53
Flow Rate x 10 ⁻³ /TKN CONC. C2		$Y_T = [(0.1282 \pm 0.0720) + (0.6506 \pm 0.7204)\beta] \beta X_T + \frac{a_t(1-\beta)^{-1}}{(1 - 0.5891 \pm 0.1495\beta)}$	540.2	117	4.62
Flow Rate + TKN Conc./ TKN Conc	D	$Y_T = \frac{[(0.09538 \pm 0.00225)\beta] W_T}{(1 - 0.6306 \pm 0.0748\beta)} - [0.07762 \pm 0.04708]\beta^3 X_T + \frac{a_t}{(1-\beta)}$	321.8	117	2.75

The parameter in model C2 which passes through zero was eliminated, but the new model which was developed, (0, 0, 1), did not pass the model residual checks and is not reported.

* Loading units are in g/d, concentration units are in mg/l, flow rate in l/d.

TABLE 4.3

DIAGNOSTIC CHECKS APPLIED TO THE MODEL RESIDUALS

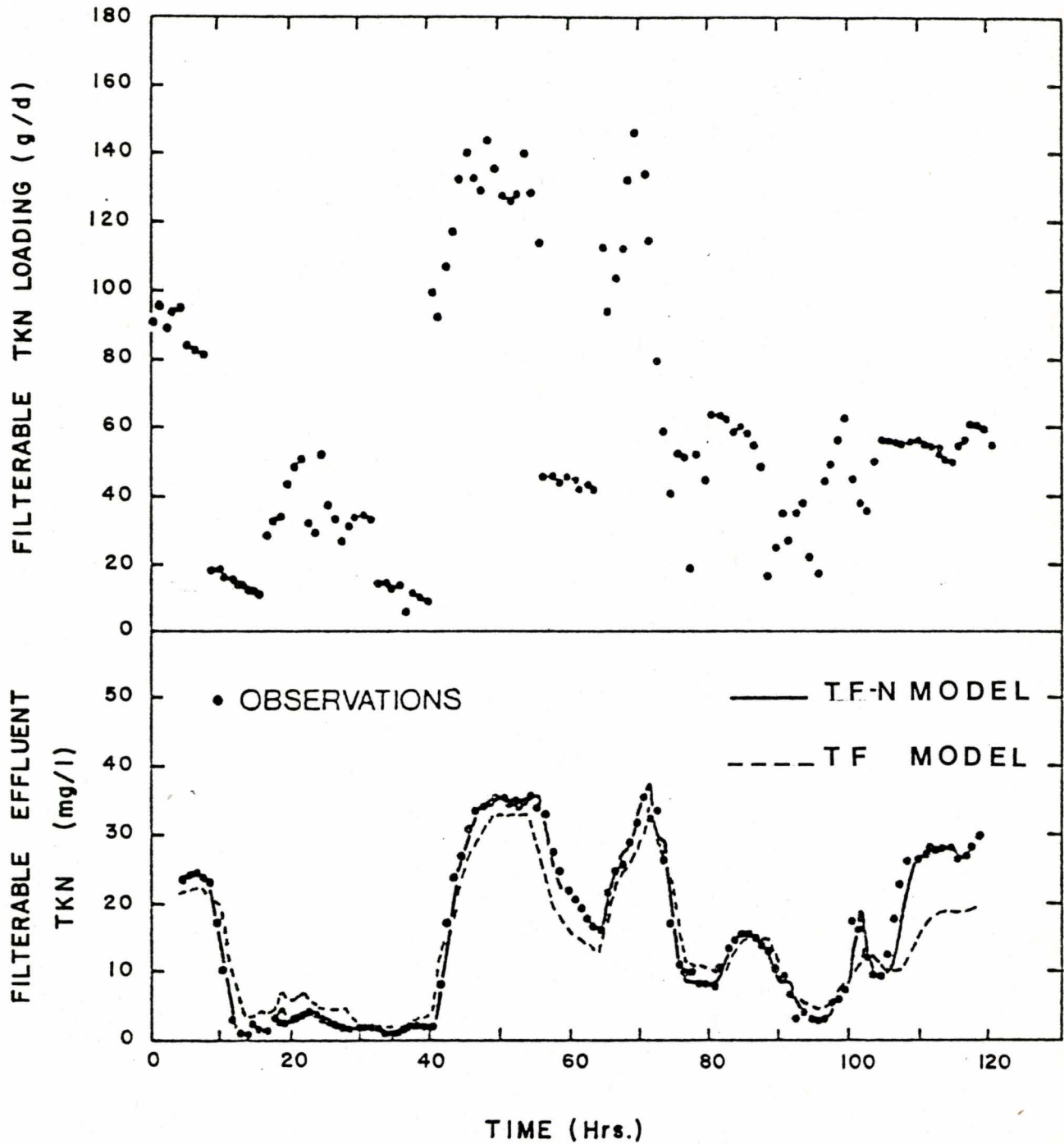
Input Variable	Model No.	Residual Autocorrelation Results		Cross Correlation Results		
		Q	$X^2, \gamma = 0.95$	$Sx^1\hat{a}$	$Sw^1\hat{a}$	$X^2, \gamma = 0.95$
TKN Load	A1	12.8	31.41	25.5		32.67
	A2	14.5	31.41	20.2		32.67
Log TKN Load	A3	14.72	31.41	17.36		32.67
TKN CONC.	B1	22.7	31.41	15.0		32.67
	B2	22.9	31.41	16.9		32.67
	B3	21.3	31.41	16.1		32.67
Flow Rate	C1	12.6	31.41	15.46		32.67
	C2	14.1	31.41	23.1		32.67
Flow Rate + TKN CONC.	D	16.8	31.41	21.7	23.9	32.67

where $Q = n \sum_{k=1}^{20} r_{aa}^2(k)$ $S = n \sum_{k=0}^{20} r_{x^1a}^2(k)$

- $r_a(k)$ = estimate of cross correlation function at lag K, or the autocorrelation function at lag K.
- a = model residuals
- x^1 = $(1-B)X$
- X = influent condition model was developed for (load or concentration)
- w = influent flow condition
- K = lag
- n = number of observations

FIGURE 4-8

COMPARISON OF EFFLUENT FILTERABLE TKN RESPONSE AND MODEL(A2) FIT



The results demonstrate that effluent filterable TKN concentration is best modelled using influent filterable TKN loading with this technique. Since the TKN load model had the lowest residual mean square, strong consideration should be given to basing RBC design criteria on TKN mass loading rather than hydraulic loading.

4.3.4 TF-N Model Analysis to Determine Response Times

The TF-N models developed in this study can be used to determine the approximate time at which a response in effluent conditions occurs to step changes in influent conditions. The analysis involves the projection of the impulse response function back through the time axis, or by calculation methods using the parameter values of the TF models (see Appendix C). Such an analysis revealed that there is an approximate time delay of 10 minutes before an effluent response occurs to a step change in influent loading. This is fast for an effluent treatment system. Sutton (1976) obtained an approximate 60 minute time delay for his separate activated sludge pilot plant when it was subjected to step inputs in TKN load.

A response to concentration variations in the input was calculated to be in the order of 20 minutes, while the response to hydraulic loading was almost instantaneous. A more accurate indication of the effluent response patterns and the system time delay may have been obtained using a shorter time interval between samples.

4.4 System Response to Natural Diurnal Variations

One of the objectives of this work was to determine the effect of diurnal inputs on instantaneous treatment efficiencies. In conjunction with this objective, it was desired to use an application of time series modelling, namely forecasting, to ensure that the model would describe

the effluent variability of the RBC for a completely independent set of data.

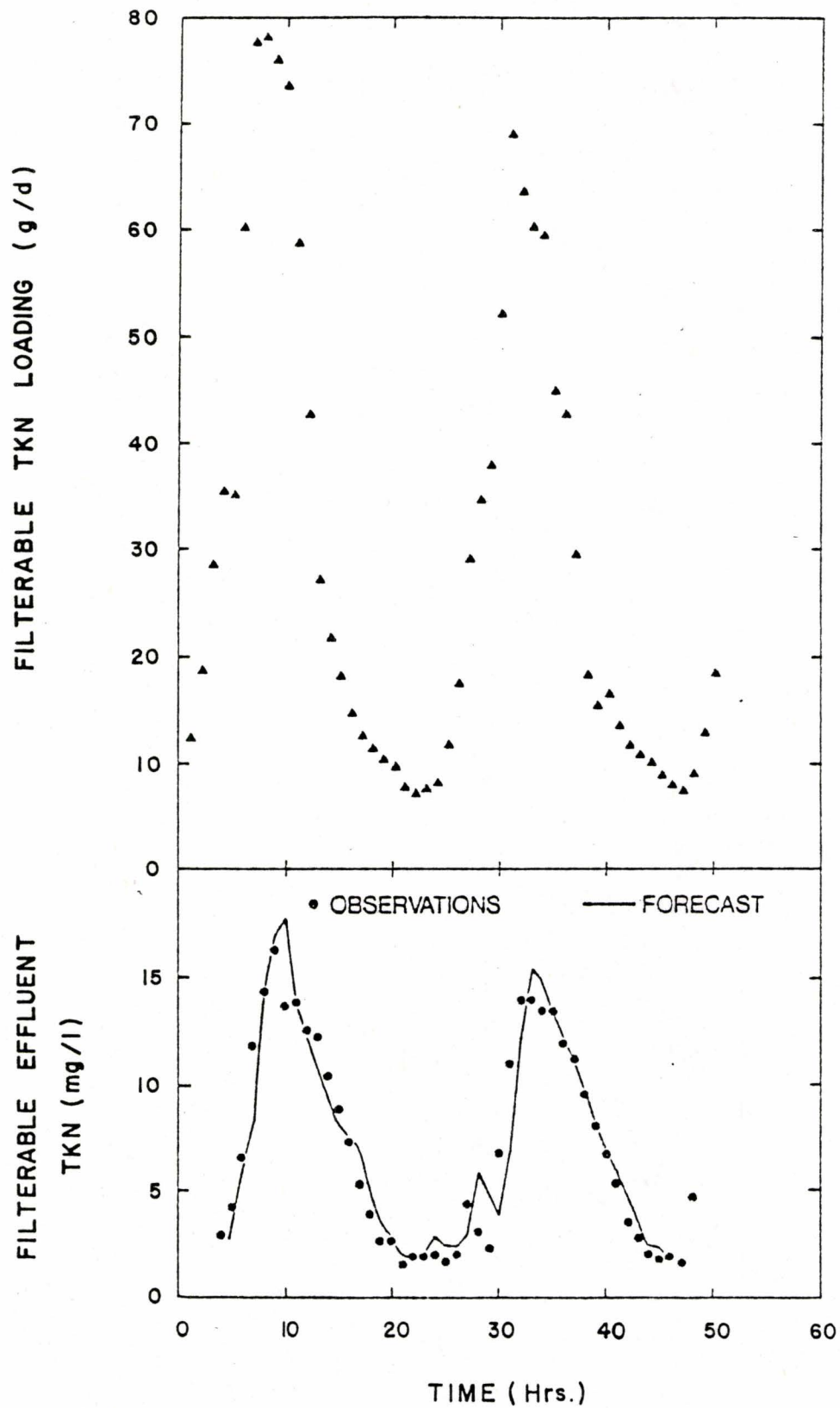
The natural diurnal variation in influent flow and concentration normally encountered at a small wastewater treatment plant was approximated on the same 0.5 meter RBC used to collect the dynamic data. The unit was subjected to a sinusoidal variation in flow synchronous to the natural concentration fluctuations of Burlington STP sewage, with a peak/average/minimum flow ratio of 2.5/1/0.5. Influent and effluent concentration and flow were monitored at hourly intervals for a period of 2 days.

4.4.1 TF-N Model Forecast Results

The forecast filtered effluent TKN concentration successfully predicted the observed daily effluent variation. (Figure 4.9) Response and recovery times agreed with the experimental data and substantiated the transfer function-noise model developed in this study. Fluctuations in effluent concentrations were extremely smooth, which seems to be a characteristic of nitrifying systems. Peak effluent concentrations lagged the influent peak by two hours. Details concerning the forecast procedures are provided in Appendix C.

The large fluctuations in effluent TKN concentration demonstrate the poor treatment performance of the RBC when operated in the nitrification mode under variable loading conditions (Figure 4.9). It is apparent that if stable effluent quality is to be produced using the RBC, process modifications or flow equalization must be provided to damp out any influent fluctuations.

FIGURE 4-9
EFFLUENT FILTERABLE TKN RESPONSE AND FORECAST
FOR DIURNAL LOADING



4.5 Comparison of RBC TF-N Models with Activated Sludge

A comparison of the sensitivity of the RBC system to conventional combined and separate nitrifying activated sludge systems may be made by calculating the gains of the TF-N models describing these systems. Dynamic TF-N models describing the effluent TKN response of activated sludge pilot plants were developed by Sutton (1976). The models given in Table 4.4 describe the dynamic relationship of influent TKN load to effluent TKN concentration. The influent flow and concentration data of the pilot plants were of comparable magnitude, and the systems were linear over the flow and concentration regimes investigated.

A comparison of the gains of the activated sludge systems with the RBC indicates that for a unit step change in the influent, 5 to 6 times greater response in effluent TKN concentrations may be expected with the RBC. This confirms the observations of Reimer et al (1975), who operated a pilot plant RBC system in parallel with a pilot plant activated sludge unit. *welner*

Since the response of the RBC to input fluctuations is so much larger than an activated sludge system, greater effluent variability may be expected from an RBC treatment plant, especially when servicing small communities. To stabilize effluent concentrations and to provide consistently good effluent quality, flow equalization should be incorporated into the design of RBC treatment plants.

4.6 Steady State Design

Although steady state design methods cannot describe the temporal relationship in the effluent when the influent to the RBC is variable, the relationship between influent load and mass removal is desirable. Steady state design is feasible as long as the effluent variability is

Table 4.4

Transfer Function-Noise Substrate Loading Models for RBC
and Activated Sludge Processes

System	Model	Calculated Gain	Ratio RBC to A/S
RBC	$Y_T = \frac{[(0.08110 \pm 0.01724) + (0.0460 \pm 0.02755)\beta + (0.02815 \pm 0.02813)\beta^2] \beta X_T}{(1 - 0.3994 \pm 0.2390\beta)}$ $+ \frac{a_t (1-\beta)^{-1}}{(1 - 0.2203 \pm 0.1823\beta)}$	0.259	
Combined A/S system	$Y_T = \frac{0.011X_{t-1}}{(1 - 0.786\beta)} + \frac{(1-\beta)^{-1}}{(1 - 1.066\beta + 0.336\beta^2)} a_t$	0.051	5.0/1
Separate A/S System	$Y_T = \frac{(0.012 + 0.008\beta) \beta X_t}{1 - 0.550\beta} + \frac{(1-\beta)^{-1}}{(1 - 1.394\beta + 0.540\beta^2)} a_t$	0.044	5.9/1

Where X_t represents time varying influent TKN load, g/d
 Y_t represents time varying effluent TKN concentration, mg/l

kept in mind, and steps are taken to reduce influent variability.

The development of dynamic transfer function noise models has indicated that influent mass loading should be considered as the primary design criterion in nitrifying systems. Although this relationship has not been proven for carbon removal, there is a strong indication that this may be the case from Figures 4.4 and 4.5.

A number of mass removal versus influent mass loading plots have been developed using the time series data obtained in this study. Mass removal calculations were based on a lag separation of 1 between influent and effluent for both the carbon oxidation mode and the carbon oxidation plus nitrification mode. This interval corresponds to the average retention times in the system, as determined by the tracer studies (Appendix D). Data points occurring during a step change and up to two hours after the step were not included in the results in order to allow the unit to recover to a "pseudo" steady state condition. All plots were standardized to a unit area basis.

4.6.1 Carbon Oxidation Mode

Figures 4.10 and 4.11 present the TOC and COD mass removal plots for the 0.5 meter RBC. The equivalent TOC and COD mass removal plots for the 2.0 meter RBC, which was run in parallel with the 0.5 meter RBC, are given in Figures 4.12 and 4.13.

Mass carbon removal generally appears to be a linear function with mass loading. However, when the TOC plots (Figures 4.10 and 4.12) are compared with the COD plots (Figures 4.11 and 4.13), respectively, a limiting TOC influent loading condition appears to have been approached on the 2.0 meter and 0.5 meter RBCs.

FIGURE 4-10

STEADY STATE DESIGN DATA FOR FILTERABLE TOC REMOVAL,
0.5m RBC, CARBON OXIDATION MODE

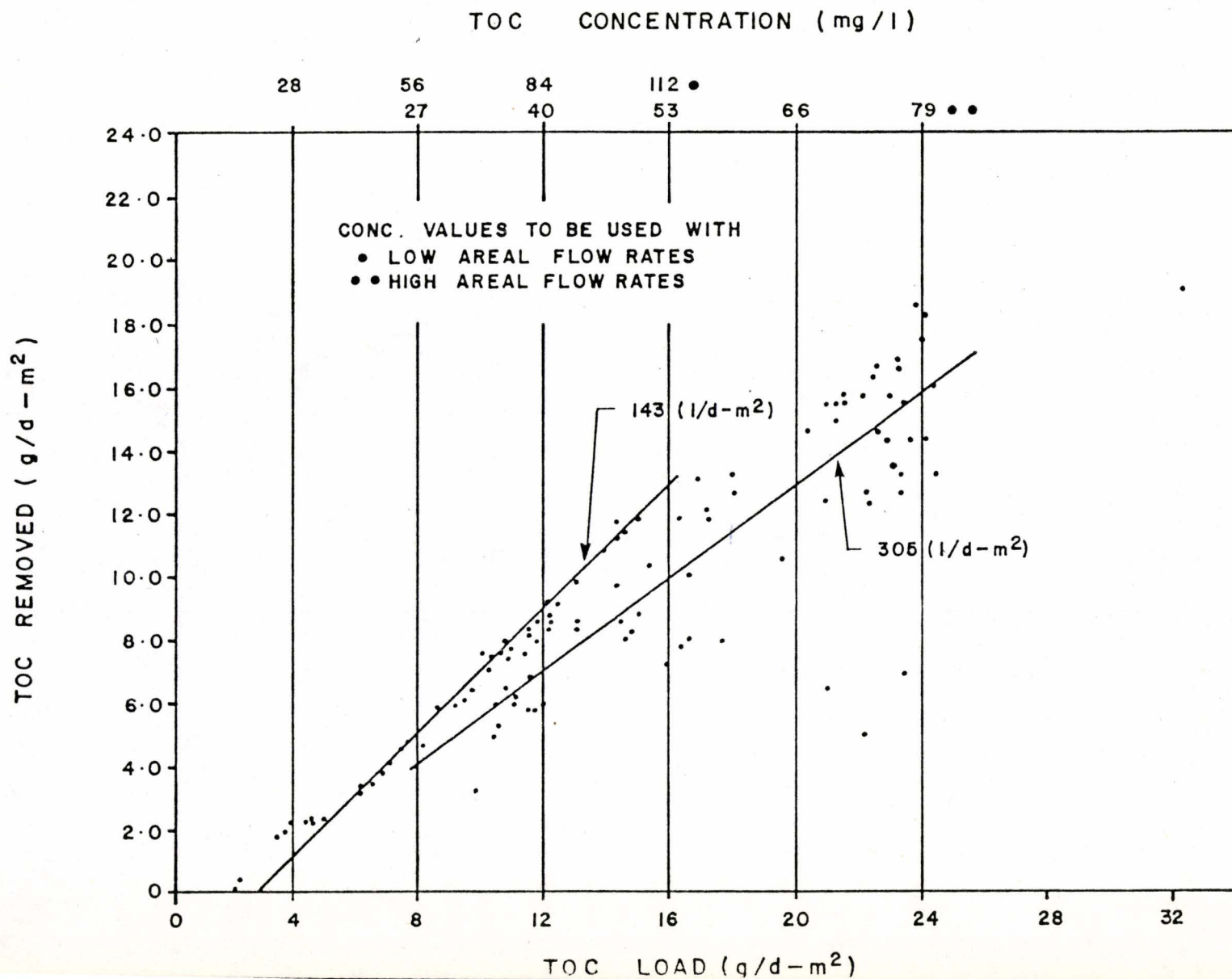


FIGURE 4-11

STEADY STATE DESIGN DATA FOR FILTERABLE COD REMOVAL,
0.5m RBC, CARBON OXIDATION MODE

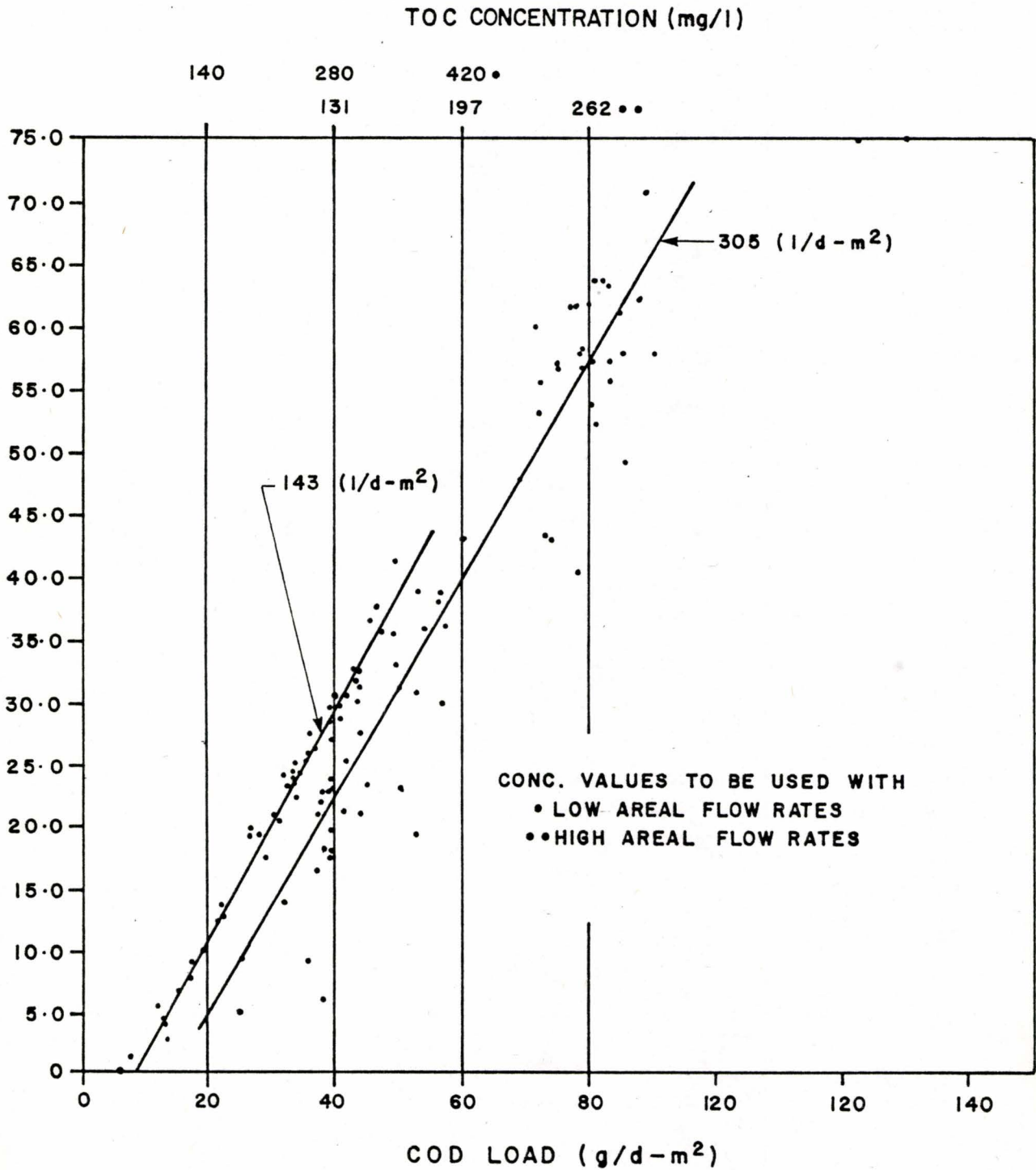


FIGURE 4-12

STEADY STATE DESIGN DATA FOR FILTERABLE TOC REMOVAL,
2.0m RBC, CARBON OXIDATION MODE

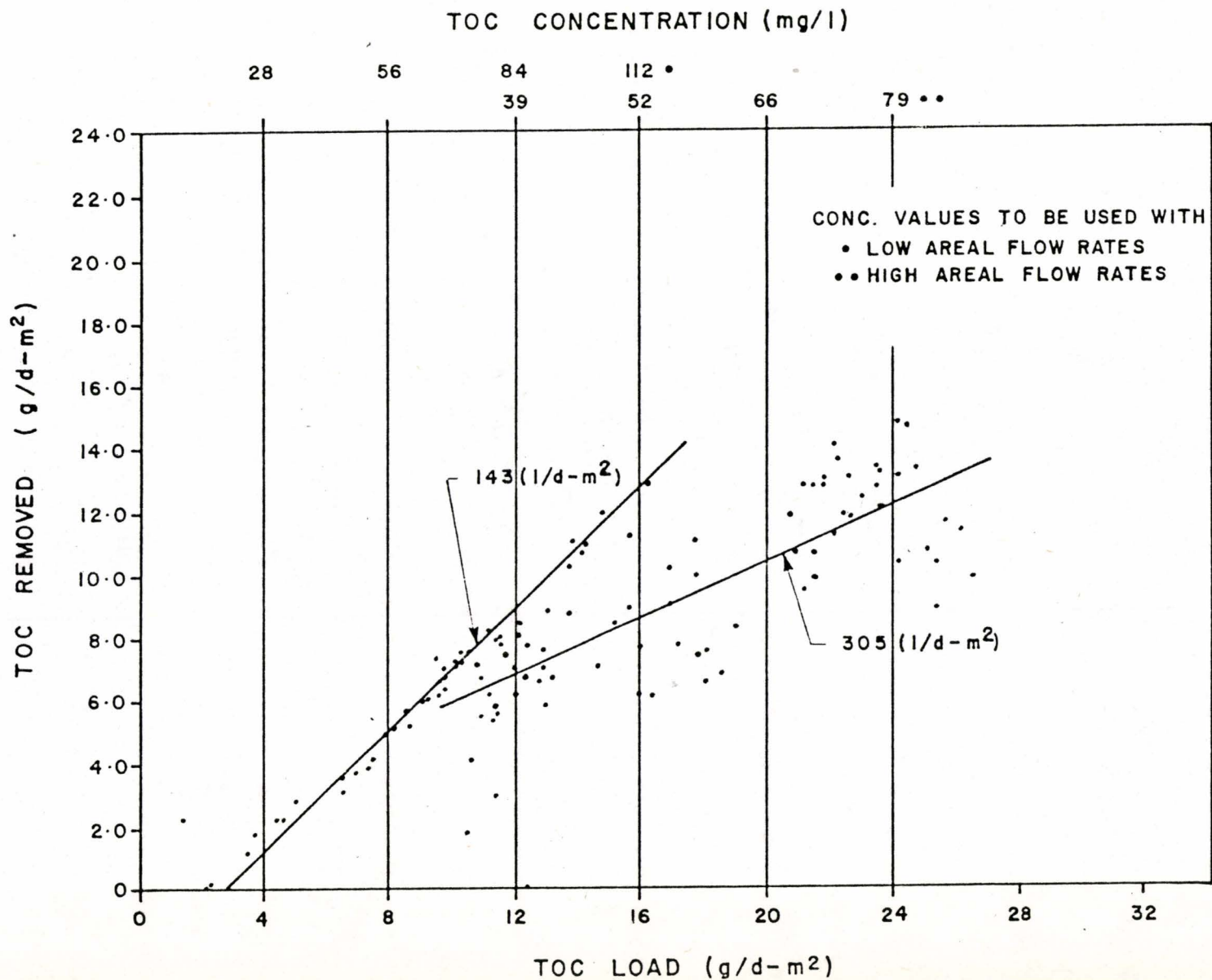
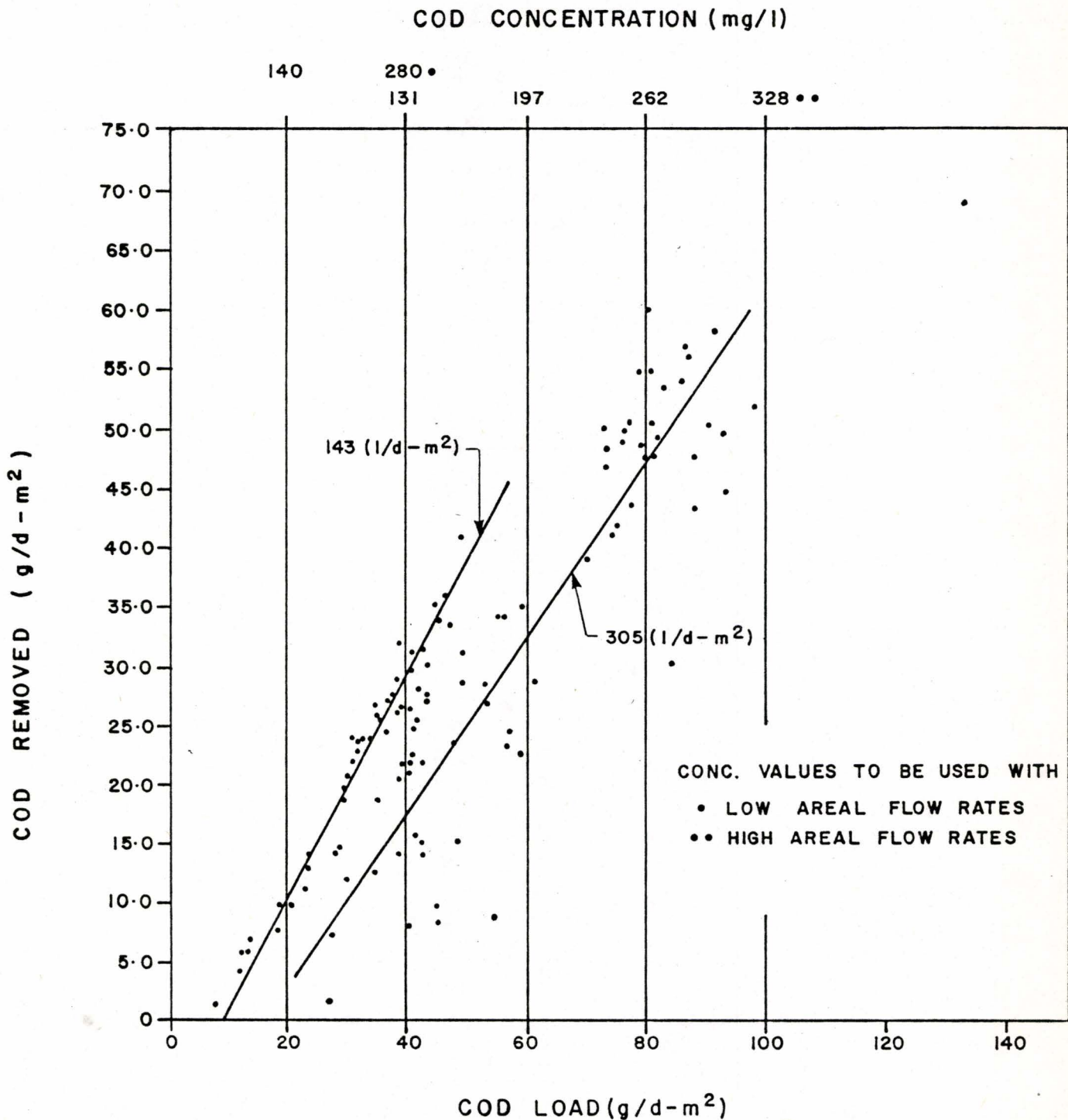


FIGURE 4-13

STEADY STATE DESIGN DATA FOR FILTERABLE COD REMOVAL,
2.0m RBC, CARBON OXIDATION MODE



It is interesting to note that two linear relationships are apparent at low loadings on Figures 4.10 to 4.13. These relationships coincide with the two levels of hydraulic loading used in the experimental designs (143 l/d-m² and 305 l/d-m²). The low hydraulic loading in each case provided for slightly improved treatment efficiency in terms of mass removal and stability. The variability in mass removal at the high loadings indicates that the RBCs were approaching unstable treatment conditions.

The presence of non-biodegradable residual organics is demonstrated by the fact that the plots do not pass through the origin.

4.6.2 Carbon Oxidation Plus Nitrification Mode

Mass loading plots for TOC and COD removal with the 0.5 metre RBC operating in the nitrification mode are given in Figures 4.14 and 4.15. Linear mass removal is apparent on both figures. Two levels of mass removal are apparent at equal mass loadings, with greater mass removal being obtained at the lower hydraulic loading.

The filterable TKN mass removal versus influent TKN load plot is presented in Figure 4.16. In contrast to carbon oxidation, a limiting mass removal was obtained at loadings greater than 2.0 g/d-m². Beyond this influent loading, mass removal becomes unstable with a maximum of 2 g/d-m² TKN being removed.

As in the carbon oxidation plots, a separate linear relationship exists at low loadings. This line projects through the origin, as would be expected.

FIGURE 4-14

STEADY STATE DESIGN DATA FOR FILTERABLE TOC REMOVAL,
0.5 m RBC, CARBON OXIDATION PLUS NITRIFICATION MODE

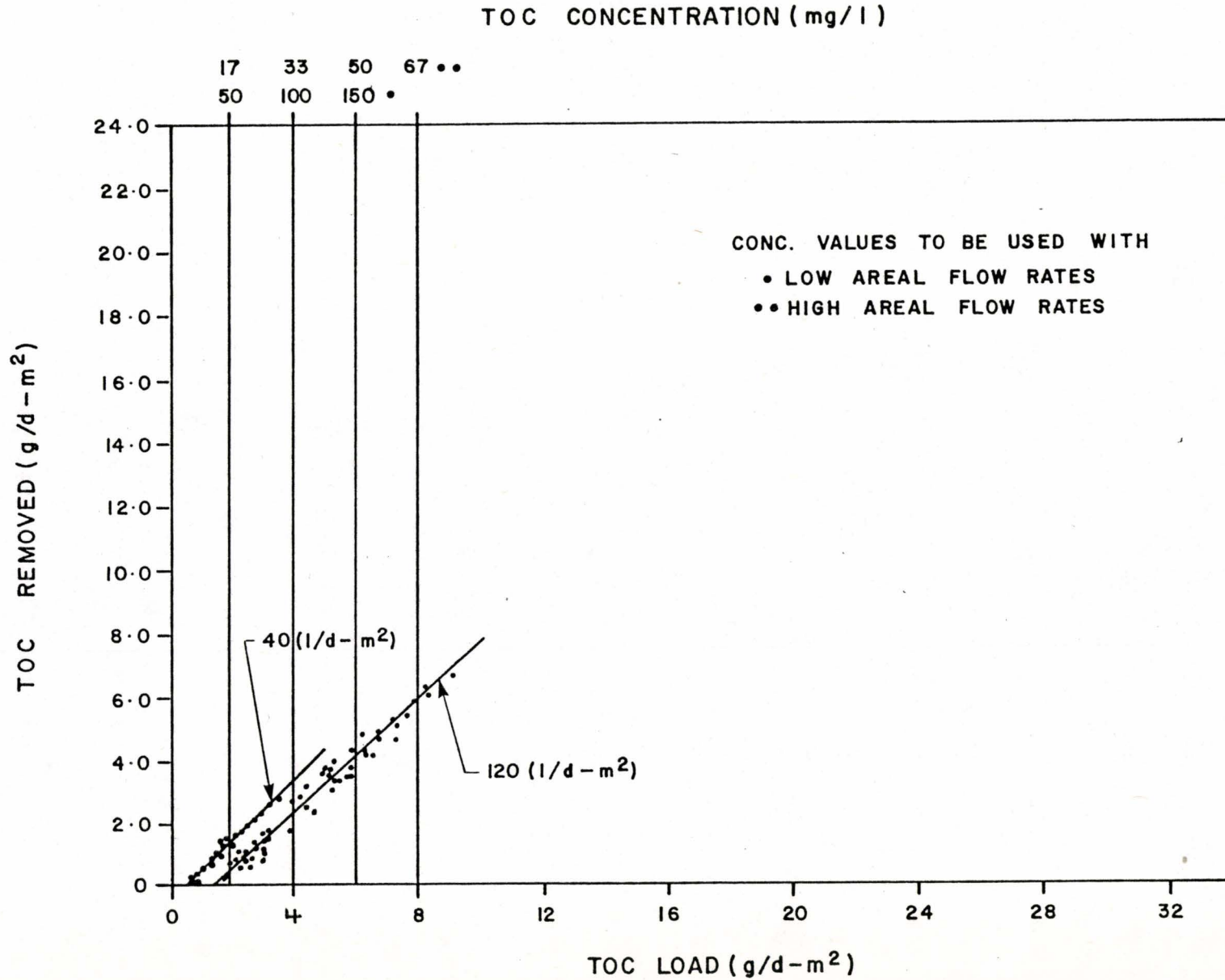


FIGURE 4-15

STEADY STATE DESIGN DATA FOR FILTERABLE COD REMOVAL,
0.5 m RBC, CARBON OXIDATION PLUS NITRIFICATION MODE

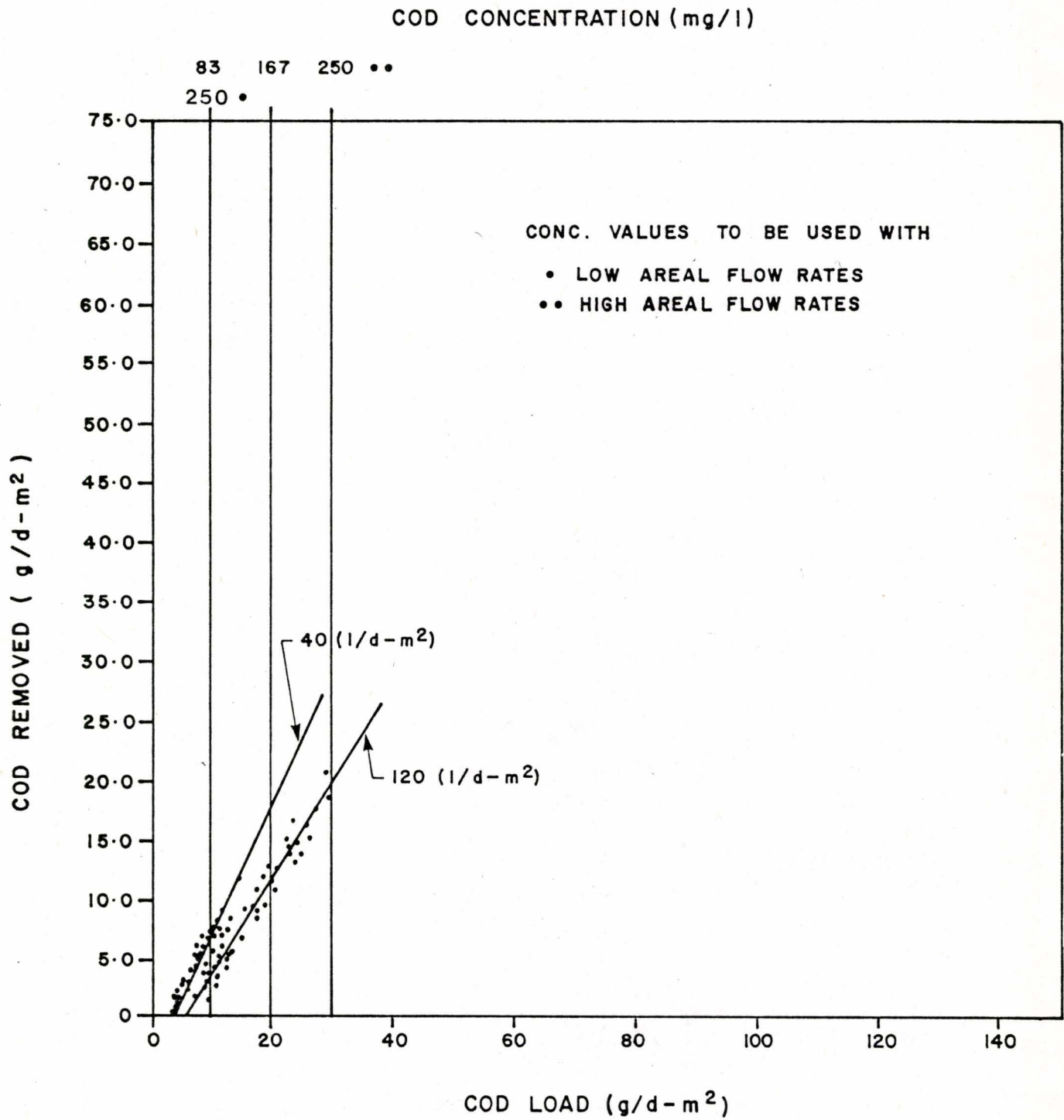
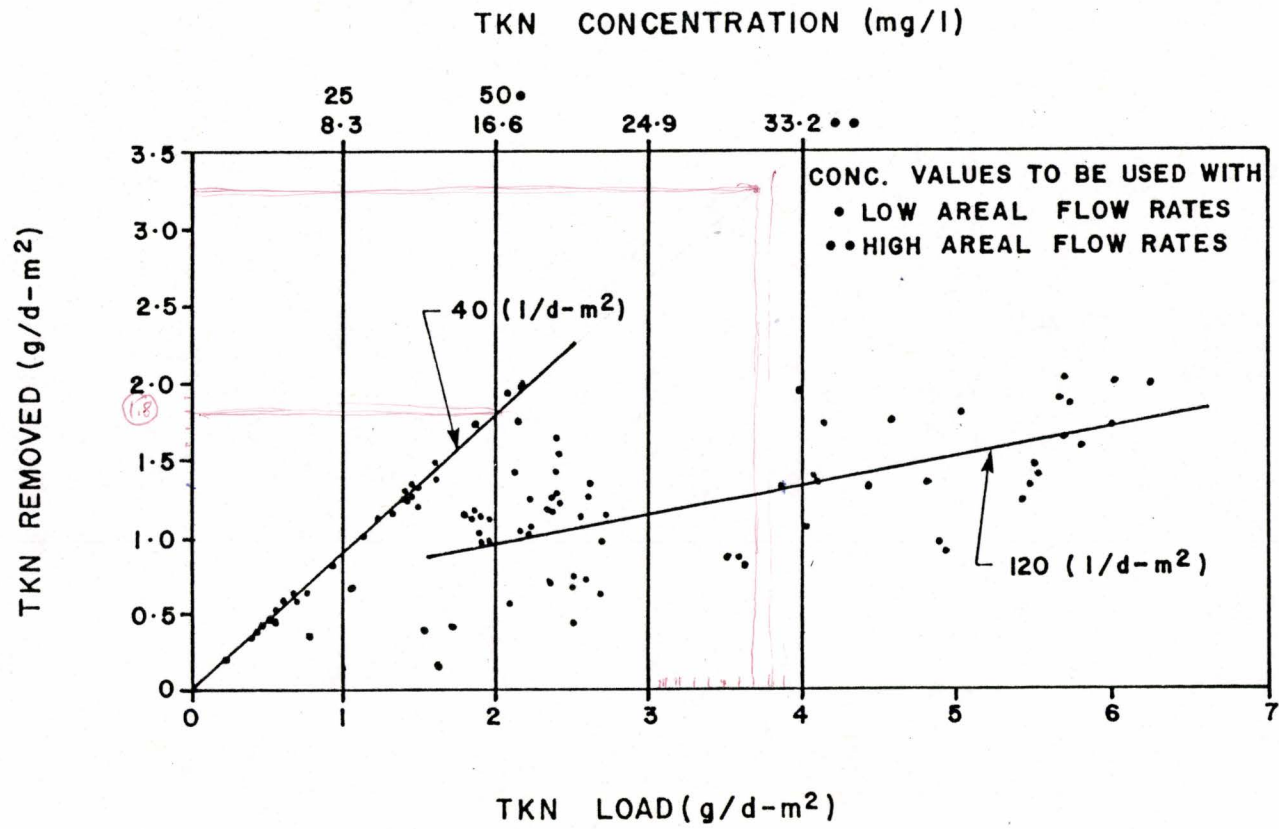


FIGURE 4-16

STEADY STATE DESIGN DATA FOR FILTERABLE TKN REMOVAL,
0.5m RBC, CARBON OXIDATION PLUS NITRIFICATION MODE



4.6.3 Use of Mass Loading Plots in Design

Figures 4.10 to 4.16 may be used for design purposes when considering influent loading conditions of minimal variability. The calculation procedure to determine the bio-disc surface area required to provide effective biological treatment is as follows:

1. Determine the influent concentration and flow rate for design.
2. Determine the effluent quality desired and from this, the mass removal required.
3. Compare the known influent concentration with the concentration isopleths on the plots. This will define the areal mass loading, RBC surface area, areal hydraulic loading and the areal mass removal. If the calculated mass removal is not sufficient to provide the predetermined effluent quality, then a larger surface area is required, thereby reducing the areal mass loading, and improving treatment. An iterative procedure must be continued until adequate mass removal is obtained.

4.7 Scale-Up

Scale-up is always an important consideration in any pilot scale study. Some correlation between the results obtained on a pilot scale unit and a full scale unit is necessary before the results obtained in the study are readily applicable. Since the 0.5 metre RBC and the 2.0 metre RBC were operated in parallel, and their areal loadings were comparable (within 3.5% at the high flow rate and 0.6% at the low flow rate) it was possible to evaluate the mass removal capabilities of the units (Figures 4.17 and 4.18).

FIGURE 4-17
COMPARISON OF TOC MASS REMOVAL 0.5m RBC
VERSUS 2.0m RBC

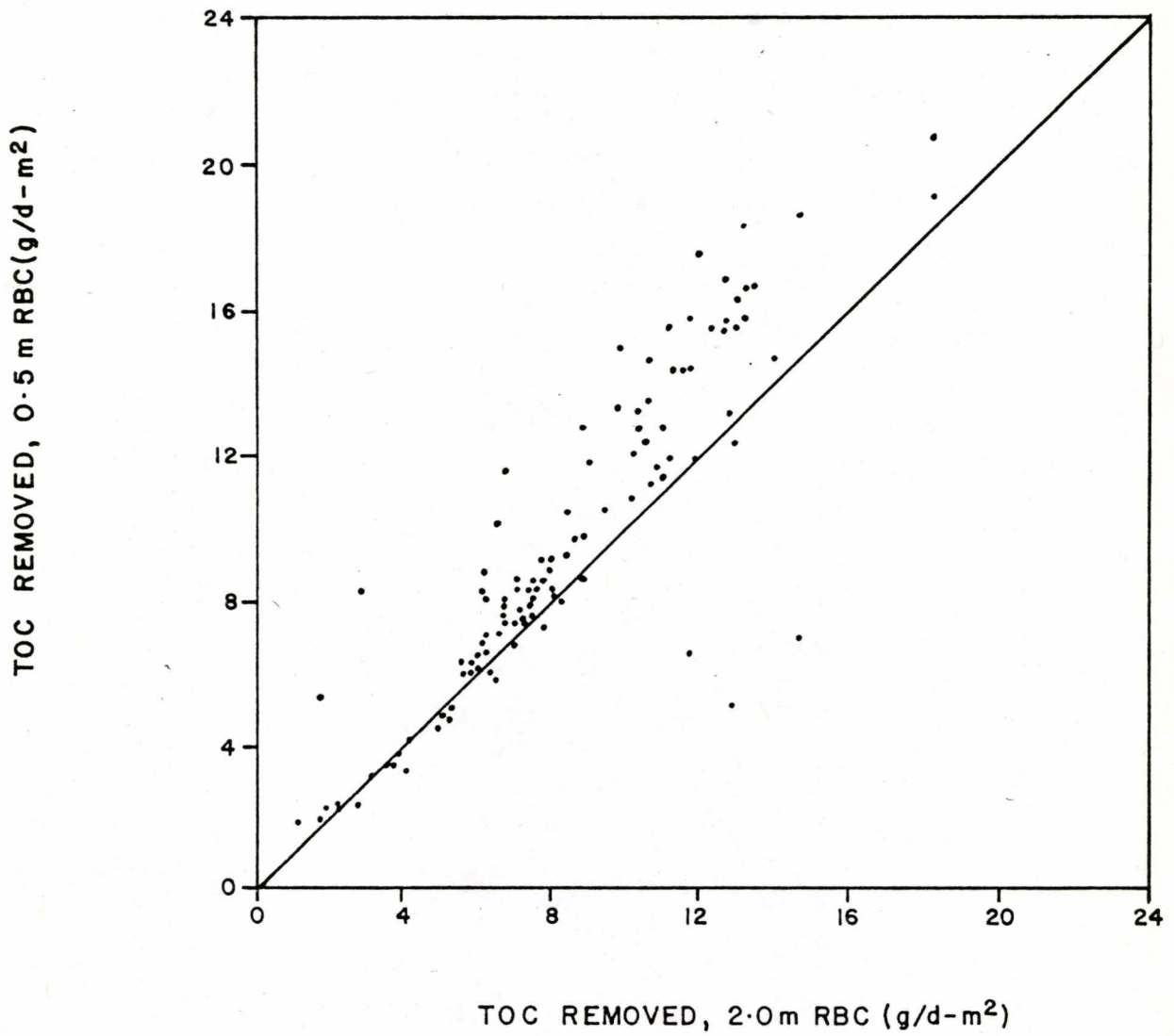
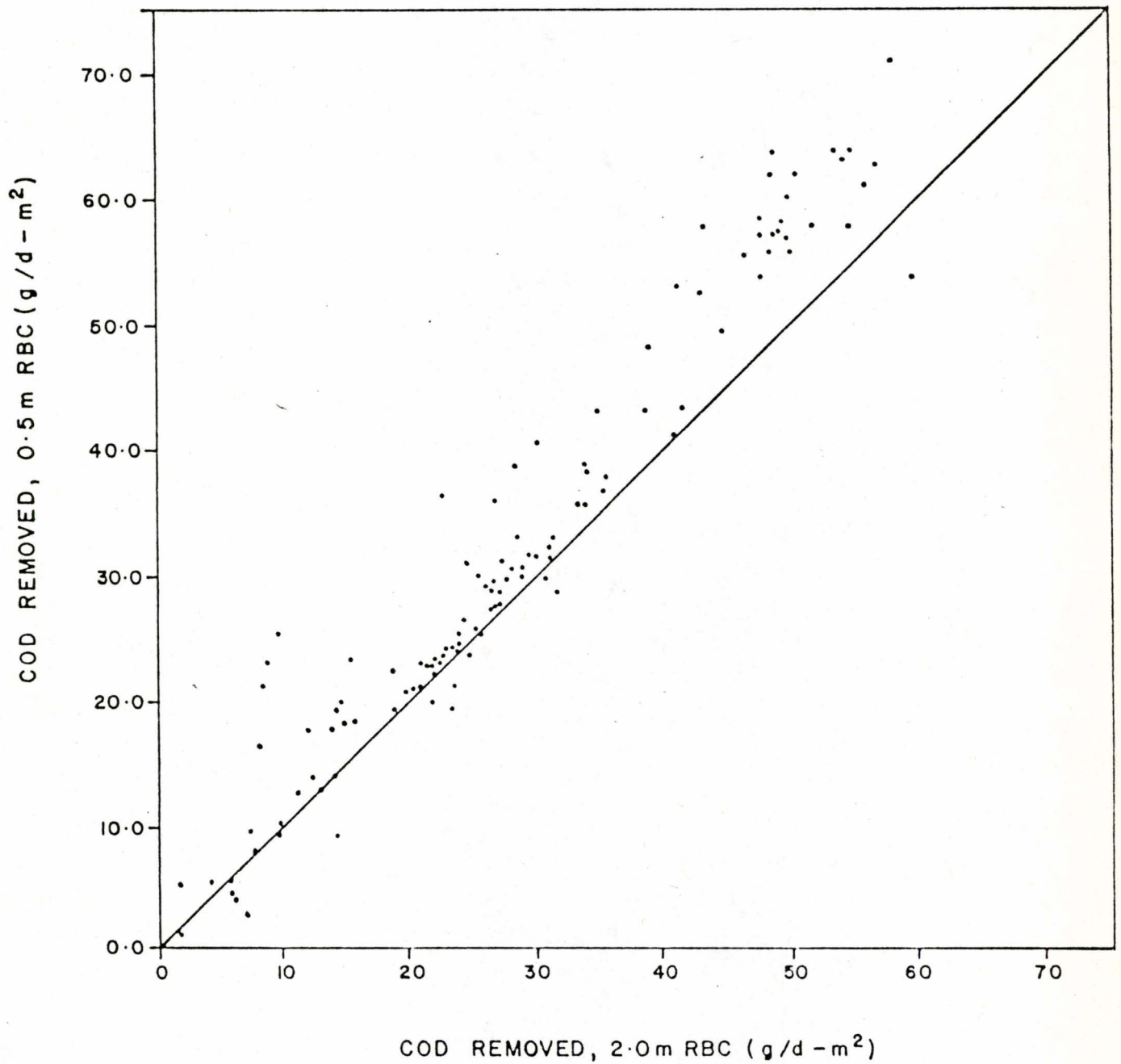


FIGURE 4-18
COMPARISON OF COD MASS REMOVAL 0.5 m RBC
VERSUS 2.0m RBC



As discussed in Section 4.2.2, a difference in performance of the two RBCs was observed through examination of the cross correlation results (Figures 4.4 and 4.5). The paired data shown on Figures 4.17 and 4.18 for TOC and COD mass removal demonstrate that the 0.5 metre RBC oxidized more carbon per unit surface area than did the 2.0 metre RBC. The difference in mass removal increases with loading to the units.

The mixing characteristics of the two reactors, as determined by dye studies (Appendix D), showed that mixing in the 0.5 metre RBC approached two CSTRs in series, while mixing in the 2.0 metre unit approached plug flow (7 CSTRs in series). If the biological oxidation reaction proceeds by first order kinetics, as indicated by Antonie (1970), then one would expect the 2.0 metre unit to have better removal than the 0.5 metre unit. However, improved performance was not observed, suggesting that the reaction does not follow a first order relationship and/or that some other effect is limiting the reaction.

Chesner and Molof (1976) demonstrated that at increased loading rates, dissolved oxygen concentration tends to decrease. They pointed out that as present design maintains a constant peripheral tip velocity, rotational speeds must decrease with increasing disc diameter, thereby decreasing aeration capacity. A dissolved oxygen limitation may therefore occur as loading and reactor size are increased. Dissolved oxygen was not regularly monitored during the experimental runs; therefore, one can only assume that this would explain the observations.

5.0 Conclusions

An evaluation of the response of rotating biological contactors (RBCs) operated under non-steady influent conditions indicates that:

1. The RBC is sensitive to influent fluctuations of an organic, inorganic and hydraulic nature in terms of maintaining consistently good effluent quality.
2. A statistically adequate representation of the dynamic TKN effluent response can be obtained using time series modelling techniques.
3. More emphasis should be placed on using influent mass loading as a design criteria.
4. The models developed indicate that:
 - a) the response in effluent filterable TKN is predicted most precisely by influent filterable TKN loading. Models based on influent TKN concentration and flow were not as precise in predicting effluent response;
 - b) positive effluent TKN response can be expected for increases in TKN loading, TKN concentration and hydraulic loading;
 - c) effluent response occurs almost instantaneously, with a duration of 3 to 4 hours; and
 - d) greater effluent variability may be expected with an RBC as compared to either a separate or combined activated sludge system operating under nitrifying conditions and at similar levels of removal.

5. Cross correlation results for carbon oxidation indicate that:
 - a) a positive TOC response can be expected for increases in TOC loading and concentration with the 0.5 metre RBC;
 - b) there is little correlation between effluent carbon concentration and hydraulic loading with the 0.5 metre RBC;
 - c) a positive TOC response can be expected for increases in TOC loading, concentration and hydraulic loading with the 2.0 metre RBC; and
 - d) the duration of the response is on the order of 1 hour.

6. An analysis of the raw data indicated that:
 - a) TOC and COD removal is a linear function of mass loading;
 - b) TKN mass removal is a linear function of TKN mass loading at loadings below 2.1 g/d-m^2 ;
 - c) TKN mass removal becomes unstable at loadings above 2.1 g/d-m^2 . The instability appears to be a function of flow rate rather than mass loading; and
 - d) areal organic mass removal was greater for the 0.5 metre unit when operated in parallel with the 2.0 metre unit.

6.0 Recommendations for Future Work

1. A detailed investigation at the pilot scale level to develop further loading data on which to base design, under both nitrifying and carbon oxidation modes of operation is recommended.
2. Develop transfer function-noise models based on the carbon oxidation data obtained in this study and examine the significance of mass carbon loading in design.
3. Develop design criteria for full scale RBCs based on pilot scale operating data. The investigation should encompass the mechanical scale-up consideration on which RBCs are currently designed.

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APPENDIX ADATA LISTING

This appendix contains a complete listing of the general operating procedures, including flow rates, influent and effluent temperatures, dissolved oxygen concentration, suspended solids, chemical additions, etc. as well as the analytical results obtained during the experimental runs. The results are presented in chronological order. Abbreviations and symbols used in the listings are defined in Appendix E.

GENERAL OPERATING DATA

SERIES A -START UP AND ACCLIMITIZATION

A-1 HYDRAULIC VOLUME AND DYE STUDY(0.5M UNIT)

C25 05 0.5 M1 RPM=13.0 WATER.FEED CONNECTED.HOOD INSTALLED
C 31 05 0.5 M1 DYE STUDY AT 0.813 L/M. RAW SEWAGE STARTED AT 1705
C01 06 0.5 M1 VOLUME=141.8 LITRES

GENERAL OPERATING DATA

A-2 ACCLIMITIZATION PERFORMANCE OF 0.5M1 + 0.5M2 UNITS

DAYMON	PILOT UNIT	SAMPLE IDENTITY	FLOW L/M	PH	ALK MG/L	SS MG/L	VSS MG/L	30 MIN SETTLE ML/L	DO MG/L	TEMP DEG-C
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01 06	0.5M1	RFG12	0.76							28.5
01 06	0.5M1	EFG12	0.76	7.7						23.8
01 06	0.5M1	AMB.AIR								20.7
01 06	0.5M1	INT.AIR								21.4

02 06	0.5M1	RFG08	0.68	6.7					1.3	28.0
02 06	0.5M1	EFG08	0.68	7.6					5.9	23.4
02 06	0.5M1	AMB.AIR								22.8
02 06	0.5M1	INT.AIR								23.2

C LIGHT FILM OF BROWN SLIME APPEARS IN STAGE 1

03 06	0.5M1	RFG08	0.65	7.0					4.0	26.3
03 06	0.5M1	EFG08	0.65	7.9					6.0	23.7
03 06	0.5M1	AMB.AIR								24.3
03 06	0.5M1	INT.AIR								24.1

04 06	0.5M1	RFG08	0.81	7.0					1.2	25.6
04 06	0.5M1	EFG08	0.81	8.0					7.1	24.6
04 06	0.5M1	EFC0807	0.81			50	45			23.8
04 06	0.5M1	AMB.AIR								23.8
04 06	0.5M1	INT.AIR								24.3

C GROWTH IS VERY HEAVY IN STG.1 AND LIGHT IN STG.2,3+4

C SECOND BIO SURF INSTALLED(0.5 M2) RPM=13.0 04/06/76

07 06	0.5M1	RFG08	0.81	6.9					0.8	28.7
07 06	0.5M1	EFG08	0.81	7.1					6.4	25.2
07 06	0.5M1	AMB.AIR								26.0
07 06	0.5M1	INT.AIR								25.9

07 06	0.5M2	RFG08	0.90	6.9					0.8	28.4
07 06	0.5M2	EFG08	0.90	8.0					6.9	23.6
07 06	0.5M2	AMB.AIR								26.0

08 06	0.5M1	RFG08	0.84	6.7					1.0	25.5
08 06	0.5M1	EFG08	0.84	7.0					6.1	25.8
08 06	0.5M1	EFC0807	0.84			42	34			26.4
08 06	0.5M1	AMB.AIR								26.4
08 06	0.5M1	INT.AIR								26.5

C HEAVY GROWTH IN FIRST 3 STAGES

GENERAL OPERATING DATA

DAYMON	PILOT UNIT	SAMPLE IDENTITY	FLOW L/M	PH	ALK MG/L	SS MG/L	VSS LMG/L	30 MIN SETTLE ML/L	DO MG/L	TEMP DEG-C
08 06	0.5M2	RFG08	0.85	6.7					1.0	25.5
08 06	0.5M2	EFG08	0.85	7.6					6.8	24.3
08 06	0.5M2	EFC0807	0.85			122	87			
08 06	0.5M2	AMB.AIR								26.4
C GROWTH HEAVY IN STG.1 AND LIGHT IN STG.2,3+4										
09 06	0.5M1	RFG08	0.77	6.8					1.0	27.2
09 06	0.5M1	EFG08	0.77	7.4					5.5	26.3
09 06	0.5M1	AMB.AIR								26.9
09 06	0.5M1	INT.AIR								26.8
09 06	0.5M2	RFG08	0.91	6.8					1.0	27.2
09 06	0.5M2	EFG08	0.91	7.3					6.2	24.9
09 06	0.5M2	AMBAIR								26.9
10 06	0.5M1	RFG08	0.75	6.8					1.2	25.8
10 06	0.5M1	EFG08	0.75	7.0					5.8	26.1
10 06	0.5M1	EFC0807	0.75			68	43			
10 06	0.5M1	AMB.AIR								26.4
10 06	0.5M1	INT.AIR								26.4
10 06	0.5M2	RFG08	0.82	6.8					1.2	25.8
10 06	0.5M2	EFG08	0.82	7.3					6.5	24.7
10 06	0.5M2	EFC0807	0.82			167	116			
10 06	0.5M2	AMB.AIR								26.4
C HOOD INSTALLED AT 1130										
11 06	0.5M1	RFG08	0.74	6.8					1.2	27.8
11 06	0.5M1	EFG08	0.74	7.5					5.5	26.0
11 06	0.5M1	AMB.AIR								25.1
11 06	0.5M1	INT.AIR								25.2
11 06	0.5M2	RFG08	0.85	6.8						27.5
11 06	0.5M2	EFG08	0.85	7.5					1.2	25.4
11 06	0.5M2	AMB.AIR							6.1	25.1
11 06	0.5M2	INT.AIR								25.2
14 06	0.5M1	RFG08	0.73	6.8					0.4	18.5
14 06	0.5M1	EFG08	0.73	7.0					6.4	21.2
14 06	0.5M1	AMB.AIR								27.1
14 06	0.5M1	INT.AIR								26.7

C RAW FEED LINE CHANGED CAUSING COOLER TEMPERATURES

GENERAL OPERATING DATA

DAYMON	PILOT UNIT	SAMPLE IDENTITY	FLOW L/M	PH	ALK MG/L	SS MG/L	VSS LMG/L	30 MIN SETTLE ML/L	DO MG/L	TEMP DEG-C
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25 06	0.5M1	RFG08	0.61	6.7					1.3	20.0
25 06	0.5M1	EFG08	0.61	7.0					6.4	22.5
25 06	0.5M1	AMB.AIR								26.6
25 06	0.5M1	INT.AIR								26.2

25 06	0.5M2	RFG08	1.01	6.7					1.1	20.2
25 06	0.5M2	EFG08	1.01	7.0					6.2	22.1
25 06	0.5M2	AMB.AIR								26.6
25 06	0.5M2	INT.AIR								26.0

C FURTHER COMPOSITES WERE TAKEN OF BOTH UNITS (0.5M)
 C MORE DATA FOR COMPARATIVE PURPOSES

16 08	0.5M1		3.27							
16 08	0.5M2		3.27							

17 08	0.5M1	RFG12	3.27	7.6					2.5	21.0
17 08	0.5M1	EFG12	3.27	7.7					5.7	21.3

17 08	0.5M1+2AMB.AIR									25.0
17 08	0.5M1+2RFC1009				370	186				

17 08	0.5M2	RFG12	3.25	7.6					2.5	21.0
17 08	0.5M2	EFG12	3.25	7.7					5.5	21.4

22 08	0.5M1+2RFC1413				94	84				
22 08	0.5M1		3.27							
22 08	0.5M2		3.25							

23 08	0.5M1+2RFC1313				98	90				
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23 08	0.5M1		3.04							
23 08	0.5M2		3.04							

SAMPLES COLLECTED ON 23/08 SAT IN FRIDGE FOR 24HR. BEFORE PREPARATION.

C 24 08 1500HR. 0.5M1 MOVED OUTSIDE AND CONNECTED TO SAME FEED AS 2.0M

C END OF EXPERIMENT

GENERAL OPERATING DATA

A-3 HYDRAULIC VOLUME AND DYE STUDY OF 2.0M UNIT

07 2.0 M. RAW SEWAGE. STARTED USING FEED SCOOPS. 1300-FEED SHUTOFF
07 2.0 M RAW SEWAGE STARTED USING DIRECT FEED RPM=3.0
07 2.0 M B10 SURF NOT LEVEL. WIER SET TO MINIMUM HEIGHT
07 2.0 M BASIC VOLUME (NO FLOW CONDITIONS)=3364.0 LITRES
07 2.0 M ACCUMULATED SLUDGE IN THE BOTTOM OF EACH STAGE
07 2.0 M B10 SURF SHUT OFF FOR 6.0HR WHILE PIPING INSTALLED
07 2.0 M DYE STUDY AT 37.2L/M. HEAD VOL=104.4L. VOLUME=3468.4 L.
07 2.0M DYE STUDY AT 100.3 L/M. HEAD VOL=181.6L. TOTAL VOL=3545.6 L.
07 2.0 M PUMP SHUT OFF AT 1400 B10 SURF STILL ROTATING
07 2.0 M PUMP STARTED UP AT 1900. STG1+2 SHOW A HEAVY GROWTH

END OF EXPERIMENT

GENERAL OPERATING DATA

DAYMON	PILOT UNIT	SAMPLE IDENTITY	FLOW L/M	PH	ALK MG/L	SS MG/L	VSS LMG/L	30 MIN SETTLE ML/L	DO MG/L	TEMP DEG-C
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03 08	2.0 M	RFG08	111.68	6.8						18.7
03 08	2.0 M	EFG08	111.68	6.8						17.7
03 08	2.0 M	STAGE1	111.68							
03 08	2.0 M	7 STAGE4	111.68					4.0	17.7	
03 08	2.0 M	AMB.AIR								16.0

C FEED PUMP IS PUMPING SOME AIR .MAY AFFECT D.O.

04 08	2.0 M	RFG08	122.6	6.7						19.5
04 08	2.0 M	RFC0807			200					
04 08	2.0 M	EFG08	122.6	6.8						16.4
04 08	2.0 M	STAGE1	122.6					4.1	18.7	
04 08	2.0 M	STAGE4	122.6					4.6	16.4	
04 08	2.0 M	AMB.AIR								19.8

~~C STILL PUMPING AIR. SYSTEM WAS RECALIBRATED IN THE PM OF THE 3RD
C PUMP SHUT OFF DURING THE NIGHT . MAY AFFECT COMPOSITE
C ELECTRICAL FAILURE IN FLOW MONITOR CAUSED SHUT DOWN AT 1400~~

C PUMP RESTARTED AT 0900. COMPOSITE STARTED AT 1000 -0508

05 08	2.0 M	RFG10	124.8	6.7						18.7
05 08	2.0 M	RFC1009			408	351				
05 08	2.0 M	EFG10	124.8	6.8						20.4
05 08	2.0 M	STAGE1	124.8					3.4	19.2	
05 08	2.0 M	STAGE4	124.8					4.2	19.4	
05 08	2.0 M	AMB.AIR								20.7

C HOOD INSTALLED ON 0608

C END OF EXPERIMENT. A TOTAL OF 4 COMPOSITE SAMPLES WERE COLLECTED

GENERAL OPERATING DATA

LYMON	PILOT UNIT	SAMPLE IDENTITY	FLOW L/M	PH	ALK MG/L	SS MG/L	VSS LMG/L	30 MIN SETTLE ML/L	DO MG/L	TEMP DEG-C
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15 07	0.5M1	RFC1009	.	.		216	162			
15 07	0.5M1	RFG08	.	.					19.6	
15 07	0.5M1	EFG08							20.3	

15 07	0.5M2	RFC1009				325	235			
15 07	0.5M2	RFG08							19.7	
15 07	0.5M2	EFG08							20.3	
15 07	0.5M2	AMB AIR							24.4	

24HR.COMPOSITES PLUS HOURLY SAMPLING ON 07+08.

07 07	0.5M1	RFC1009				204	169			
07 07	0.5M2	RFC1009				200	170			

08 07	0.5M1	RFG08		6.4				0.4	20.1	
08 07	0.5M1	RFC1009				227	179			
08 07	0.5M1	STAGE 1		6.3				2.8	21.4	
08 07	0.5M1	STAGE 4		6.2				4.1	22.4	
08 07	0.5M1	AMB.AIR							25.3	
08 07	0.5M2	INT.AIR							24.9	

08 07	0.5M2	RFG08	1.08	6.3				0.8	20.1	
08 07	0.5M2	RFC1009				185	149			
08 07	0.5M2	STAGE 4	1.08	6.0				4.2	21.9	
08 07	0.5M2	AMB.AIR							25.3	
08 07	0.5M2	INT.AIR							25.0	

IT WAS NOT POSSIBLE TO OBTAIN ACCESS TO STAGE 1

09 07	0.5M1	RFG08		6.4	266			0.5	20.0	
09 07	0.5M1	STAGE 1		6.4				3.2	21.0	
09 07	0.5M1	STAGE 4		5.9				4.0	21.8	
09 07	0.5M1	AMB.AIR							25.8	
09 07	0.5M1	INT.AIR							24.9	
09 07	0.5M1	EFG08			198					

09 07	0.5M2	RFG08	1.11	6.5	264			1.1	20.2	
09 07	0.5M2	STAGE 1	1.11	6.3				4.2	21.6	
09 07	0.5M2	AMB.AIR							25.8	
09 07	0.5M2	INT.AIR							24.8	
09 07	0.5M2	EFG08			179					

ANOTHER COMPOSITE SAMPLE RUN WILL BE DONE AT A LATER DATE

GENERAL OPERATING DATA

21 07 COMPOSITE SAMPLE 1110 . 22/07 COMPOSITE SAMPLE TAKEN
 THE FLOW SPLITTER WAS ADDED ON 16/07 AT 1500.5 DAY ACCLIMITIZATION

AYMON	PILOT SAMPLE	FLOW	PH	ALK	SS	VSS	30 MIN	DO	TEMP
UNIT	IDENTITY	L/M		MG/L	MG/L	MG/L	SETTLE	MG/L	DEG-C
							ML/L		

21 07	0.5M1	RFG11							19.8
21 07	0.5M1	EFG11							20.9
21 07	0.5M1	AMB.AIR							24.1

21 07	0.5M2	RFG11							19.8
21 07	0.5M2	EFG11							20.7

22 07	0.5M1	RFG08							19.4
22 07	0.5M1	EFG08							20.3
22 07	0.5M1	AMB.AIR							22.8

22 07	0.5M2	RFG08							19.4
22 07	0.5M2	EFG08							20.6

END OF EXPERIMENT

GENERAL OPERATING DATA

SERIES B-2 ORGANIC CARBON REMOVAL MODE

TIME	FLOW (L/M)	WEIGHTED VOL. (MLS)	TIME	FLOW (L/M)	WEIGHTED VOL. (MLS)
0	2.52	74	13	3.06	90
1	2.22	65	14	3.42	100
2	2.06	60.7	15	4.65	137
3	1.91	56	16	5.86	172
4	1.84	54	17	6.54	192
5	1.82	53	18	6.81	200
6	1.82	53	19	6.58	193
7	1.82	53	20	6.36	173
8	1.82	53	21	4.99	141
9	1.91	56	22	3.40	100
10	2.04	60	23	2.84	83
11	2.27	67	24	2.52	74
12	2.63	77	AVERAGE=3.36 L/M.		

AYMON UNIT	PILOT SAMPLE IDENTITY	FLOW L/M	PH	ALK MG/L	SS MG/L	VSS MG/L	30 MIN SETTLE ML/L	DO MG/L	TEMP DEG-C
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23/07 TO 28/07 . ACCLIMITIZATION

23 07 0.5M2 3.13
 23 07 0.5M1 FLOW FROM FLOW CONTROLLER. FLOWS ARE LISTED .

28 07 24HR. COMPOSITE BEGAN AT 0900

28 07 0.5M1	RFC0908	275	241
28 07 0.5M1	EFC0908	202	
28 07 0.5M2	RFC0908	146	130
28 07 0.5M2	EFC0908	333	

29 07 0.5M1	1000	2.0			
29 07 0.5M1	RFG09	6.9			
29 07 0.5M1	EFG09	6.8			
29 07 0.5M1	STAGE1			3.7	20.8
29 07 0.5M1	STAGE4			4.3	21.2
29 07 0.5M1	AMB. AIR				23.1

29 07 0.5M2	RFG09	3.4	6.9		
29 07 0.5M2	EFG09	3.4	6.8		
29 07 0.5M2	STAGE1	3.4	6.8		20.8
29 07 0.5M2	STAGE4	3.4	6.8	4.2	20.7
29 07 0.5M2	AMB. AIR				23.1

30 07 WASTED SLUDGE FROM THE SEWAGE PLANT IN THE INFLUENT AT 0900

04 08 0.5M1	STAGE1	6.5		1.8	20.4
04 08 0.5M1	STAGE4	7.0		3.8	21.0
04 08 0.5M1+2	AMB. AIR				22.8
04 08 0.5M2	STAGE1	6.5		1.8	20.2
04 08 0.5M2	STAGE4	6.9		3.8	21.0

GENERAL OPERATING DATA

DAYMON	PILOT UNIT	SAMPLE IDENTITY	FLOW L/M	PH	ALK MG/L	SS MG/L	VSS LMG/L	30 MIN SETTLE ML/L	DO MG/L	TEMP DEG-C
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04	08	0.5M1	RFC0908			253	181			
04	08	0.5M1	EFC0908			197				
04	08	0.5M2	RFC0908			256				
04	08	0.5M2	EFC0908			228				

05	08	0.5M1	1530	5.13						
05	08	0.5M1	RFG15		6.0				1.4	20.4
05	08	0.5M1	EFG15		6.06				3.0	21.2
05	08	0.5M2	RFG15	3.27	6.0				1.4	20.4
05	08	0.5M2	EFG15	3.27	6.8				3.1	21.5

06	08	0.5M1	RFG		6.7				1.7	20.5
06	08	0.5M1	EFG		6.9				3.0	20.8
06	08	0.5M1+2AMB.AIR								23.5
06	08	0.5M2	RFG15	3.5	6.7				1.7	20.5
06	08	0.5M2	EFG15	3.5	6.79				2.9	20.2

07	08	0.5M1	RFG13		6.7					20.6
07	08	0.5M1	EFG13		6.9					21.1
07	08	0.5M2	RFG13	3.47	6.7					20.6
07	08	0.5M2	EFG13	3.47	6.9					21.0

09	08	0.5M1	1530	5.08						
09	08	0.5M1	RFG							20.8
09	08	0.5M1	EFG							21.0
09	08	0.5M1+2AMB.AIR								24.3
09	08	0.5M2	EFG09	3.44						21.2
09	08	0.5M2	RFG09	3.44						20.8

END OF EXPERIMENT

GENERAL OPERATING DATA

SERIES C DYNAMIC RESPONSE (0.5M UNIT)

C-1 NITRIFICATION MODE

07 AVERAGE FLOW SET AT 1.3 L/M
 LL CHANGES TO FLOW DURING THIS RUN WERE MADE 15 MIN. BEFORE THE HOUR
 LL SPIKE CHANGES DURING THIS RUN WERE MADE 10 MIN. BEFORE THE HOUR
 H4CL=3G/L; NAHCO3= 9.0 G/L, DEXTROSE= 5G/L. THIS SOLUTION WAS ADDED AT 1100HR
 N 10/07 AT A FLOW RATE OF 12.9 ML/MIN UNTIL 2345HR ON 11/07
 SPIKE CONCENTRATIONS. TKN SPIKE=7.71G NH4CL + 23.13G NAHCO3 IN ONE LITRE
 TOC SPIKE=7.0G DEXTROSE IN ONE LITRE
 THESE ARE THE CONCENTRATIONS USED FROM 12/07 TO 17/07

F=FLOW OF RAW FEED IN L/M , DW=FLOW OF DILUTION WATER IN L/M
 DC=FLOW OF TOC SPIKE IN ML/MIN , TKN=FLOW OF TKN SPIKE IN ML/MIN

MON	PILOT UNIT	SAMPLE IDENTITY	FLOW L/M	PH	ALK MG/L	SS MG/L	VSS MG/L	30 MIN SETTLE ML/L	DO MG/L	TEMP DEG-C	CHEMICALS	RF	DW	TOC	TKN
07	0.5MI	0										.672	0	0	14.2
07	0.5MI	1200										.672	.46	0	14.2
07	0.5MI	1600										1.83	1.14		28.4
07	0.5MI	RFG12		6.8	368					19.0					
07	0.5MI	EFG12		7.0	190					20.3					
07	0.5MI	0										.672	0	0	0
07	0.5MI	0800										.672	0	14.2	14.2
07	0.5MI	1600										1.97	1.14	0	0
07	0.5MI	RFG10		7.6	390				3	20.5					
07	0.5MI	EFG10		7.3	230				4.3	21.0					
07	0.5MI	0										.454	0	14.2	0
07	0.5MI	0800										1.94	0	0	28.4
07	0.5MI	1200										1.94	1.14	39.7	0
07	0.5MI	1600										1.94	0	26.1	0
07	0.5MI	RFG10								21.5					
07	0.5MI	EFG10								22.5					
07	0.5MI	0										.633	0	13.0	17.0
07	0.5MI	0800										1.94	0	26.0	34.0
07	0.5MI	1300										1.94	0	0	26.1
07	0.5MI	1600										1.94	0	0	0
07	0.5MI	RFG14		7.2						20.5					
07	0.5MI	EFG14		7.1	360					21.0					
07	0.5MI	0										1.94	0	0	0
07	0.5MI	0800										.672	0	13.0	0
07	0.5MI	1200										.672	0.57	13.0	0
07	0.5MI	1400										.672	0.57	0	0
07	0.5MI	1600										1.94	0	0	0

GENERAL OPERATING DATA

ION	PILOT UNIT	SAMPLE IDENTITY	FLOW L/M	PH	ALK MG/L	SS MG/L	VSS LMG/L	30 MIN SETTLE ML/L	DO MG/L	TEMP DEG-C	CHEMICALS			
											RF	DW	TO	TKN
07	0.5MI	RFG14								21.0				
07	0.5MI	EFG14								22.1				
07	0.5MI	0									.672	0	0	22.7
07	0.5MI	1600		END	OF	EXPERIMENT					0	0	0	0

BIO-SURF PILOT PLANT TESTING

1976

ANALYTICAL RESULTS - SERIES A1 + A2 (ACCLIMATIZATION)

MON	PILOT UNIT	SAMPLE IDENTITY	SETTLED SAMPLE			UNFILTERED SAMPLE			FILTERED SAMPLE						
			BOD	COD	TKN	BOD	COD	TKN	BOD	COD	TOC	NH3N	NO2N	NO3N	TKN
06	.5M1	EFC0908				35	204	20.7	15	46	26	13.0	0.2	0.2	16.2
06	.5M1+2	RFC0908										14.6	0.1	0.1	
06	.5M1	EFC0908				19	108	20.6	7	21	18	13.0	0.4	0.2	15.8
06	.5M1+2	RFC0908				91	293	26.2	33	105	31	17.0	0.1	1.6	19.9
06	.5M1	EFC0908				16	96	3.8	5	33	17	0.8	10.1	0.0	2.3
06	.5M2	EFC0908				30	138	19.6	9	75	22	15.0	0.4	0.0	15.4
06	.5M1+2	RFC0908				100	264	24.1	32	92	33	16.0	0.0	0.0	17.9
06	.5M1	EFC0908				16	75	6.2	6	59	15	0.8	17.0	1.5	1.7
06	.5M2	EFC0908				35	205	18.9	9	54	19	12.0	1.2	0.0	13.6
06	.5M1+2	RFC0908										18.0	0.0	0.0	
06	.5M1	EFC0908				15	96	4.4	4	75	10	2.0	3.6	8.0	3.3
06	.5M2	EFC0908				18	155	5.5	4	54	10	1.0	9.1	0.0	2.7
06	.5M1	EFC0908				23	192	6.1	3	42	10	1.0	2.9	7.8	2.6
06	.5M2	EFC0908				23	172	9.7	5	46	10	1.0	9.7	0.0	2.2
06	.5M1+2	RFC0908				100	310	25.2	32	96	36	19.0	0.0	0.2	19.2
06	.5M1	EFC0908				42	190	6.4	4	54	10	0.9	1.0	6.7	1.6
06	.5M2	EFC0908				34	206	4.7	4	54	10	0.7	5.6	1.3	3.0
08	.5M1+2MRFC	1009	27.9			109	233	31.8		90	32	20.0	0.0	0.5	21.4
08	.5M1	EFC1009	22	46	20.8	109		22.1		60	17	14.0	0.3	0.6	18.8
08	.5M2	EFC1009	18	37	21.6			23.1		60	18	18.0	0.1	0.2	19.4
08	.5M1+2MRFC	1413	108			56	226	22.1		63	19	15.0	0.2	0.3	17.4
08	.5M1	EFC1413		133	17.6			19.7		49	14	13.0	1.2	1.1	15.1
08	.5M2	EFC1413	22	97	19.5			21.6		50	15	14.0	0.5	0.8	15.6

BIO-SURF PILOT PLANT TESTING
1976

ANALYTICAL RESULTS - SERIES B1 (COMPOSITES)

MON	PILOT	SAMPLE	SETTLED			UNFILTERED			FILTERED						
			UNIT	IDENTITY	SAMPLE	BOD	COO	TKN	BOD	COO	TKN	BOD	COO	TKN	NO ₂ N
	06	.5M1	RFC0908			17.8	81	194	51.8	64	32	11.0	0.1	0.6	15.8
	06	.5M1	EFC0908	8	69	5.5			19.9	49	14	4.6	1.0	4.2	4.8
	06	.5M2	RFC0908			16.8	72	205	24.2	62	26	11.0	0.1	0.7	16.2
	06	.5M2	EFC0908	11	117	7.3			11.8	42	14	1.0	1.5	2.9	5.2
	07	.5M1	RFC0908			13.9	103	320	25.4	66	32	5.0	0.0	0.2	9.5
	07	.5M1	EFC0908	12	75	5.9			8.0	32	15	2.9	0.8	3.3	3.0
	07	.5M2	RFC0908			17.8	78	219	17.3	59	27	8.0	0.0	0.4	9.6
	07	.5M2	EFC0908	9	65	3.2			4.8	41	13	0.6	0.6	3.2	1.7
	07	.5M1	RFC0908			25.5	119	340	27.4	73	46	18.5	0.0	0.3	20.4
	07	.5M1	EFC0908	14	103	12.3			15.2	66	27	7.0	0.1	5.3	10.0
	07	.5M2	RFC0908			22.9	88	298	24.7	68	39	14.0	0.0	0.2	19.8
	07	.5M2	EFC0908	10	77	8.3			11.2	58	22	4.0	0.7	5.9	7.3
	07	.5M1	RFC0908			27.2	137	376	30.1	70	48	16.0	0.1	0.1	20.1
	07	.5M1	EFC0908	19	89	10.9			16.7	54	22	7.2	0.5	4.6	9.0
	07	.5M2	RFC0908			23.5	103	322	25.1	56	37	14.0	0.0	0.0	17.1
	07	.5M2	EFC0908	14	94	6.7			11.2	49	16	3.4	0.9	5.3	4.9
	07	.5M1	RFC0908			24.7	131	359	30.3	0	40	15.0	0.4	0.3	19.6
	07	.5M1	EFC0908	30	98	12.3			15.3	0	18	7.2	0.7	4.4	9.2
	07	.5M2	RFC0908			27.5	119	290	26.3	0	36	15.0	0.2	0.0	17.2
	07	.5M2	EFC0908	37	83	5.9			10.1	0	15	3.0	1.5	5.3	4.3
	07	.5M1	RFC1110			299	148	448	315	148	50	28.0	0.4	0.7	29.8
	07	.5M1	EFC1110	30	120	16.0			23.0	78	21	10.8	2.6	2.8	14.2
	07	.5M2	RFC1110			237	134	415	24.8	130	38	21.0	0.3	0.5	21.9
	07	.5M2	EFC1110	11	87	5.8			11.7	65	17	4.0	1.2	2.7	4.2

FEED AND EFFLUENT SAMPLES IN .5M1 WERE INTERCHANGED BY ACCIDENT

	07	.5M1	RFC1110			440	33.8		38.2	390	50	29.0	0.1	0.6	31.7	
	07	.5M1	EFC1110			315		145	369	27.9	162	7.0	2.5	2.2	9.2	
	07	.5M2	RFC1110			203		30	427	28.4	148	40	17.0	0.0	0.3	20.2
	07	.5M2	EFC1110	144	71	4.2			7.6	63	18	3.0	2.0	4.9	4.4	

BIO-SURF PILOT PLANT TESTING
1976

ANALYTICAL RESULTS - SERIES B2 (COMPOSITES)

MON	PILOT UNIT	SAMPLE IDENTITY	SETTLED SAMPLE			UNFILTERED SAMPLE			FILTERED SAMPLE						
			BOD	COD	TKN	BOD	COD	TKN	BOD	COD	TOC	NH3N	NO2N	NO3N	TKN
07	.5M1	RFC0908		230		106	255	25.8		101	31	21.0	0.0	0.0	21.9
07	.5M1	EFC0908	34	142	23.3			27.3		70	24	17.0	0.6	0.1	19.5
07	.5M2	RFC0908		180		105	272	26.7		75	30	14.0	0.0	0.0	19.9
07	.5M2	EFC0908	23	105	18.8			22.6		49	20	11.0	0.5	0.1	13.8
07	.5M1	RFC0908		192		114	293	21.3		88	25	14.0	0.1	0.5	17.7
07	.5M1	EFC0908	24	88	14.8		167	18.4		50	18	12.0	0.6	0.1	13.0
07	.5M2	RFC0908		117	17.8	29	251	25.9		60	18	13.3	0.6	0.1	15.9
07	.5M2	EFC0908	116	192	21.2		393	24.3		72	22	13.0	0.3	0.0	15.6
08	.5M1	RFC0908		241		109	328	29.1		98	34	13.0	0.0	0.4	16.1
08	.5M1	EFC0908	37	183	24.4			26.9		66	22	18.0	0.2	0.3	22.0
08	.5M2	RFC0908		261		79	274	26.4		82	29	17.0	0.0	0.2	21.3
08	.5M2	EFC0908	22	129	21.4			22.7		57	17	14.0	0.1	0.3	19.3
08	.5M1	RFC1514		266		109	353	31.3		91	29	18.0	0.0	0.4	23.5
08	.5M1	EFC1514	24	141	21.5			22.8		60	18	14.0	0.1	0.3	18.9
08	.5M2	RFC1514		224		106	295	29.2		74	25	18.0	0.0	0.3	21.2
08	.5M2	EFC1514	32	174	23.8			20.3		66	19	11.0	0.1	0.3	16.9
08	.5M1	RFC0908		213		131	303	30.9		84	33	19.0	0.0	0.2	23.2
08	.5M1	EFC0908	38	160	24.3			29.3		65	22	15.0	0.1	0.4	19.3
08	.5M2	RFC0908		225		103	254	26.8		74	26	18.0	0.0	0.2	20.8
08	.5M2	EFC0908	23	98	23.8			25.1		60	19	18.6	0.0	0.3	24.3
08	.5M1	RFC1009		242		103	303	27.0		93	32	23.0	0.0	0.4	24.1
08	.5M1	EFC1009	33	168	21.7			25.6		74	21	19.0	0.4	0.4	19.1
08	.5M2	RFC1009		213		72	250	23.5		76	30	20.0	0.0	0.2	20.9
08	.5M2	EFC1009	23	131	19.7			23.4		71	20	17.0	0.4	0.4	17.8

BIO-SUPP PILOT PLANT TESTING
1976

ANALYTICAL RESULTS - SERIES B1 (HOURLY)

Y	MON	PILOT	SAMPLE	FLOW RATE	UNFILTERED	FILTERED							
						UNIT	IDENTITY	IGPM	BOD	COD	TKN	BOD	COD
7	07	.5M1	RFG 900	0.130				41	15	8.3	0.3	0.6	11.3
7	07	.5M1	RFG1000	0.145				49	16	11.0	0.2	0.3	13.0
7	07	.5M1	RFG1100	0.162				49	15	15.0	0.0	0.1	17.6
7	07	.5M1	RFG1200	0.182				86	31	20.0	0.0	0.0	24.0
7	07	.5M1	RFG1300	0.210				102	47	23.0	0.2	0.0	25.8
7	07	.5M1	RFG1400	0.229				90	43	20.0	0.0	0.0	23.5
7	07	.5M1	RFG1500	0.324				169	78	21.0	0.0	0.0	28.4
7	07	.5M1	RFG1600	0.414				159	92	21.0	0.0	0.0	28.7
7	07	.5M1	RFG1700	0.471				153	70	19.0	0.0	0.0	25.3
7	07	.5M1	RFG1800	0.484				135	72	18.0	0.0	0.1	24.0
7	07	.5M1	RFG1900	0.459				142	74	19.0	0.0	0.0	24.5
7	07	.5M1	RFG2000	0.409				96	67	18.0	0.0	0.1	21.9
7	07	.5M1	RFG2100	0.329				98	58	16.0	0.0	0.0	19.9
7	07	.5M1	RFG2200	0.235				90	53	14.0	0.0	0.0	17.6
7	07	.5M1	RFG2300	0.205				57	39	13.0	0.0	0.0	16.2
7	07	.5M1	RFG2400	0.179				74	37	12.0	0.0	0.0	15.4
8	07	.5M1	RFG 100	0.155				65	32	13.0	0.0	0.0	14.5
8	07	.5M1	RFG -20	0.140				57	24	13.0	0.0	0.0	13.7
8	07	.5M1	RFG 300	0.125				51	23	12.0	0.0	0.0	14.1
8	07	.5M1	RFG 400	0.115				59	24	13.0	0.0	0.0	14.0
8	07	.5M1	RFG 500	0.115				53	19	12.0	0.0	0.0	13.0
8	07	.5M1	RFG 600	0.115				41	17	10.0	0.0	0.0	10.2
8	07	.5M1	RFG 700	0.115				45	17	9.0	0.1	0.0	9.4
8	07	.5M1	RFG 800	0.120				43	16	9.0	0.2	0.5	9.7
8	07	.5M1	RFG 900	0.130				47	15	8.0	0.3	0.3	9.9
8	07	.5M1	RFG1000	0.145				42	16	10.0	0.2	0.2	12.6
8	07	.5M1	RFG1100	0.162				48	17	14.0	0.2	0.0	16.8
8	07	.5M1	RFG1200	0.182				53	41	21.0	0.0	0.1	24.5
8	07	.5M1	RFG1300	0.210				73	50	22.0	0.0	0.0	25.4
8	07	.5M1	RFG1400	0.229				62	53	23.0	0.0	0.0	25.3
8	07	.5M1	RFG1500	0.324				82	60	20.0	0.0	0.0	24.6
8	07	.5M1	RFG1600	0.414				121	75	21.0	0.0	0.0	25.5
8	07	.5M1	RFG1700	0.471				101	59	18.0	0.0	0.0	20.7
8	07	.5M1	RFG1800	0.484				95	56	16.0	0.0	0.0	19.0
8	07	.5M1	RFG1900	0.459				190	61	18.0	0.1	0.9	19.8
8	07	.5M1	RFG2000	0.409				89	48	14.0	0.9	0.0	16.8
8	07	.5M1	RFG2100	0.329				95	57	16.0	0.1	0.0	20.0
8	07	.5M1	RFG2200	0.235				82	51	16.0	0.0	0.0	19.3
8	07	.5M1	RFG2300	0.205				62	33	12.0	0.0	0.0	13.8
8	07	.5M1	RFG2400	0.179				55	30	12.0	0.0	0.1	13.3
9	07	.5M1	RFG 100	0.155				69	41	14.0	0.0	0.0	16.6
9	07	.5M1	RFG 200	0.140				77	42	13.0	0.0	0.0	15.0
9	07	.5M1	RFG 300	0.125				54	24	13.0	0.5	0.0	14.6
9	07	.5M1	RFG 400	0.115				56	24	12.0	0.8	0.1	14.6
9	07	.5M1	RFG 500	0.115				54	31	12.0	0.4	0.4	13.8
9	07	.5M1	RFG 600	0.115				54	16	9.0	0.6	1.4	11.9
9	07	.5M1	RFG 700	0.115				49	16	8.0	0.3	2.1	10.9
9	07	.5M1	RFG 800	0.120				49	18	8.0	0.5	2.2	9.8
9	07	.5M1	RFG 900	0.130				45	14	2.0	8.0	14.1	11.0
9	07	.5M1	RFG1000	0.145				48	15	9.0	0.4	3.3	13.8
9	07	.5M1	RFG1100	0.162				63	22	11.0	0.3	1.0	17.7

ANALYTICAL RESULTS - SERIES B1 (HOURLY)

MON	PILOT	SAMPLE	FLOW RATE	UNIT IDENTITY	IGPM	UNFILTERED			FILTERED					
						BOD	COD	TKN	BOD	COD	TOC	NH3N	NO2N	NO3N
7	07	.5M1	EFG 900		0.130				57	15	0.9	0.2	7.1	1.4
7	07	.5M1	EFG1000		0.145				49	14	0.7	0.2	7.3	1.1
7	07	.5M1	EFG1100		0.162				45	14	0.8	0.3	7.4	1.8
7	07	.5M1	EFG1200		0.182				41	14	1.2	0.3	7.8	2.1
7	07	.5M1	EFG1300		0.210				41	14	2.3	0.5	8.3	3.0
7	07	.5M1	EFG1400		0.229				41	15	4.1	0.6	8.2	4.3
7	07	.5M1	EFG1500		0.324				41	15	6.5	0.6	7.8	6.6
7	07	.5M1	EFG1600		0.414				57	18	10.3	0.5	4.0	11.8
7	07	.5M1	EFG1700		0.471				57	22	12.8	0.5	2.0	14.4
7	07	.5M1	EFG1800		0.484				55	25	13.5	0.4	1.7	16.3
7	07	.5M1	EFG1900		0.459				61	25	12.5	0.4	1.5	13.7
7	07	.5M1	EFG2000		0.409				61	28	12.5	0.5	1.7	13.9
7	07	.5M1	EFG2100		0.329				57	22	11.9	0.5	1.9	12.7
7	07	.5M1	EFG2200		0.235				45	19	10.6	0.5	2.4	12.3
7	07	.5M1	EFG2300		0.205				45	17	9.0	0.5	3.3	10.5
7	07	.5M1	EFG2400		0.179				41	17	7.4	0.5	4.2	8.9
8	07	.5M1	EFG 100		0.155				45	18	5.7	0.6	5.0	7.3
8	07	.5M1	EFG 200		0.140				45	16	4.0	0.6	5.8	5.4
8	07	.5M1	EFG 300		0.125				41	17	2.6	0.6	6.6	3.9
8	07	.5M1	EFG 400		0.115				42	16	1.4	0.6	7.3	2.7
8	07	.5M1	EFG 500		0.115				46	16	1.1	0.5	7.5	2.7
8	07	.5M1	EFG 600		0.115				44	17	0.6	0.4	7.5	1.6
8	07	.5M1	EFG 700		0.115				44	15	0.6	0.3	7.3	2.0
8	07	.5M1	EFG 800		0.120				10	13	0.5	0.3	7.1	2.1
8	07	.5M1	EFG 900		0.130				53	14	0.9	0.3	6.4	2.2
8	07	.5M1	EFG1000		0.145				51	13	0.7	0.3	6.4	1.7
8	07	.5M1	EFG1100		0.162				51	14	0.6	0.3	7.1	2.1
8	07	.5M1	EFG1200		0.182				67	13	3.4	1.1	8.7	4.4
8	07	.5M1	EFG1300		0.210				49	13	1.7	0.8	8.8	3.1
8	07	.5M1	EFG1400		0.229				47	13	0.9	0.5	8.0	2.3
8	07	.5M1	EFG1500		0.324				46	14	5.7	1.0	7.7	6.8
8	07	.5M1	EFG1600		0.414				50	16	9.1	0.8	5.2	11.1
8	07	.5M1	EFG1700		0.471				54	19	12.0	0.7	3.1	13.9
8	07	.5M1	EFG1800		0.484				55	18	12.0	0.7	2.2	14.0
8	07	.5M1	EFG1900		0.459				55	25	12.0	0.7	2.1	13.5
8	07	.5M1	EFG2000		0.409				55	25	12.0	0.9	2.3	13.5
8	07	.5M1	EFG2100		0.329				55	19	10.0	0.8	2.7	12.0
8	07	.5M1	EFG2200		0.235				52	21	10.0	0.8	3.2	11.3
8	07	.5M1	EFG2300		0.205				52	17	8.6	0.8	3.9	9.6
8	07	.5M1	EFG2400		0.179				49	16	7.0	0.8	5.0	8.1
9	07	.5M1	EFG 100		0.155				50	17	5.4	0.9	5.8	6.7
9	07	.5M1	EFG 200		0.140				49	16	3.9	0.9	6.6	5.4
9	07	.5M1	EFG 300		0.125				53	16	2.4	0.9	7.5	3.6
9	07	.5M1	EFG 400		0.115				53	15	1.2	0.8	8.3	2.9
9	07	.5M1	EFG 500		0.115				53	16	0.7	0.6	8.6	2.1
9	07	.5M1	EFG 600		0.115				53	16	0.5	0.4	8.4	1.9
9	07	.5M1	EFG 700		0.115				53	17	0.4	0.4	8.4	2.0
9	07	.5M1	EFG 800		0.120				53	17	0.3	0.3	8.3	1.8
9	07	.5M1	EFG 900		0.130				53	13	3.7	0.5	1.0	4.8
9	07	.5M1	EFG1000		0.145				52	13	0.7	0.3	7.7	4.5
9	07	.5M1	EFG1100		0.162				49	13	0.6	0.3	7.7	5.0

BIO-SURF PILOT PLANT TESTING
1976

ANALYTICAL RESULTS - SERIES B2 (HOURLY)

Y	MON	PILOT UNIT	SAMPLE IDENTITY	FLOW RATE IGPM	UNFILTERED SAMPLE			FILTERED SAMPLE					
					BOD	COD	TKN	BOD	COD	TOC	NH3N	NO2N	NO3N
3	08	.5M1	RFG09	0.42				119	31	12.0	0.3	0.2	14.1
3	08	.5M1	RFG10	0.45				46	13	12.0	0.2	0.0	13.8
3	08	.5M1	RFG11	0.50				51	14	15.0	0.2	0.0	16.2
3	08	.5M1	RFG12	0.58				59	17	19.0	0.1	0.0	21.3
3	08	.5M1	RFG13	0.675				118	38	31.0	0.2	0.1	33.0
3	08	.5M1	RFG14	0.754				158	53	36.0	0.1	0.0	37.3
3	08	.5M1	RFG15	1.025				184	64	41.0	0.0	0.0	42.8
3	08	.5M1	RFG16	1.29				168	59	31.0	0.0	0.0	33.1
3	08	.5M1	RFG17	1.44				150	50	26.0	0.0	0.0	27.9
3	08	.5M1	RFG18	1.50				128	31	22.0	0.0	0.0	23.0
3	08	.5M1	RFG19	1.45				168	60	28.0	0.0	0.0	29.7
3	08	.5M1	RFG20	1.41				160	52	26.0	0.1	0.1	27.6
3	08	.5M1	RFG21	1.1				158	51	26.0	0.1	0.1	27.7
3	08	.5M1	RFG22	0.75				132	37	22.0	0.1	0.1	23.9
3	08	.5M1	RFG23	0.625				98	27	15.0	0.1	0.0	17.2
3	08	.5M1	RFG24	0.555				92	24	14.0	0.0	0.1	15.1
4	08	.5M1	RFG01	0.49				84	32	14.0	0.1	0.1	15.2
4	08	.5M1	RFG02	0.455				96	27	14.0	0.1	0.0	15.4
4	08	.5M1	RFG03	0.42				80	22	14.0	0.1	0.0	15.0
4	08	.5M1	RFG04	0.405				70	18	14.0	0.1	0.1	15.3
4	08	.5M1	RFG05	0.40				59	16	14.0	0.1	0.3	15.2
4	08	.5M1	RFG06	0.40				53	15	13.0	0.1	0.4	14.9
4	08	.5M1	RFG07	0.40				50	14	12.0	0.1	0.4	13.2
4	08	.5M1	RFG08	0.40				50	14	12.0	0.1	0.5	13.0
4	08	.5M1	RFG09	0.42				45	14	11.0	0.2	0.6	12.9
4	08	.5M1	RFG10	0.45				66	18	12.0	0.2	0.5	13.8
4	08	.5M1	RFG11	0.50				82	24	17.0	0.7	0.0	19.2
4	08	.5M1	RFG12	0.58				89	22	19.0	0.5	0.1	22.0
4	08	.5M1	RFG13	0.675				148	37	27.0	0.0	0.0	29.1
4	08	.5M1	RFG14	0.754				224	67	37.0	0.0	0.0	41.0
4	08	.5M1	RFG15	1.025				164	46	32.0	0.0	0.0	33.1
4	08	.5M1	RFG16	1.29				206	57	31.0	0.0	0.0	32.1
4	08	.5M1	RFG17	1.44				182	53	33.0	0.0	0.0	33.1
4	08	.5M1	RFG18	1.50				130	35	22.0	0.0	0.0	23.1
4	08	.5M1	RFG19	1.45				160	55	28.0	0.0	0.0	29.1
4	08	.5M1	RFG20	1.41				122	35	19.0	0.4	0.0	19.1
4	08	.5M1	RFG21	1.1				114	31	15.0	0.5	1.4	16.1
4	08	.5M1	RFG22	0.75				109	31	14.0	0.3	0.1	15.1
4	08	.5M1	RFG23	0.625				100	29	14.0	0.2	0.1	15.1
4	08	.5M1	RFG24	0.555				100	48	13.0	0.2	0.0	14.1
5	08	.5M1	RFG01	0.49				89	30	14.0	0.1	0.0	15.1
5	08	.5M1	RFG02	0.455				88	24	14.0	0.1	0.2	15.1
5	08	.5M1	RFG03	0.42				81	21	15.0	0.1	0.3	15.1
5	08	.5M1	RFG04	0.405				74	21	14.0	0.2	0.4	15.1
5	08	.5M1	RFG05	0.40				69	23	14.0	0.2	0.5	15.1
5	08	.5M1	RFG06	0.40				65	20	13.0	0.2	0.5	14.1
5	08	.5M1	RFG07	0.40				55	17	13.0	0.1	0.8	13.1
5	08	.5M1	RFG08	0.40				54	20	11.0	0.2	0.9	12.1
5	08	.5M1	RFG09	0.42				52	17	14.0	0.6	0.8	15.1
5	08	.5M1	RFG10	0.45				54	15	11.0	0.3	0.7	12.1
5	08	.5M1	RFG11	0.50				56	16	15.0	0.2	0.4	15.1
5	08	.5M1	RFG12	0.58				71	21	19.0	0.2	0.2	19.1
5	08	.5M1	RFG13	0.675				96	25	25.0	0.5	0.2	26.1

ANALYTICAL RESULTS - SERIES B2 (HOURLY)

MON	PILOT	SAMPLE	FLOW RATE IGPM	UNFILTERED		FILTERED						
				BOD	COD TKN SAMPLE	BOD	COD	TOC	NH3N	NO2N	NO3N	TKN
	UNIT	IDENTITY										
08	.5M1	RFG14	0.754			131	37	25.0	0.0	0.0	26.0	
08	.5M1	RFG15	1.025			133	37	27.0	0.0	0.0	27.7	
08	.5M1	RFG16	1.29			194	57	29.0	0.0	0.0	29.5	
08	.5M1	RFG17	1.44			146	42	24.0	0.1	0.1	25.5	
08	.5M1	RFG18	1.50			135	34	20.0	0.1	0.0	21.6	
08	.5M1	RFG19	1.45			160	50	28.0	0.0	0.0	29.5	
08	.5M1	RFG20	1.41			139	40	23.0	0.2	0.0	24.0	
08	.5M1	RFG21	1.1			88	27	18.0	0.6	0.4	19.3	
08	.5M1	RFG22	0.75			127	39	19.0	0.3	0.1	19.3	
08	.5M1	RFG23	0.625			117	27	18.0	0.3	0.1	19.1	
08	.5M1	RFG24	0.555			78	33	15.0	0.4	0.2	15.5	
08	.5M1	RFG01	0.49			74	20	14.0	0.4	0.4	15.3	
08	.5M1	RFG02	0.455			79	21	16.0	0.3	0.3	16.7	
08	.5M1	RFG03	0.42			71	20	16.0	0.2	0.2	17.3	
08	.5M1	RFG04	0.405			63	19	16.0	0.1	0.1	17.2	
08	.5M1	RFG05	0.40			61	19	16.0	0.1	0.2	16.9	
08	.5M1	RFG06	0.40			55	16	15.0	0.1	0.3	16.1	
08	.5M1	RFG07	0.40			55	15	14.0	0.2	0.4	15.0	
08	.5M1	RFG08	0.40			56	14	13.0	0.2	0.5	14.4	
08	.5M1	EFG09	0.42			83	23	5.0	2.7	0.6	7.1	
08	.5M1	EFG10	0.45			46	14	5.0	3.3	0.6	6.8	
08	.5M1	EFG11	0.50			43	14	6.0	3.7	0.8	7.8	
08	.5M1	EFG12	0.58			41	13	9.0	3.3	0.7	10.3	
08	.5M1	EFG13	0.675			48	15	15.0	1.9	0.3	16.9	
08	.5M1	EFG14	0.754			65	21	22.0	0.4	0.2	23.2	
08	.5M1	EFG15	1.025			89	27	26.0	0.0	0.0	27.8	
08	.5M1	EFG16	1.29			115	38	27.0	0.0	0.0	28.9	
08	.5M1	EFG17	1.44			115	37	24.0	0.0	0.0	26.1	
08	.5M1	EFG18	1.50			90	25	18.0	0.0	0.0	20.0	
08	.5M1	EFG19	1.45			116	38	22.0	0.0	0.0	23.7	
08	.5M1	EFG20	1.41			111	33	22.0	0.0	0.0	23.9	
08	.5M1	EFG21	1.1			86	24	17.0	0.0	0.0	19.1	
08	.5M1	EFG22	0.75			85	25	16.0	0.0	0.0	17.6	
08	.5M1	EFG23	0.625			73	21	14.0	0.0	0.0	14.9	
08	.5M1	EFG24	0.555			65	21	13.0	0.0	0.2	14.7	
08	.5M1	EFG01	0.49			57	20	12.0	0.2	0.2	13.7	
08	.5M1	EFG02	0.455			60	18	11.0	0.4	0.2	12.9	
08	.5M1	EFG03	0.42			59	19	11.0	0.5	0.2	12.8	
08	.5M1	EFG04	0.405			55	17	11.0	0.8	0.3	12.8	
08	.5M1	EFG05	0.40			54	16	10.0	0.8	0.3	12.1	
08	.5M1	EFG06	0.40			49	17	10.0	0.9	0.4	11.9	
08	.5M1	EFG07	0.40			50	16	9.0	1.1	0.4	10.9	
08	.5M1	EFG08	0.40			46	16	9.0	1.4	0.5	11.0	
08	.5M1	EFG09	0.42			47	14	9.0	1.1	0.5	10.6	
08	.5M1	EFG10	0.45			45	13	9.0	1.0	0.6	10.5	
08	.5M1	EFG11	0.50			44	13	9.0	1.2	0.4	10.9	
08	.5M1	EFG12	0.58			49	16	12.0	0.9	0.3	13.5	
08	.5M1	EFG13	0.675			50	16	16.0	0.9	0.3	18.0	
08	.5M1	EFG14	0.754			75	20	20.0	0.2	0.1	22.2	
08	.5M1	EFG15	1.025			102	27	23.0	0.0	0.0	24.9	
08	.5M1	EFG16	1.29			141	39	25.0	0.0	0.0	25.9	
08	.5M1	EFG17	1.44			166	43	29.0	0.0	0.0	31.1	
08	.5M1	EFG18	1.50			119	32	21.0	0.0	0.0	22.7	

Y	MON	PILOT	SAMPLE	FLOW RATE IGPM	ANALYTICAL RESULTS - SERIES B2 (HOURLY)									
					UNFILTERED SAMPLE			FILTERED SAMPLE						
		UNIT	IDENTITY		BOD	COD	TKN	BOD	COD	TOC	NH3N	NO2N	NO3N	TKN
4	08	.5M1	EFG19	1.45				96	27	21.0	0.0	0.0	22.4	
4	08	.5M1	EFG20	1.41				88	22	19.0	0.0	0.0	19.9	
4	08	.5M1	EFG21	1.1				92	28	15.0	0.0	0.0	16.6	
4	08	.5M1	EFG22	0.75				75	20	15.0	0.0	0.0	16.5	
4	08	.5M1	EFG23	0.625				64	20	15.0	0.0	0.0	16.2	
4	08	.5M1	EFG24	0.555				60	19	14.0	0.2	0.1	15.8	
5	08	.5M1	EFG01	0.49				57	19	13.0	0.2	0.1	14.5	
5	08	.5M1	EFG02	0.455				57	17	13.0	0.3	0.0	14.2	
5	08	.5M1	EFG03	0.42				58	19	13.0	0.5	0.0	13.9	
5	08	.5M1	EFG04	0.405				55	17	13.0	0.6	0.1	14.2	
5	08	.5M1	EFG05	0.40				52	16	13.0	0.7	0.1	14.3	
5	08	.5M1	EFG06	0.40				52	16	13.0	0.8	0.1	14.1	
5	08	.5M1	EFG07	0.40				50	19	12.0	0.9	0.1	13.8	
5	08	.5M1	EFG08	0.40				50	16	11.0	1.0	0.2	12.9	
5	08	.5M1	EFG09	0.42				43	13	11.0	0.1	0.2	12.3	
5	08	.5M1	EFG10	0.45				44	13	11.0	0.6	0.4	11.9	
5	08	.5M1	EFG11	0.50				45	14	11.0	0.7	0.3	12.6	
5	08	.5M1	EFG12	0.58				46	13	13.0	0.8	0.3	14.9	
5	08	.5M1	EFG13	0.675				47	14	17.0	0.7	0.3	20.0	
5	08	.5M1	EFG14	0.754				58	15	21.0	0.5	0.3	23.3	
5	08	.5M1	EFG15	1.025				60	18	21.0	0.3	0.2	22.7	
5	08	.5M1	EFG16	1.29				90	21	24.0	0.0	0.0	25.7	
5	08	.5M1	EFG17	1.44				92	22	23.0	0.0	0.0	24.1	
5	08	.5M1	EFG18	1.50				97	22	20.0	0.0	0.0	21.0	
5	08	.5M1	EFG19	1.45				109	24	24.0	0.0	0.0	25.7	
5	08	.5M1	EFG20	1.41				103	31	23.0	0.0	0.0	23.7	
5	08	.5M1	EFG21	1.1				68	19	17.0	0.0	0.2	18.1	
5	08	.5M1	EFG22	0.75				77	20	17.0	0.3	0.1	18.5	
5	08	.5M1	EFG23	0.625				82	19	16.0	0.0	0.2	17.6	
5	08	.5M1	EFG24	0.555				64	17	15.0	0.0	0.3	15.6	
6	08	.5M1	EFG01	0.49				54	16	14.0	0.1	0.3	14.7	
6	08	.5M1	EFG02	0.455				51	15	14.0	0.2	0.3	14.9	
6	08	.5M1	EFG03	0.42				51	14	14.0	0.3	0.2	15.7	
6	08	.5M1	EFG04	0.405				52	15	15.0	0.3	0.3	16.0	
6	08	.5M1	EFG05	0.40				54	15	15.0	0.4	0.3	15.8	
6	08	.5M1	EFG06	0.40				52	15	14.0	0.4	0.4	15.7	
6	08	.5M1	EFG07	0.40				50	14	14.0	0.6	0.3	14.7	
6	08	.5M1	EFG08	0.40				50	14	13.0	0.7	0.4	13.9	

BIO-SURF PILOT PLANT TESTING
 1976

ANALYTICAL RESULTS - SERIES C2 + E2 (30 MIN)

MON	PILOT SAMPLE UNIT IDENTITY	FLOW RATE IGPM	UNFILTERED SAMPLE			FILTERED SAMPLE					
			BOD	COO	TKN	BOD	COO	TOC	NH3N	NO2N	NO3N
09	.5M1+2MRFG1000			222	69	22.0	0.0	0.0	23.3		
09	.5M1+2MRFG1030			250	74	21.0	0.0	0.0	22.0		
09	.5M1+2MRFG1100			231	74	22.0	0.0	0.0	22.8		
09	.5M1+2MRFG1130			277	100	20.0	0.0	0.0	21.7		
09	.5M1+2MRFG1200			212	65	18.0	0.0	0.0	19.5		
09	.5M1+2MRFG1230			272	80	16.0	0.0	0.0	17.8		
09	.5M1+2MRFG1300			328	117	10.0	0.0	0.0	11.1		
09	.5M1+2MRFG1330			340	113	16.0	0.0	0.0	17.2		
09	.5M1+2MRFG1400			300	99	12.0	0.0	0.0	13.1		
09	.5M1+2MRFG1430			363	145	17.0	0.0	0.0	18.0		
09	.5M1+2MRFG1500			175	78	17.0	0.0	0.0	17.9		
09	.5M1+2MRFG1530			131	58	9.0	0.0	0.3	11.1		
09	.5M1+2MRFG1600			144	55	7.0	0.0	0.0	7.9		
09	.5M1+2MRFG1630			86	34	6.0	0.0	0.0	7.1		
09	.5M1+2MRFG1700			111	41	7.0	0.0	0.0	7.4		
09	.5M1+2MRFG1730			87	36	4.0	0.0	0.0	5.8		
09	.5M1+2MRFG1800			129	51	13.0	0.0	0.0	14.4		
09	.5M1+2MRFG1830			184	57	12.0	0.0	0.0	12.9		
09	.5M1+2MRFG1900			144	58	16.0	0.0	0.0	17.6		
09	.5M1+2MRFG1930			186	53	13.0	0.0	0.0	13.9		
09	.5M1+2MRFG2000			184	63	14.0	0.0	0.0	15.5		
09	.5M1+2MRFG2030			182	56	9.0	0.0	0.0	10.6		
09	.5M1+2MRFG2100			148	45	5.0	0.0	0.0	6.6		
09	.5M1+2MRFG2130			132	47	7.0	0.0	0.0	7.7		
09	.5M1+2MRFG2200			117	42	8.0	0.0	0.0	9.0		
09	.5M1+2MRFG2230			152	51	9.0	0.0	0.0	9.9		
09	.5M1+2MRFG2300			192	53	15.0	0.0	0.0	16.3		
09	.5M1+2MRFG2330			151	48	12.0	0.0	0.0	13.1		
09	.5M1+2MRFG2400			118	42	9.0	0.0	0.0	10.7		
09	.5M1+2MRFG0030			140	51	9.0	0.0	0.0	11.2		
09	.5M1+2MRFG0100			342	103	12.0	0.0	0.0	13.5		
09	.5M1+2MRFG0130			320	98	9.0	0.0	0.0	11.2		
09	.5M1+2MRFG0200			312	95	17.0	0.0	0.0	18.3		
09	.5M1+2MRFG0230			230	73	11.0	0.0	0.0	12.7		
09	.5M1+2MRFG0300			230	70	21.0	0.0	0.0	23.0		
09	.5M1+2MRFG0330			235	72	12.0	0.0	0.0	13.7		
09	.5M1+2MRFG0400			246	71	15.0	0.0	0.2	16.9		
09	.5M1+2MRFG0430			242	70	16.0	0.0	0.2	16.9		
09	.5M1+2MRFG0500			207	59	16.0	0.0	0.1	17.2		
09	.5M1+2MRFG0530			201	62	15.0	0.0	0.0	16.1		
09	.5M1+2MRFG0600			131	39	14.0	0.0	0.0	15.1		
09	.5M1+2MRFG0630			141	41	14.0	0.0	0.0	14.6		
09	.5M1+2MRFG0700			139	41	12.0	0.0	0.0	13.3		
09	.5M1+2MRFG0730			137	39	13.0	0.0	0.0	13.7		
09	.5M1+2MRFG0800			133	37	13.0	0.0	0.1	14.1		
09	.5M1+2MRFG0830			135	39	13.0	0.0	0.3	13.9		
09	.5M1+2MRFG0900			140	41	13.0	0.1	0.3	13.9		
09	.5M1+2MRFG0930			119	38	14.0	0.0	0.1	14.8		
09	.5M1+2MRFG1000			102	42	18.0	0.0	0.2	20.7		
09	.5M1+2MRFG1030			125	42	19.0	0.0	0.1	23.7		
09	.5M1+2MRFG1100			37	14	10.0	0.0	0.1	13.8		
09	.5M1+2MRFG1130			39	15	11.0	0.0	0.2	13.0		
09	.5M1+2MRFG1200			52	25	11.0	0.0	0.1	13.5		

ANALYTICAL RESULTS - SERIES C2 + E2 (30 MIN)

Y	MON	PILOT SAMPLE UNIT IDENTITY	FLOW RATE IGPM	UNFILTERED SAMPLE			FILTERED SAMPLE					
				BOD	COD	TKN	BOD	COD	TOC	NH3N	NO2N	NO3N
3	09	.5M1+2MRFG1230					81	27	11.0	0.0	0.1	13.5
3	09	.5M1+2MRFG1300					51	23	10.0	0.0	0.0	12.6
3	09	.5M1+2MRFG1330					92	34	12.0	0.0	0.0	13.9
3	09	.5M1+2MRFG1400					89	31	13.0	0.0	0.0	13.4
3	09	.5M1+2MRFG1430					86	31	11.0	0.0	0.0	12.8
3	09	.5M1+2MRFG1500					79	30	12.0	0.0	0.7	13.1
3	09	.5M1+2MRFG1530					104	39	13.0	0.2	0.6	14.3
3	09	.5M1+2MRFG1600					185	62	19.0	0.5	0.6	20.5
3	09	.5M1+2MRFG1630					200	69	22.0	0.0	0.1	23.8
3	09	.5M1+2MRFG1700					138	40	16.0	0.0	0.1	17.1
3	09	.5M1+2MRFG1730					125	42	16.0	0.0	0.0	16.3
3	09	.5M1+2MRFG1800					138	52	19.0	0.0	0.1	21.5
3	09	.5M1+2MRFG1830					138	52	19.0	0.3	4.8	20.1
3	09	.5M1+2MRFG1900					158	53	19.0	0.1	2.3	20.6
3	09	.5M1+2MRFG1930					155	51	18.0	0.1	0.5	20.1
3	09	.5M1+2MRFG2000					134	48	14.0	0.0	0.2	16.2
3	09	.5M1+2MRFG2030					145	46	16.0	0.0	0.4	17.7
3	09	.5M1+2MRFG2100					438	123	28.0	0.0	0.1	29.0
3	09	.5M1+2MRFG2130					278	115	18.0	0.0	0.0	22.0
3	09	.5M1+2MRFG2200					306	87	19.0	0.0	0.0	21.4
3	09	.5M1+2MRFG2230					322	83	19.0	0.0	0.0	21.7
3	09	.5M1+2MRFG2300					263	83	18.0	0.0	0.0	19.9
3	09	.5M1+2MRFG2330					290	82	19.0	0.0	0.0	20.2
3	09	.5M1+2MRFG2400					288	79	18.0	0.0	0.1	19.2
3	09	.5M1+2MRFG0030					298	84	18.0	0.0	0.1	19.2
3	09	.5M1+2MRFG0100					305	86	18.0	0.0	0.0	19.8
3	09	.5M1+2MRFG0130					296	82	18.0	0.0	0.0	19.2
3	09	.5M1+2MRFG0200					280	79	18.0	0.0	0.0	18.8
3	09	.5M1+2MRFG0230					298	84	18.0	0.0	0.0	19.4
3	09	.5M1+2MRFG0300					300	83	19.0	0.0	0.0	20.8
3	09	.5M1+2MRFG0330					268	75	19.0	0.0	0.0	20.5
3	09	.5M1+2MRFG0400					230	70	18.0	0.0	0.0	20.4
3	09	.5M1+2MRFG0430					230	71	18.0	0.0	0.0	19.5
3	09	.5M1+2MRFG0500					237	67	18.0	0.0	0.0	19.5
3	09	.5M1+2MRFG0530					224	70	18.0	0.0	0.0	19.2
3	09	.5M1+2MRFG0600					230	71	17.0	0.0	0.0	18.1
3	09	.5M1+2MRFG0630					236	70	17.0	0.0	0.0	17.7
3	09	.5M1+2MRFG0700					252	73	17.0	0.0	0.0	18.2
3	09	.5M1+2MRFG0730					263	78	17.0	0.0	0.4	18.1
3	09	.5M1+2MRFG0800					263	75	16.0	0.0	0.2	17.0
3	09	.5M1+2MRFG0830					253	74	16.0	0.0	0.4	17.6
3	09	.5M1+2MRFG0900					253	75	16.0	0.0	0.2	16.9
3	09	.5M1+2MRFG0930					280	81	18.0	0.1	0.4	19.3
3	09	.5M1+2MRFG1000					276	81	21.0	0.0	0.1	22.2
3	09	.5M1+2MRFG1030					242	74	18.0	0.0	0.0	23.0
3	09	.5M1+2MRFG1100					280	85	25.0	0.0	0.0	28.4
3	09	.5M1+2MRFG1130					310	89	24.0	0.0	0.0	26.1
3	09	.5M1+2MRFG1200					255	73	10.0	0.0	0.0	13.5
3	09	.5M1+2MRFG1230					270	82	12.0	0.0	0.1	13.8
3	09	.5M1+2MRFG1300					253	71	11.0	0.0	0.0	12.2
3	09	.5M1+2MRFG1330					292	84	12.0	0.0	0.1	14.1
3	09	.5M1+2MRFG1400					276	75	9.0	0.1	0.1	11.6
3	09	.5M1+2MRFG1430					200	63	12.0	0.0	0.0	14.1
3	09	.5M1+2MRFG1500					312	77	10.0	0.0	0.1	12.3
3	09	.5M1+2MRFG1530					296	80	11.0	0.0	0.1	11.6

ANALYTICAL RESULTS - SERIES C2 + E2 (30 MIN)

Y	MON	PILOT SAMPLE UNIT IDENTITY	FLOW RATE IGPM	UNFILTERED SAMPLE			FILTERED SAMPLE					
				BOD	COU	TKN	BOD	COU	TOC	NH3N	NO2N	NO3N
9	09	.5M1+2MRFG1600					282	83	9.0	0.0	0.0	11.5
9	09	.5M1+2MRFG1630					296	79	10.0	0.0	0.3	12.0
9	09	.5M1+2MRFG1700					170	45	8.0	0.0	0.2	10.1
9	09	.5M1+2MRFG1730					148	44	7.0	0.1	0.4	9.4
9	09	.5M1+2MRFG1800					136	41	7.0	0.1	0.3	9.2
9	09	.5M1+2MRFG1830					148	44	7.0	0.1	0.6	9.5
9	09	.5M1+2MRFG1900					180	58	11.0	0.1	0.5	12.1
9	09	.5M1+2MRFG1930					168	52	10.0	0.1	0.4	10.2
9	09	.5M1+2MRFG2000					192	61	11.0	0.1	0.3	11.5
9	09	.5M1+2MRFG2030					190	61	12.0	0.1	0.3	12.1
9	09	.5M1+2MRFG2100					204	58	12.0	0.1	0.2	12.6
9	09	.5M1+2MRFG2130					210	60	11.0	0.1	0.2	11.2
9	09	.5M1+2MRFG2200					296	99	19.0	0.0	0.1	22.2
9	09	.5M1+2MRFG2230					290	84	16.0	0.0	0.0	17.7
9	09	.5M1+2MRFG2300					282	86	15.0	0.0	0.0	16.5
9	09	.5M1+2MRFG2330					302	90	18.0	0.0	0.0	18.2
9	09	.5M1+2MRFG2400					272	81	15.0	0.0	0.0	17.7
0	09	.5M1+2MRFG0030					276	80	15.0	0.0	0.0	17.7
0	09	.5M1+2MRFG0100					282	79	15.0	0.0	0.0	17.7
0	09	.5M1+2MRFG0130					270	84	15.0	0.0	0.0	16.5
0	09	.5M1+2MRFG0200					254	75	16.0	0.0	0.0	17.1
0	09	.5M1+2MRFG0230					280	81	16.0	0.0	0.0	16.9
0	09	.5M1+2MRFG0300					270	82	16.0	0.0	0.0	16.3
0	09	.5M1+2MRFG0330					286	80	15.0	0.0	0.0	15.8
0	09	.5M1+2MRFG0400					448	136	13.0	0.0	0.4	15.0
0	09	.5M1+2MRFG0430					362	78	13.0	0.0	0.6	14.8
0	09	.5M1+2MRFG0500					282	77	12.0	0.1	0.5	14.8
0	09	.5M1+2MRFG0530					276	77	13.0	0.1	0.6	14.8
0	09	.5M1+2MRFG0600					268	72	12.0	0.1	0.6	14.1
0	09	.5M1+2MRFG0630					276	76	13.0	0.0	0.3	13.9
0	09	.5M1+2MRFG0700					184	36	13.0	0.0	0.4	14.8
0	09	.5M1+2MRFG0730					236	71	12.0	0.0	0.3	12.0
0	09	.5M1+2MRFG0800					252	74	11.0	0.0	0.4	11.0

BIO-SURF PILOT PLANT TESTING
1976

ANALYTICAL RESULTS - SERIES C2 (30 MIN)

DAY	MON	PILOT UNIT	SAMPLE IDENTITY	FLOW RATE IGPM	UNFILTERED SAMPLE			FILTERED SAMPLE					
					BOD	COD	TKN	BOD	COD	TOC	NH3N	NO2N	NO3N
7	09	.5M1	EFG1000	0.508				54	17	9.0	0.2	0.3	9.0
7	09	.5M1	EFG1030	0.508				55	17	9.0	0.2	0.3	10.1
7	09	.5M1	EFG1100	0.508				59	19	16.0	0.4	0.2	16.6
7	09	.5M1	EFG1130	0.508				66	20	10.0	0.1	0.1	10.7
7	09	.5M1	EFG1200	0.508				64	21	8.0	0.0	0.1	9.5
7	09	.5M1	EFG1230	0.508				70	23	20.0	0.1	0.3	22.6
7	09	.5M1	EFG1300	0.508				66	24	20.0	0.0	0.3	21.7
7	09	.5M1	EFG1330	0.508				80	26	17.0	0.0	0.2	18.6
7	09	.5M1	EFG1400	0.508				93	31	17.0	0.0	0.1	17.0
7	09	.5M1	EFG1430	0.508				92	32	17.0	0.0	0.2	18.5
7	09	.5M1	EFG1500	1.01				105	36	15.0	0.0	0.1	16.3
7	09	.5M1	EFG1530	1.01				95	35	9.0	0.0	0.2	10.0
7	09	.5M1	EFG1600	1.01				110	23	14.0	0.0	0.2	15.6
7	09	.5M1	EFG1630	1.01				70	30	18.0	0.0	0.3	19.3
7	09	.5M1	EFG1700	1.01				69	23	9.0	0.0	0.2	11.0
7	09	.5M1	EFG1730	1.01				63	21	12.0	0.0	0.2	13.0
7	09	.5M1	EFG1800	1.01				55	19	11.0	0.2	0.3	11.0
7	09	.5M1	EFG1830	1.01				72	23	10.0	0.1	0.3	10.0
7	09	.5M1	EFG1900	1.01				76	30	13.0	0.3	0.2	14.0
7	09	.5M1	EFG1930	1.01				56	30	14.0	0.2	0.3	14.0
7	09	.5M1	EFG2000	0.507				43	24	12.0	0.2	0.3	14.0
7	09	.5M1	EFG2030	0.507				47	22	11.0	0.2	0.3	11.0
7	09	.5M1	EFG2100	0.507				49	24	10.0	0.2	0.3	11.0
7	09	.5M1	EFG2130	0.507				62	22	7.0	0.1	0.2	9.0
7	09	.5M1	EFG2200	0.507				63	21	5.0	0.0	0.2	7.0
7	09	.5M1	EFG2230	0.507				63	21	5.0	0.2	0.2	6.0
7	09	.5M1	EFG2300	0.507				63	20	4.0	0.1	0.1	5.0
7	09	.5M1	EFG2330	0.507				58	20	5.0	0.1	2.0	6.0
7	09	.5M1	EFG2400	0.507				56	20	5.0	0.1	0.2	6.0
8	09	.5M1	EFG0030	0.507				55	19	9.0	0.2	0.2	9.0
8	09	.5M1	EFG0100	0.511				53	19	9.0	0.2	0.3	11.0
8	09	.5M1	EFG0130	0.511				57	21	7.0	0.2	0.2	8.0
8	09	.5M1	EFG0200	0.511				60	21	7.0	0.1	0.2	7.0
8	09	.5M1	EFG0230	0.511				59	21	6.0	0.1	0.1	7.0
8	09	.5M1	EFG0300	0.511				57	21	7.0	0.1	0.1	7.0
8	09	.5M1	EFG0330	0.511				58	20	10.0	0.1	0.3	11.0
8	09	.5M1	EFG0400	0.511				67	20	9.0	0.1	0.1	10.0
8	09	.5M1	EFG0430	0.511				68	20	10.0	0.1	0.2	10.0
8	09	.5M1	EFG0500	0.511				67	20	11.0	0.2	0.2	12.0
8	09	.5M1	EFG0530	0.511				63	19	11.0	0.3	0.1	11.0
8	09	.5M1	EFG0600	0.991				61	19	11.0	0.1	0.2	11.0
8	09	.5M1	EFG0630	0.991				57	18	12.0	0.1	0.1	12.0
8	09	.5M1	EFG0700	0.991				59	17	12.0	0.1	0.1	13.0
8	09	.5M1	EFG0730	0.991				58	17	11.0	0.1	0.1	12.0
8	09	.5M1	EFG0800	0.991				56	17	11.0	0.1	0.1	12.0
8	09	.5M1	EFG0830	0.991				55	16	11.0	0.2	0.1	12.0
8	09	.5M1	EFG0900	0.991				54	17	11.0	0.1	0.1	12.0
8	09	.5M1	EFG0930	0.991				56	17	11.0	0.1	0.2	12.0
8	09	.5M1	EFG1000	0.991				40	15	13.0	0.3	0.4	13.0
8	09	.5M1	EFG1030	0.991				40	14	15.0	0.1	0.2	16.0
8	09	.5M1	EFG1100	0.509				51	16	16.0	0.0	0.2	17.0
8	09	.5M1	EFG1130	0.509				48	14	16.0	0.0	0.3	16.0
8	09	.5M1	EFG1200	0.509				44	13	11.0	0.1	0.3	14.0

ANALYTICAL RESULTS - SERIES C2 (30 MIN)													
Y	MON	PILOT UNIT	SAMPLE IDENTITY	FLOW RATE IGPM	UNFILTERED SAMPLE			FILTERED SAMPLE					
					BOD	COD	TKN	BOD	COD	TOC	NH3N	NO2N	NO3N
8	09	.5M1	EFG1230	0.509				44	12	9.0	3.1	0.4	11.3
8	09	.5M1	EFG1300	0.509				43	12	10.0	0.2	0.3	12.4
8	09	.5M1	EFG1330	0.509				42	11	10.0	0.2	0.4	11.0
8	09	.5M1	EFG1400	0.509				73	18	15.0	0.0	0.3	16.3
8	09	.5M1	EFG1430	0.509				61	16	14.0	0.0	0.2	15.7
8	09	.5M1	EFG1500	0.509				56	15	13.0	0.0	0.5	13.7
8	09	.5M1	EFG1530	0.509				42	15	11.0	0.1	0.5	12.5
8	09	.5M1	EFG1600	1.00				342	96	9.0	0.1	0.3	13.6
8	09	.5M1	EFG1630	1.00				117	34	12.0	0.0	0.1	13.5
8	09	.5M1	EFG1700	1.00				94	32	13.0	0.0	0.3	14.4
8	09	.5M1	EFG1730	1.00				69	20	11.0	0.1	0.2	13.3
8	09	.5M1	EFG1800	1.00				93	21	11.0	0.1	0.3	12.9
8	09	.5M1	EFG1830	1.00				76	23	13.0	0.1	0.4	13.5
8	09	.5M1	EFG1900	1.00				74	23	13.0	0.4	1.6	13.7
8	09	.5M1	EFG1930	1.00				76	22	13.0	0.4	0.9	14.3
8	09	.5M1	EFG2000	1.00				81	21	13.0	0.1	0.4	14.2
8	09	.5M1	EFG2030	1.00				70	19	12.0	0.1	0.3	13.0
8	09	.5M1	EFG2100	0.991				87	23	12.0	0.0	0.4	13.4
8	09	.5M1	EFG2130	0.991				132	49	19.0	0.1	0.2	22.7
8	09	.5M1	EFG2200	0.991				134	47	20.0	0.2	0.1	22.4
8	09	.5M1	EFG2230	0.991				130	40	19.0	0.0	0.1	21.1
8	09	.5M1	EFG2300	0.991				115	38	18.0	0.0	0.0	20.9
8	09	.5M1	EFG2330	0.991				110	36	18.0	0.0	0.0	19.5
8	09	.5M1	EFG2400	0.991				103	34	16.0	0.0	0.0	17.9
9	09	.5M1	EFG0300	0.991				96	34	18.0	0.0	0.0	19.7
9	09	.5M1	EFG0100	0.991				99	33	18.0	0.0	0.0	19.6
9	09	.5M1	EFG0130	0.991				98	35	18.0	0.0	0.0	19.6
9	09	.5M1	EFG0200	0.509				103	34	18.0	0.0	0.0	19.3
9	09	.5M1	EFG0230	0.509				80	27	18.0	0.0	0.0	18.7
9	09	.5M1	EFG0300	0.509				78	25	18.0	0.0	0.0	18.6
9	09	.5M1	EFG0330	0.509				75	22	17.0	0.0	0.0	17.9
9	09	.5M1	EFG0400	0.509				71	22	18.0	0.0	0.0	19.1
9	09	.5M1	EFG0430	0.509				68	22	14.0	0.0	0.1	15.6
9	09	.5M1	EFG0500	0.509				63	20	15.0	0.0	0.1	15.9
9	09	.5M1	EFG0530	0.509				69	23	15.0	0.0	0.1	15.9
9	09	.5M1	EFG0600	0.509				64	21	15.0	0.0	0.2	16.2
9	09	.5M1	EFG0630	0.509				63	20	14.0	0.0	0.1	15.8
9	09	.5M1	EFG0700	1.01				62	19	14.0	0.0	0.1	15.1
9	09	.5M1	EFG0730	1.01				41	19	11.0	0.0	0.1	12.7
9	09	.5M1	EFG0800	1.01				63	21	11.0	0.0	0.1	12.1
9	09	.5M1	EFG0830	1.01				64	21	11.0	0.0	0.1	12.5
9	09	.5M1	EFG0900	1.01				59	20	10.0	0.0	0.1	11.1
9	09	.5M1	EFG0930	1.01				58	20	10.0	0.0	0.1	11.6
9	09	.5M1	EFG1000	1.01				63	23	12.0	0.0	0.1	13.5
9	09	.5M1	EFG1030	1.01				77	22	14.0	0.0	0.0	15.3
9	09	.5M1	EFG1100	1.01				74	22	17.0	0.0	0.0	18.3
9	09	.5M1	EFG1130	1.01				92	29	21.0	0.0	0.0	21.6
9	09	.5M1	EFG1200	1.01				109	35	20.0	0.0	0.0	22.2
9	09	.5M1	EFG1230	1.01				103	30	11.0	0.0	0.1	14.6
9	09	.5M1	EFG1300	1.01				54	58	7.0	0.0	0.0	10.3
9	09	.5M1	EFG1330	1.01				67	20	5.0	0.0	0.0	6.6
9	09	.5M1	EFG1400	1.01				70	20	4.0	0.0	0.0	5.0
9	09	.5M1	EFG1430	1.01				72	21	4.0	0.0	0.0	6.3
9	09	.5M1	EFG1500	1.01				74	23	4.0	0.0	0.0	7.3
9	09	.5M1	EFG1530	1.01				64	22	4.0	0.0	0.0	6.0

ANALYTICAL RESULTS - SERIES C2 (30 MIN)

Y	MON	PILOT UNIT	SAMPLE IDENTITY	FLOW RATE IGPM	UNFILTERED SAMPLE			FILTERED SAMPLE					
					BOD	COD	TKN	BOD	COD	TOC	NH3N	NO2N	NO3N
9	09	.5M1	EFG1600	1.01				82	25	3.0	0.0	0.0	6.0
9	09	.5M1	EFG1630	1.01				58	18	4.0	0.0	0.1	6.4
9	09	.5M1	EFG1700	1.04				54	15	4.0	0.0	0.1	5.5
9	09	.5M1	EFG1730	1.04				64	18	3.0	0.0	0.2	5.5
9	09	.5M1	EFG1800	1.04				55	16	4.0	0.0	0.0	6.3
9	09	.5M1	EFG1830	1.04				36	13	3.0	0.2	0.0	5.4
9	09	.5M1	EFG1900	1.04				42	15	4.0	0.2	0.2	5.5
9	09	.5M1	EFG1930	1.04				48	17	5.0	0.1	0.3	7.1
9	09	.5M1	EFG2000	1.04				56	17	5.0	0.1	0.2	7.0
9	09	.5M1	EFG2030	1.04				60	16	6.0	0.1	0.1	7.7
9	09	.5M1	EFG2100	1.04				61	18	6.0	0.0	0.3	6.4
9	09	.5M1	EFG2130	1.04				58	18	6.0	0.0	0.2	7.8
9	09	.5M1	EFG2200	0.507				59	16	6.0	0.0	0.0	7.3
9	09	.5M1	EFG2230	0.507				68	18	8.0	0.1	0.1	9.0
9	09	.5M1	EFG2300	0.507				79	21	9.0	0.0	0.0	12.0
9	09	.5M1	EFG2330	0.507				83	23	11.0	0.0	0.0	13.2
9	09	.5M1	EFG2400	0.507				83	22	12.0	0.0	0.1	14.0
9	09	.5M1	EFG0030	0.507				83	22	11.0	0.0	1.3	14.5
9	09	.5M1	EFG0100	0.507				77	23	12.0	0.0	0.1	13.8
9	09	.5M1	EFG0130	0.507				75	22	11.0	0.1	0.1	14.3
9	09	.5M1	EFG0200	0.507				72	20	11.0	0.1	0.2	13.8
9	09	.5M1	EFG0230	0.507				71	24	11.0	0.0	0.4	13.0
9	09	.5M1	EFG0300	1.03				73	21	11.0	0.0	0.1	14.4
9	09	.5M1	EFG0330	1.03				71	22	11.0	0.0	0.4	13.5
9	09	.5M1	EFG0400	1.03				89	27	11.0	0.0	0.3	12.6
9	09	.5M1	EFG0430	1.03				112	36	10.0	0.0	0.3	10.8
9	09	.5M1	EFG0500	1.03				88	29	8.0	0.0	0.6	9.9
9	09	.5M1	EFG0530	1.03				63	20	8.0	0.0	0.1	10.0
9	09	.5M1	EFG0600	1.03				79	27	7.0	0.0	1.4	9.0
9	09	.5M1	EFG0630	1.03				56	50	7.0	0.0	0.1	9.0
9	09	.5M1	EFG0700	1.03				57	59	7.0	0.0	0.1	8.4
9	09	.5M1	EFG0730	1.03				61	18	7.0	0.1	0.1	8.9
9	09	.5M1	EFG0800	1.03				59	18	7.0	0.0	0.1	8.9

BIO-SURF PILOT PLANT TESTING
1976

ANALYTICAL RESULTS - SERIES E2 (30 MIN)

Y	MON	PILOT UNIT	SAMPLE IDENTITY	FLOW RATE IGPM	UNFILTERED SAMPLE			FILTERED SAMPLE					
					BOD	COO	TKN	BOD	COO	TOC	NH3N	NO2N	NO3N
7	09	2M	EFG1000	15.37				47	17	7.0	0.1	0.1	8.4
7	09	2M	EFG1030	15.37				49	17	8.0	0.0	0.2	8.6
7	09	2M	EFG1100	15.37				57	20	11.0	0.0	0.2	12.4
7	09	2M	EFG1130	15.37				59	20	20.0	0.1	0.1	21.3
7	09	2M	EFG1200	15.37				68	20	15.0	0.0	0.2	16.2
7	09	2M	EFG1230	15.37				70	22	24.0	0.1	0.1	26.1
7	09	2M	EFG1300	15.37				72	21	22.0	0.1	0.1	24.1
7	09	2M	EFG1330	15.37				82	24	19.0	0.0	0.0	21.2
7	09	2M	EFG1400	15.37				98	32	19.0	0.0	0.0	19.2
7	09	2M	EFG1430	15.37				116	36	17.0	0.0	0.0	19.2
7	09	2M	EFG1500	34.60				169	41	16.0	0.0	0.0	18.0
7	09	2M	EFG1530	34.60				147	36	19.0	0.0	0.0	19.6
7	09	2M	EFG1600	34.60				132	37	15.0	0.0	0.0	16.1
7	09	2M	EFG1630	34.60				117	30	9.0	0.0	0.1	9.7
7	09	2M	EFG1700	34.60				81	21	13.0	0.0	0.2	14.9
7	09	2M	EFG1730	34.60				71	20	7.0	0.0	0.0	8.0
7	09	2M	EFG1800	34.60				64	19	10.0	0.0	0.3	11.1
7	09	2M	EFG1830	34.60				103	31	14.0	0.0	0.5	15.3
7	09	2M	EFG1900	34.60				105	33	15.0	0.0	0.1	15.9
7	09	2M	EFG1930	34.60				113	34	12.0	0.0	0.5	13.6
7	09	2M	EFG2000	17.11				93	24	13.0	0.0	0.7	13.8
7	09	2M	EFG2030	17.11				89	22	13.0	0.1	0.4	13.8
7	09	2M	EFG2100	17.11				90	22	13.0	0.1	0.4	14.1
7	09	2M	EFG2130	17.11				76	21	9.0	0.1	0.2	10.2
7	09	2M	EFG2200	17.11				69	22	9.0	0.1	0.1	10.2
7	09	2M	EFG2230	17.11				68	22	10.0	0.1	0.1	10.7
7	09	2M	EFG2300	17.11				68	19	11.0	0.1	0.2	11.1
7	09	2M	EFG2330	17.11				70	20	10.0	0.1	0.1	11.1
7	09	2M	EFG2400	17.11				60	21	10.0	0.1	0.1	10.6
8	09	2M	EFG0030	17.11				55	19	10.0	0.1	0.2	10.9
8	09	2M	EFG0100	16.00				62	21	9.0	0.1	0.1	10.1
8	09	2M	EFG0130	16.00				56	20	10.0	0.0	0.0	10.7
8	09	2M	EFG0200	16.00				71	24	09.0	0.0	0.0	9.2
8	09	2M	EFG0230	16.00				66	24	11.0	0.0	0.0	11.1
8	09	2M	EFG0300	16.00				64	21	11.0	0.0	0.1	12.1
8	09	2M	EFG0330	16.00				65	21	12.0	0.0	0.0	12.2
8	09	2M	EFG0400	16.00				68	20	12.0	0.0	0.1	13.1
8	09	2M	EFG0430	16.00				70	21	11.0	0.0	0.0	12.6
8	09	2M	EFG0500	16.00				64	20	11.0	0.0	0.0	11.7
8	09	2M	EFG0530	16.00				62	20	12.0	0.0	0.0	12.9
8	09	2M	EFG0600	32.40				61	19	12.0	0.0	0.0	13.7
8	09	2M	EFG0630	32.40				61	19	13.0	0.0	0.1	13.9
8	09	2M	EFG0700	32.40				64	20	12.0	0.0	0.1	13.1
8	09	2M	EFG0730	32.40				64	20	12.0	0.0	0.0	13.4
8	09	2M	EFG0800	32.40				65	20	11.0	0.0	0.0	12.6
8	09	2M	EFG0830	32.40				58	18	11.0	0.1	0.1	12.9
8	09	2M	EFG0900	32.40				61	19	11.0	0.0	0.0	12.1
8	09	2M	EFG0930	32.40				55	17	12.0	0.0	0.0	12.9
8	09	2M	EFG1000	32.40				55	17	18.0	0.0	0.0	20.1
8	09	2M	EFG1030	32.40				61	19	20.0	0.0	0.0	24.8
8	09	2M	EFG1100	16.20				60	19	24.0	0.0	0.0	25.1
8	09	2M	EFG1130	16.20				50	16	20.0	0.0	0.2	21.8
8	09	2M	EFG1200	16.20				40	14	14.0	0.0	0.3	16.6

ANALYTICAL RESULTS - SERIES E2 (30 MIN)

Y	MON	PILOT	SAMPLE	FLOW RATE	UNFILTERED			FILTERED						
					SAMPLE			SAMPLE						
			UNIT	IGPM	BOD	COD	TKN	BOD	COD	TOC	NH3N	NO2N	NO3N	TKN
8	09	2M	EFG1230	16.20				42	13	16.0	0.1	0.3	16.2	
8	09	2M	EFG1300	16.20				42	14	14.0	0.1	0.6	15.5	
8	09	2M	EFG1330	16.20				42	15	11.0	0.3	0.4	14.5	
8	09	2M	EFG1400	16.20				45	15	15.6	0.1	0.4	17.1	
8	09	2M	EFG1430	16.20				49	16	18.0	0.1	0.3	18.2	
8	09	2M	EFG1500	16.20				47	16	18.0	0.1	0.4	18.3	
8	09	2M	EFG1530	16.20				51	15	14.0	0.1	0.5	15.6	
8	09	2M	EFG1600	34.10				540	46	13.0	0.0	0.0	15.1	
8	09	2M	EFG1630	34.10				109	35	18.0	0.0	0.0	19.1	
8	09	2M	EFG1700	34.10				106	38	19.0	0.0	0.0	21.5	
8	09	2M	EFG1730	34.10				67	64	18.0	0.0	0.0	18.4	
8	09	2M	EFG1800	34.10				79	23	14.0	0.0	0.0	16.4	
8	09	2M	EFG1830	34.10				93	32	15.0	0.0	0.5	16.8	
8	09	2M	EFG1900	34.10				89	27	22.0	0.2	2.4	23.3	
8	09	2M	EFG1930	34.10				108	33	14.0	0.1	0.4	15.6	
8	09	2M	EFG2000	34.10				78	22	13.0	0.0	0.1	14.1	
8	09	2M	EFG2030	34.10				83	25	12.0	0.0	0.0	13.1	
8	09	2M	EFG2100	34.00				143	46	15.0	0.0	0.0	16.3	
8	09	2M	EFG2130	34.00				211	63	19.0	0.0	0.0	20.8	
8	09	2M	EFG2200	34.00				179	55	16.0	0.0	0.0	17.7	
8	09	2M	EFG2230	34.00				159	55	15.0	0.0	0.0	16.2	
8	09	2M	EFG2300	34.00				152	54	14.0	0.0	0.0	15.0	
8	09	2M	EFG2330	34.00				136	49	14.0	0.0	0.0	15.1	
8	09	2M	EFG2400	34.00				148	47	13.0	0.0	0.0	13.9	
9	09	2M	EFG0030	34.00				131	45	13.0	0.0	0.0	14.2	
9	09	2M	EFG0100	34.00				133	46	13.0	0.0	0.0	14.1	
9	09	2M	EFG0130	34.00				143	49	13.0	0.0	0.0	14.1	
9	09	2M	EFG0200	15.20				120	42	13.0	0.0	0.0	14.1	
9	09	2M	EFG0230	15.20				89	30	13.0	0.0	0.0	14.1	
9	09	2M	EFG0300	15.20				81	27	13.0	0.0	0.0	13.7	
9	09	2M	EFG0330	15.20				72	25	14.0	0.0	0.0	15.6	
9	09	2M	EFG0400	15.20				70	23	14.0	0.0	0.0	14.9	
9	09	2M	EFG0430	15.20				65	25	13.0	0.0	0.2	13.6	
9	09	2M	EFG0500	15.20				63	22	12.0	0.0	0.0	13.6	
9	09	2M	EFG0530	15.20				63	23	13.0	0.0	0.1	13.8	
9	09	2M	EFG0600	15.20				63	22	12.0	0.0	0.0	13.1	
9	09	2M	EFG0630	15.20				58	20	13.0	0.0	0.1	14.6	
9	09	2M	EFG0700	32.30				58	23	12.0	0.0	0.0	13.3	
9	09	2M	EFG0730	32.30				79	29	12.0	0.0	0.0	13.3	
9	09	2M	EFG0800	32.30				94	33	12.0	0.0	0.0	12.9	
9	09	2M	EFG0830	32.30				91	30	11.0	0.0	0.0	11.9	
9	09	2M	EFG0900	32.30				86	30	11.0	0.0	0.0	12.6	
9	09	2M	EFG0930	32.30				92	31	10.0	0.0	0.1	11.2	
9	09	2M	EFG1000	32.30				105	35	13.0	0.0	0.0	14.1	
9	09	2M	EFG1030	32.30				111	37	15.0	0.0	0.0	17.8	
9	09	2M	EFG1100	32.30				107	40	17.0	0.0	0.0	19.9	
9	09	2M	EFG1130	32.30				73	88	20.0	0.0	0.0	24.6	
9	09	2M	EFG1200	32.80				122	45	21.0	0.0	0.0	24.2	
9	09	2M	EFG1230	32.80				113	37	10.0	0.0	0.0	13.0	
9	09	2M	EFG1300	32.80				105	32	6.0	0.0	0.1	8.3	
9	09	2M	EFG1330	32.80				113	35	6.0	0.0	0.1	8.6	
9	09	2M	EFG1400	32.80				108	39	5.0	0.0	0.1	7.6	
9	09	2M	EFG1430	32.80				114	37	5.0	0.0	0.1	6.2	
9	09	2M	EFG1500	32.80				123	40	5.0	0.0	0.0	6.2	
9	09	2M	EFG1530	32.80				114	37	4.0	0.0	0.0	5.0	

ANALYTICAL RESULTS - SERIES E2 (30 MIN)

MON	PILOT	SAMPLE	FLOW RATE	UNFILTERED			FILTERED						
				UNIT	IDENTITY	IGPM	BOD	COD	TKN	BOD	COD	TOC	NH3N
09	2M	EFG1230	16.20					42	13	16.0	0.1	0.3	16.2
09	2M	EFG1300	16.20					42	14	14.0	0.1	0.6	15.5
09	2M	EFG1330	16.20					42	15	11.0	0.3	0.4	14.5
09	2M	EFG1400	16.20					45	15	15.6	0.1	0.4	17.1
09	2M	EFG1430	16.20					49	16	18.0	0.1	0.3	18.2
09	2M	EFG1500	16.20					47	16	18.0	0.1	0.4	18.3
09	2M	EFG1530	16.20					51	15	14.0	0.1	0.5	15.6
09	2M	EFG1600	34.10					540	46	13.0	0.0	0.0	15.1
09	2M	EFG1630	34.10					109	35	18.0	0.0	0.0	19.1
09	2M	EFG1700	34.10					106	38	19.0	0.0	0.0	21.5
09	2M	EFG1730	34.10					67	64	18.0	0.0	0.0	18.4
09	2M	EFG1800	34.10					79	23	14.0	0.0	0.0	16.4
09	2M	EFG1830	34.10					93	32	15.0	0.0	0.5	16.8
09	2M	EFG1900	34.10					89	27	22.0	0.2	2.4	23.3
09	2M	EFG1930	34.10					108	33	14.0	0.1	0.4	15.6
09	2M	EFG2000	34.10					78	22	13.0	0.0	0.1	14.1
09	2M	EFG2030	34.10					83	25	12.0	0.0	0.0	13.1
09	2M	EFG2100	34.00					143	46	15.0	0.0	0.0	16.3
09	2M	EFG2130	34.00					211	63	19.0	0.0	0.0	20.8
09	2M	EFG2200	34.00					179	55	16.0	0.0	0.0	17.7
09	2M	EFG2230	34.00					159	55	15.0	0.0	0.0	16.2
09	2M	EFG2300	34.00					152	54	14.0	0.0	0.0	15.0
09	2M	EFG2330	34.00					136	49	14.0	0.0	0.0	15.1
09	2M	EFG2400	34.00					148	47	13.0	0.0	0.0	13.9
09	2M	EFG0030	34.00					131	45	13.0	0.0	0.0	14.2
09	2M	EFG0100	34.00					133	46	13.0	0.0	0.0	14.1
09	2M	EFG0130	34.00					143	49	13.0	0.0	0.0	14.1
09	2M	EFG0200	15.20					120	42	13.0	0.0	0.0	14.1
09	2M	EFG0230	15.20					89	30	13.0	0.0	0.0	14.1
09	2M	EFG0300	15.20					81	27	13.0	0.0	0.0	13.7
09	2M	EFG0330	15.20					72	25	14.0	0.0	0.0	15.6
09	2M	EFG0400	15.20					70	23	14.0	0.0	0.0	14.9
09	2M	EFG0430	15.20					65	25	13.0	0.0	0.2	13.6
09	2M	EFG0500	15.20					63	22	12.0	0.0	0.0	13.6
09	2M	EFG0530	15.20					63	23	13.0	0.0	0.1	13.8
09	2M	EFG0600	15.20					63	22	12.0	0.0	0.0	13.1
09	2M	EFG0630	15.20					58	20	13.0	0.0	0.1	14.6
09	2M	EFG0700	32.30					58	23	12.0	0.0	0.0	13.3
09	2M	EFG0730	32.30					79	29	12.0	0.0	0.0	13.3
09	2M	EFG0800	32.30					94	33	12.0	0.0	0.0	12.9
09	2M	EFG0830	32.30					91	30	11.0	0.0	0.0	11.9
09	2M	EFG0900	32.30					86	30	11.0	0.0	0.0	12.6
09	2M	EFG0930	32.30					92	31	10.0	0.0	0.1	11.2
09	2M	EFG1000	32.30					105	35	13.0	0.0	0.0	14.1
09	2M	EFG1030	32.30					111	37	15.0	0.0	0.0	17.8
09	2M	EFG1100	32.30					107	40	17.0	0.0	0.0	19.9
09	2M	EFG1130	32.30					73	38	20.0	0.0	0.0	24.6
09	2M	EFG1200	32.80					122	45	21.0	0.0	0.0	24.2
09	2M	EFG1230	32.80					113	37	10.0	0.0	0.0	13.0
09	2M	EFG1300	32.80					105	32	6.0	0.0	0.1	8.3
09	2M	EFG1330	32.80					113	35	6.0	0.0	0.1	8.6
09	2M	EFG1400	32.80					108	39	5.0	0.0	0.1	7.6
09	2M	EFG1430	32.80					114	37	5.0	0.0	0.1	5.2
09	2M	EFG1500	32.80					123	40	5.0	0.0	0.0	6.2
09	2M	EFG1530	32.80					114	37	4.0	0.0	0.0	5.0

ANALYTICAL RESULTS - SERIES E2 (30 MIN)												
Y	MON	PILOT SAMPLE UNIT IDENTITY	FLOW RATE IGPM	UNFILTERED SAMPLE			FILTERED SAMPLE					
				BOD	COD	TKN	BOD	COD	TOC	NH3N	NO2N	NO3N
9	09	2M	EFG1600	32.80			106	35	4.0	0.0	0.0	5.7
9	09	2M	EFG1630	32.80			100	33	6.0	0.0	0.0	6.3
9	09	2M	EFG1700	32.40			96	30	6.0	0.0	0.0	6.1
9	09	2M	EFG1730	32.40			63	22	4.0	0.0	0.1	5.7
9	09	2M	EFG1800	32.40			55	18	5.0	0.1	0.0	6.9
9	09	2M	EFG1830	32.40			45	16	4.0	0.0	0.2	4.7
9	09	2M	EFG1900	32.40			54	20	5.0	0.1	0.0	6.8
9	09	2M	EFG1930	32.40			82	23	6.0	0.0	0.1	8.6
9	09	2M	EFG2000	32.40			70	23	6.0	0.0	0.1	7.3
9	09	2M	EFG2030	32.40			75	27	6.0	0.0	0.1	8.2
9	09	2M	EFG2100	32.40			73	23	6.0	0.0	0.1	8.0
9	09	2M	EFG2130	32.40			84	27	7.0	0.0	0.1	8.4
9	09	2M	EFG2200	15.95			61	18	6.0	0.0	0.1	7.2
9	09	2M	EFG2230	15.95			77	23	8.0	0.0	0.5	10.1
9	09	2M	EFG2300	15.95			95	28	10.0	0.0	0.5	13.4
9	09	2M	EFG2330	15.95			98	32	11.0	0.0	0.1	14.9
9	09	2M	EFG2400	15.95			92	28	12.0	0.0	0.1	14.0
0	09	2M	EFG0030	15.95			88	29	13.0	0.0	0.1	14.6
0	09	2M	EFG0100	15.95			86	24	12.0	0.0	0.2	14.6
0	09	2M	EFG0130	15.95			66	59	12.0	0.0	0.3	14.0
0	09	2M	EFG0200	15.95			48	25	12.0	0.0	0.2	14.0
0	09	2M	EFG0230	15.95			84	25	12.0	0.0	0.2	14.0
0	09	2M	EFG0300	32.00			92	29	12.0	0.1	0.0	14.6
0	09	2M	EFG0330	32.00			118	40	11.0	0.3	1.0	13.5
0	09	2M	EFG0400	32.00			114	37	11.0	0.0	0.5	13.3
0	09	2M	EFG0430	32.00			149	51	10.0	0.0	0.2	10.9
0	09	2M	EFG0500	32.00			104	37	09.0	0.0	0.1	10.0
0	09	2M	EFG0530	32.00			91	30	8.0	0.0	0.9	10.7
0	09	2M	EFG0600	32.00			85	28	9.0	0.0	0.1	10.6
0	09	2M	EFG0630	32.00			92	31	8.0	0.0	0.1	10.2
0	09	2M	EFG0700	32.00			106	31	8.0	0.0	0.4	10.9
0	09	2M	EFG0730	32.00			91	30	8.0	0.0	0.2	10.8
0	09	2M	EFG0800	32.00			88	29	8.0	0.0	0.1	10.5

APPENDIX B
ANALYTICAL PROCEDURES

Total Kjeldahl Nitrogen

Total kjeldahl nitrogen analyses (organic plus ammonia nitrogen) were performed according to Technicon Auto-analyser Industrial Method 146-71A. Essentially this procedure consists of digestion of organic matter at 380°C followed by measurement of the ammonia produced using the Berthelot reaction in which the formation of a blue indo-phenol complex occurs when ammonia reacts with sodium phenate followed by the addition of sodium hypochlorite. Glycine standards were used for calibration. For keeping unfiltered samples homogenized in the sample cups the system has two air aspirators. One aspirator provides complete mixing in the cup being sampled while the second aspirator mixes the next cup on the tray.

Ammonia

Analyses of ammonia nitrogen were conducted using Technicon Auto-analyser Industrial Method 98-70W. This is essentially the same technique employed for Total kjeldahl nitrogen with the omission of the selenium dioxide/sulphuric acid/perchloric acid digestion step which ammonifies the organic nitrogen fraction. Ammonium chloride standards provided calibration.

Nitrate

Technicon Auto-analyser Industrial Method 100-70W was used for nitrite-nitrogen determinations. This technique involves a reaction between nitrite and sulphanilamide under acid conditions to form a diazo compound which in turn is

coupled with N-1-naphthylethylenediamine to form a reddish purple azo dye. Colourimetric determination is then made on the sample.

Nitrate plus Nitrite

Nitrate plus nitrite-nitrogen analyses were performed using Technicon Auto-analyser Industrial Method 100-70W. In this method, the nitrate-nitrogen is reduced to nitrite in the copper-cadium reduction column. The sample is then analysed for nitrite nitrogen as described previously.

Chemical Oxygen Demand (COD)

COD determinations on 24 hour composite samples were done according to the dichromate reflux method described in "Standard Methods" (1971). During the dynamic experiments, when as many as 600 samples were collected for COD analyses in the space of 3-5 days, a less time consuming analytical technique was employed. A modified version of Technicon Auto-analyser Industrial Method No. 268-73W was adapted for COD analysis and a Technicon Solid prep 11 sampler was introduced in place of the normal sampler. Standard solutions were prepared using ammonium chloride. The standards were first analysed using the "Standard Methods" reflux technique and then analysed on the Technicon equipment. The standard peaks produced on the Technicon System were then calibrated against the "Standard Methods" results.

This complicated approach was necessary since the sample digestion time in the Auto-analyser was shorter than that in the standard reflux test. This resulted in a lower degree of reaction completion with the Auto-analyser when heterogeneous sewage samples were tested. With this

procedure modification in effect, Auto-analyser COD results for sewage samples were generally only 5 to 7 percent lower than results obtained via the "Standard Methods" technique.

Biochemical Oxygen Demand (BOD)

The 5 day, 20°C BOD determinations were performed according to the method described in "Standard Methods" pages 489-495 (1971).

Total Organic Carbon (TOC)

Twenty micro-litre samples previously acidified and purged were injected into a Beckman Infrared Carbon Analyser. The resulting peaks were compared to a calibration curve prepared from standards using anhydrous potassium biphthalate.

Suspended Solids

GELMAN .45 micron glass fibre filters were dried, Previously Washed, for at least two hours in a 103 degree C oven. They were then cooled in a dessicator and weighed. Suspended solids determinations were made by filtering a minimum of 10 ml of solution through a filter. The filter was then re-dried at 103 degrees for two or more hours, dessicated for 15 minutes and re-weighed. The increase in weight was taken as a measure of the suspended solids.

Dissolved Oxygen

An Electronic Instruments Ltd. Dissolved Oxygen Metre Model 15A was used for dissolved oxygen determinations. It was found necessary to calibrate the probe roughly once a week.

Temperature

The D.O. metre also included a temperature probe and this was used for measurement of the feed stream and the reactor temperatures. Occasionally, a mercury thermometer was also used.

pH

pH was measured using an Orion Specific Ion Meter (Model 401) together with Fisher Combination electrodes (Cat. 1e-639-90).

Alkalinity

By using the Orion pH meter, 50 ml samples were titrated to a pH of 4.8 by addition of .02 N sulphuric acid. Results were expressed as mg/l of calcium carbonate.

APPENDIX CCALCULATION PROCEDURESIdentification of the Transfer Function Model:TKN Load vs TKN Concentration

After sufficient differencing to induce stationarity in the data, the impulse response weights of the cross correlation function (Figure 4.1) were examined in order to identify the transfer function model orders (r, s, b). In this case the model orders may be tentatively identified as a (1, 1, 1) or a (1, 2, 1) process (Figure 4.1). In order to determine which model provided the best representation of the data, the models must be built and checked, and the residual sums of squares of the models compared.

The sample calculation will continue with the (1, 1, 1) model form. The preliminary transfer function model may therefore be identified as

$$Y_t = \frac{(W_0 - W_1 \beta) \beta X_t}{(1 - \delta_1 \beta)} \quad \dots (C-1)$$

The preliminary estimates of the above model parameters were calculated from the impulse response weights:

$$W_0 = v_{j=b} = 0.083$$

$$\delta_1 W_0 - W_1 = v_{j=b+1} = 0.088$$

$$\delta_1 v_{j-1} = v_{j=b+1} = 0.065$$

Identification of the Noise Model

The model under consideration is

$$Y_t = \frac{W(\beta)}{\delta(\beta)} X_{t-b} + N_t \quad \dots (C-2)$$

where N_t is to be parsimoniously represented as

$$N_t = \frac{\theta(\beta)}{\Delta^d \phi(\beta)} a_t \quad \dots (C-3)$$

Having already identified the transfer function model, N_t may be obtained from

$$N_t = Y_t - \delta_1 Y_{t-1} - w_0 X_{t-1} + w_1 X_{t-2} + \delta_1 N_{t-1} \quad \dots (C-4)$$

Plots of the autocorrelation and partial autocorrelation functions of the noise sequence are given in Figure C-1. Examination of these functions reveals a decaying autocorrelation function which becomes negligible after lag 1. A single significant spike at lag 1 is noted for the partial autocorrelation function. This is indicative of an autoregressive process of order 1 (AR(1)).

$$\text{Therefore, } N_t = \frac{a_t}{(1 - \phi_1 \beta)} \quad \dots (C-5)$$

$$\text{with } \phi_1 = \rho_1 = 0.219$$

Our tentatively identified model is therefore

$$Y_t = \frac{(w_0 - w_1 \beta) \beta X_t}{(1 - \delta_1 \beta)} + \frac{a_t}{(1 - \phi_1 \beta)} \quad \dots (C-6)$$

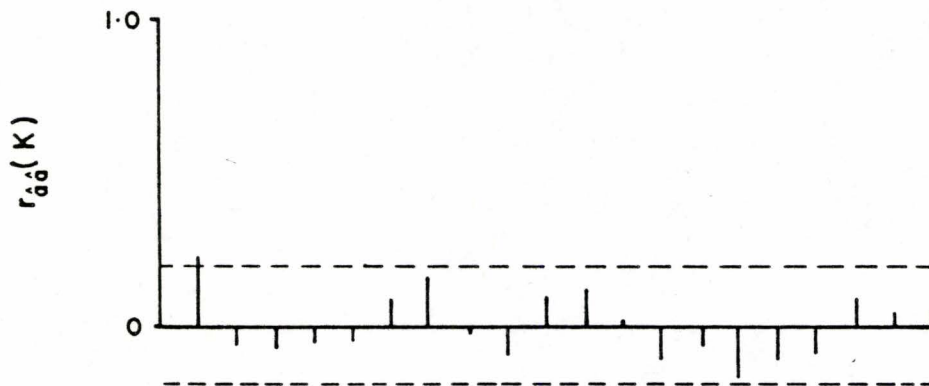
Fitting and Applying Residual Checks to the Combined
Transfer Function - Noise Model (TF-N)

The initial parameter estimates obtained from the previous

FIGURE C-1

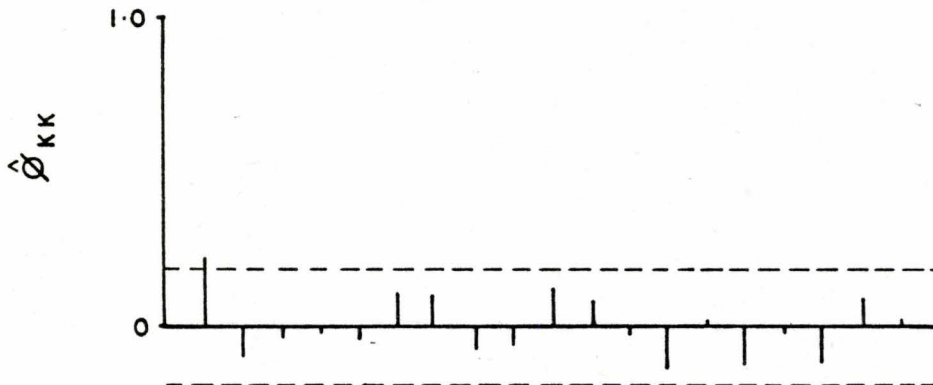
AUTOCORRELATION AND PARTIAL AUTOCORRELATION
FUNCTIONS OF TKN LOAD MODEL NOISE SEQUENCE

AUTOCORRELATION FUNCTION



APPROXIMATE 95% CONFIDENCE LIMITS

PARTIAL AUTOCORRELATION FUNCTION



0 2 4 6 8 10 12 14 16 18 20

LAG K (Hrs.)

sections were used as starting values in TS HAUS (TS HAUS is an efficient non-linear least squares program developed for use with time series analysis). The dynamic and noise parameters were estimated simultaneously. The parameter estimates and their approximate 95% confidence limits appear in Table C-1A.

TABLE C-1A
SAMPLE CALCULATION MODEL ESTIMATES

<u>Model</u>	<u>Initial Estimates</u>	<u>Final Estimates</u>	<u>95% Confidence Interval</u>
(1,1,1)	w_0 0.083	0.08312	± 0.01738
	w_1 -0.0267	-0.03450	± 0.02426
	δ_1 0.739	0.5590	± 0.1365
	ϕ_1 0.219	0.2339	± 0.1808

The two most frequent checks applied to diagnose transfer function noise model adequacy (or inadequacy as the case may be) are the autocorrelation function $\hat{r}_{aa}^{\wedge}(k)$ of the residuals from the fitted model and the cross correlation function between prewhitened input (or in our case the first difference Δx_t) and the model residuals. Assuming the form of the TF - N model is correct and that the parameter values are known, then the estimated autocorrelation function of the residuals will not be correlated and will be distributed normally about zero with variance n^{-1} , where n is the number of data points in the series. These checks are presented in Figures C-2 and C-3. The results in Figures C-2 and C-3 indicate that there is no serious transfer function model or noise model inadequacy and that the chosen models adequately represent the data.

FIGURE C-2
 AUTOCORRELATION OF THE FITTED TKN LOAD TF-N
 MODEL RESIDUALS

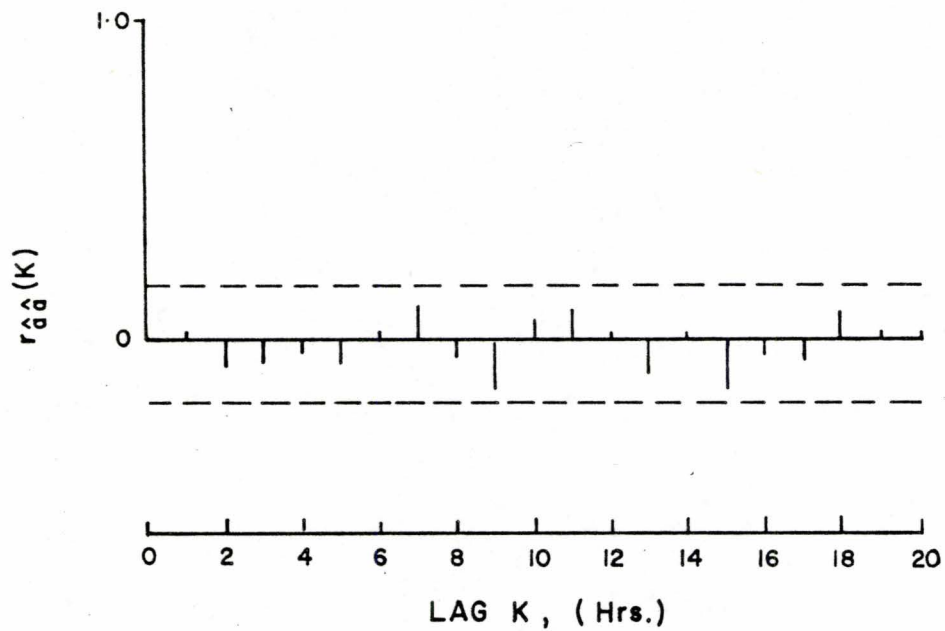
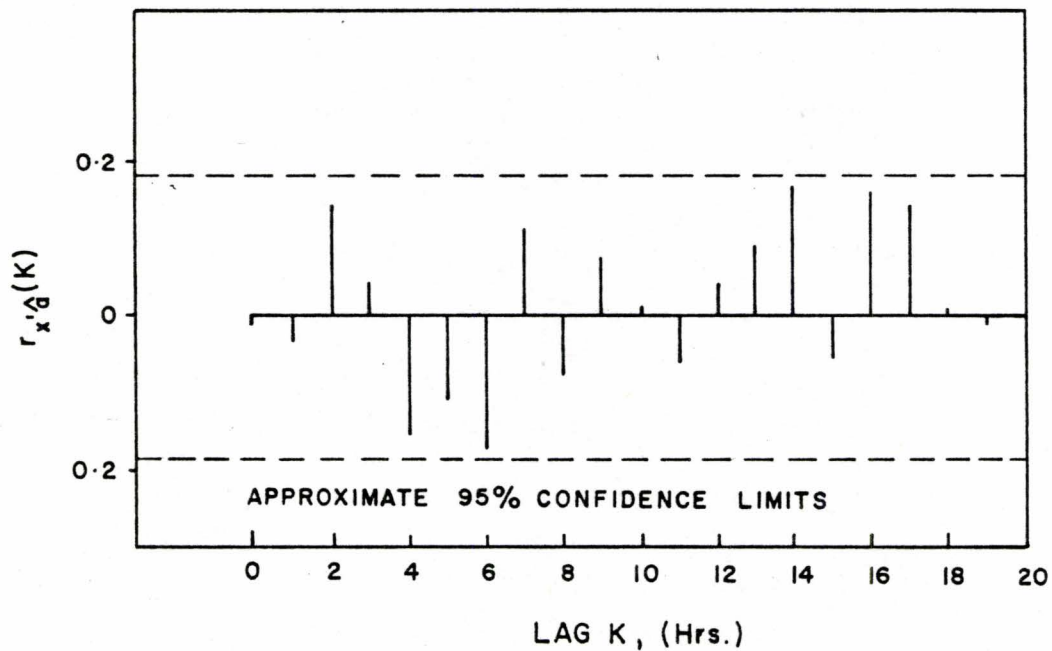


FIGURE C-3
 CROSS CORRELATION MODEL CHECK, ∇X_t vs. TF-N
 MODEL RESIDUALS



A further assessment of the residuals involves taking the first k autocorrelations and computing the Q statistic. Comparing the result to the chi-square distribution (χ^2) with $k-p-q$ degrees of freedom, where $p + q$ is the total number of parameters of the noise model, determines the significance of the residuals. In a similar manner the significance of the cross correlations between residuals and the stationary input series for each variable can be computed by comparing the S statistic to the chi-square (χ^2) distribution. These results were presented in Table 4.3 for the models obtained in this work.

A complete listing of the computer programs used is provided in Tables C1, C2, C3 and C4.

Forecasting System Response to Impulse Forcings

The forecasted effluent values discussed in Section 4.4.1 were determined using the minimum mean square error forecast with leading indicators, as developed by Box and Jenkins 1976, pgs. 402 to 405. The model used the following as input data:

- 1) past values of influent TKN load in g/day at discrete 1 hour time intervals
- 2) past values of effluent TKN concentration in mg/l at discrete time intervals
- 3) initial estimates for the error function, a_t .

The initial estimates used in (3) above were obtained by calculating residuals for the forecast and the experimental data with a_t initially set at zero. Calculated

residuals were used to replace the estimate of zero and calculations were continued in this iterative fashion.

A listing of the computer program and the plotting routine used is provided in Table C-5.

Transfer Function Calculation Procedures to Determine Response Times (From Box and Jenkins (1976))

The continuous system satisfying

$$(1+TD)Y(t) = g X (t-b-c) \quad \dots (C-7)$$

is for a pulsed input, discretely coincident with the discrete system satisfying

$$(1-\delta\beta)Y_t = (W_0 - W_1 \beta) X_{t-b-1+} \quad \dots (C-8)$$

where

$$\delta_1 = e^{-1/T}, \quad W_0 = g (1-\delta^{1-c}),$$

$$W_1 = g (\delta - \delta^{1-c})$$

The gain, g and the fractional time delay of the system, c , are independent of the sample interval. T is the average residence time in the system.

The constants for loading model A1 (Table 4.2) are as follows:

$$\begin{aligned} W_0 &= 0.08312 \\ W_1 &= -0.03450 \\ \delta_1 &= 0.559 \end{aligned}$$

Therefore
$$g = \frac{W_0 - W_1}{\delta^1} = 0.210$$

For the relationship:

$$W_0 = g (1 - \delta^{1-c}) \quad \dots (C-9)$$

the only unknown is the fractional time delay, c . Substituting into equation C-9,

$$c = 0.134$$

Since discrete sampling took place at hourly intervals, the approximate time delay of the system is about 10 minutes for loading influent variations.

Computer Program Used For Cross Correlations and Matrix
Inversion, Sample Output For Model A-1

```

LN J001      0      CROSS CORRELATIONS AND W WEIGHTS FOR SSC SYSTEM FILTER
LN J002      0      LOADING VERSUS EFFLUENT FILTERABLE TKN
LN J003      0      DIMENSION A(14,121),X(121),Y(121),Z(121),W(121)
LN J004      000000
LN J005      000000
LN J006      000000
LN J007      000000      NL = NUMBER OF LAGS
LN J008      000000
LN J009      000000
LN J010      000000
LN J011      DATA NOB/121/,NL/20/
LN J012      READ 1,((A(I,J),I=1,7),J=1,NOB)
LN J013      READ 2,((A(I,J),I=8,14),J=1,NOB)
LN J014      1  FORMAT(26X,F5.3,19X,F4.0,1X,F3.0,1X,4F5.1)
LN J015      2  FORMAT(26X,F5.3,21X,F3.0,1X,F3.0,1X,4F5.1)
LN J016      3  READ 4,(IX,IY,IW)
LN J017      4  FORMAT(3I2)
LN J018      PRINT 4,IX,IY,IW
LN J019      IF(IFEOF(60).EQ.-1) STOP
LN J020      PRINT 12
LN J021      12  FORMAT(28X,10HFILTERABLE,31X,10HFILTERABLE)
LN J022      PRINT 11
LN J023      11  FORMAT(10X,14HFLOW(IGAL/MIN),3X,12HRF TKN(MG/L),5X,
LN J024      11 5HLOADING(GM/DAY),8X,13HEFF TKN(MG/L))
LN J025      PRINT 9
LN J026      9  FORMAT(16X,1HX,16X,1HY,19X,1HZ,19X,1HW)
LN J027      SW=0.0
LN J028      SZ=0.0
LN J029      DO 5 K=1,NOB
LN J030      X(K)=A(IX,K)
LN J031      Y(K)=A(IY,K)
LN J032      W(K)=A(IW,K)
LN J033      Z(K)=(X(K)*Y(K))*6.552
LN J034      SZ=SZ+Z(K)
LN J035      SW=SW+W(K)
LN J036      PRINT 10,(X(K),Y(K),Z(K),W(K))
LN J037      10  FORMAT(9X,F10.2,7X,F10.2,10X,F10.2,10X,F10.2)
LN J038      5  CONTINUE
LN J039      AW=SW/FLOAT(NO3)
LN J040      AZ=SZ/FLOAT(NO3)
LN J041      PRINT 101
LN J042      101  FORMAT(///,24X,6HZ MEAN,15X,6HW MEAN)
LN J043      PRINT 102,AZ,AW
LN J044      102  FORMAT(/,20X,F10.3,10X,F10.3)
LN J045      DO 81 K=1,NOB
LN J046      W(K)=W(K)-AW
LN J047      81  Z(K)=Z(K)-AZ
LN J048      CALL IDENT-IF(Z,W,NO3,NL,1)
LN J049      GO TO 3
LN J050      END

```

USASI FORTRAN DIAGNOSTIC RESULTS FOR FTN.MAIN

NO ERRORS

```

0001 SUBROUTINE IDENT TF(X,Y,NOB,NL,NDIFF)
0002 C IDENTIFICATION OF THE IMPULSE RESPONSE
0003 C BY INVERSION OF MATRIX
0004 C THIS SUBROUTINE REQUIRES THE SSPLIB
0005 C SSPLIB MUST BE ATTACHED
0006 C COMMON SDA,SDX
0007 DIMENSION X(1),Y(1),AC(60),GAM(20,20),VGAM(400)
0008 DIMENSION CC1(21),CC2(21),L(400),M(400),V(20),VN(20)
0009 NDATA=NOB
0010 ND=0
0011 PRINT 18
0012 18 FORMAT(52H1 CROSS CORRELATIONS AT + AND - LAGS FOR NDIFF = 0)
0013 5 CALL CROSS(X,Y,NOB,NL,CC1,CC2)
0014 CALL ACORR(X,AC,SDZ,NOB,NL)
0015 C BUILDING THE MATRIX OF INPUT AUTOCORRELATION GAM
0016 NLL=NL-1
0017 DO 10 J=1,NLL
0018 GAM(J,J)=1.0
0019 II=NL-J+1
0020 DO 20 I=2,II
0021 IW=I+J-1
0022 20 GAM(IW,J)=AC(I-1)
0023 10 CONTINUE
0024 GAM(NL,NL)=1.0
0025 DO 30 J=2,NL
0026 JJ=J-1
0027 DO 40 I=1,JJ
0028 40 GAM(I,J)=GAM(J,I)
0029 30 CONTINUE
0030 PRINT 42,((GAM(I,J),I=1,NL),J=1,NL)
0031 42 FORMAT(10X,20F6.2)
0032 C TRANSFORMING GAM TO A VECTOR MATRIX VGAM
0033 DO 50 J=1,20
0034 DO 60 I=1,20
0035 IR=NL*(J-1)+I
0036 VGAM(IR)=GAM(I,J)
0037 60 CONTINUE
0038 50 CONTINUE
0039 C INVERSION OF MATRIX
0040 C MINV AND GMPD ARE IBM SCIENTIFIC SUBROUTINES
0041 N=NL*NL
0042 CALL MINV(VGAM,NL,D,L,M)
0043 PRINT 51,0
0044 51 FORMAT(//////,10X,4H 0 =,E20.8)
0045 NLL=NL*NL
0046 PRINT 42,(VGAM(I),I=1,NLL)
0047 C CALCULATION OF THE TRANSFER FUNCTION PARAMETERS
0048 CALL GAS003(VGAM,CC2,V,NL,NL,1)
0049 CALL GAS003(VGAM,CC1,VN,NL,NL,1)
0050 DO 16 I=1,NL
0051 V(I)=V(I)*SDA/SDX
0052 16 VN(I)=VN(I)*SDA/SDX
0053 C PLOTTING PARAMETERS V
0054 PRINT 8,ND
0055 3 FORMAT(///,10X,38H V WEIGHTS AT + AND - LAGS FOR NDIFF=,I2)
0056 CL=SQRT(VGAM(1)/FLOAT(NOB))*SDA*2.0/SDX
0057 CLN=-CL
0058 DO 2 K=1,NL
0059 T=FLOAT(K)
0060 IT=-T
0061 KK=K-1
0062 K1=-KK
0063 2 PRINT 4,K1,VN(K),KK,V(K)
0064 4 FORMAT(5X,I3,5X,F6.3,10X,I3,5X,F6.3)
0065 PRINT 12,CL
0066 12 FORMAT(//,53H APPROX. 95 PER CENT CONF LIMIT ON IMPULSE RESPONS
0067 1,F10.3)
0068 NDATA=NDATA-1
0069 ND=ND+1
0070 IF(ND.GT.NDIFF)GO TO 100
0071 DO 41 I=1,NDATA
0072 X(I)=X(I+1)-X(I)
0073 41 Y(I)=Y(I+1)-Y(I)
0074 PRINT 19
0075 19 FORMAT(52H1 CROSS CORRELATIONS AT + AND - LAGS FOR NDIFF = 1)
0076 GO TO 6

```



```

0001 SUBROUTINE ACORR(Z,AC,SDZ,N,NL)
0002 DIMENSION Z(1),AC(1)
0003 NL1 = NL+1
0004 TN = N
0005 SZ = 0.
0006 DO 13 I=1,N
0007 13 SZ = SZ+Z(I)
0008 ZBAR = SZ/TN
0009 DO 10 JJ=1,NL1
0010 J = JJ-1
0011 SZZ = 0.
0012 NN = N-J
0013 DO 11 I=1,NN
0014 K = I+J
0015 11 SZZ = SZZ + (Z(I)-ZBAR)*(Z(K)-ZBAR)
0016 10 AC(JJ) = SZZ/TN
0017 SDZ = SQRT(AC(1))
0018 VZ = AC(1)
0019 DO 12 J=1,NL
0020 12 AC(J) = AC(J+1)/VZ
0021 RETURN
0022 END

```

USASI FORTRAN DIAGNOSTIC RESULTS FOR ACORR

NO ERRORS

```

0001 SUBROUTINE CROSS(X,A,NOB,NL,CC1,CC2)
0002 COMMON SDA,SDX
0003 DIMENSION X(NOB),A(NOB)
0004 DIMENSION CC1(41),CC2(41)
0005 C CC1 ARE CROSSCORRELATIONS AT NEGATIVE LAGS
0006 C CC2 ARE CROSSCORRELATION AT POSITIVE LAGS
0007 CALL CRCORR(X,A,CC2,SDX,SDA,NOB,NL)
0008 CALL CRCORR(A,X,CC1,SDA,SDX,NOB,NL)
0009 PRINT 6
0010 6 FORMAT(79H CROSS-CORRELATIONS BETWEEN MANIPULATED VARIABLES
0011 1 RESIDUALS X(T)*A(T+K),//)
0012 NL1 = NL+1
0013 DO 7 K=1,NL1
0014 KK = K-1
0015 K1 = -KK
0016 7 PRINT 8, -K1,CC1(K),KK,CC2(K)
0017 8 FORMAT(5X,I3,5X,F6.3,10X,I3,5X,F6.3)
0018 CL = 2.0/SQRT(FLOAT(NOB))
0019 PRINT 12, CL
0020 12 FORMAT(//,55H APPROX. 95 PERCENT CONF. LIMIT ON CROSS-CORRELATIO
0021 1 =,F6.3)
0022 PRINT 9, SDX,SDA
0023 9 FORMAT(/,28H STANDARD DEVIATIONS S(X) =,F12.4,5X,6HS(A) =,E12.4
0024 9 = 0.
0025 DO 10 J=1,NL1
0026 10 Q = Q+ CC2(J)*CC2(J)
0027 Q = Q*FLOAT(NOB)
0028 NDF = NL+1
0029 PRINT 11, Q,NDF
0030 11 FORMAT(/,25H CHI SQUARED STATISTIC = ,F6.2,/,10H BASED ON(,I2,46
0031 1 NO. OF DYNAMIC PARAMETERS) DEGREES OF FREEDOM)
0032 RETURN
0033 END

```

USASI FORTRAN DIAGNOSTIC RESULTS FOR CROSS

NO ERRORS


```

N J0001 SUBROUTINE CRCORR(X,Y,CC,SDX,SDY,N,NL)
N J0002 DIMENSION X(N),Y(N),CC(1)
N J0003 SX = 0.
N J0004 SY = 0.
N J0005 SXX = 0.
N J0006 SYY = 0.
N J0007 DO 2 I=1,N
N J0008 SX = SX+X(I)
N J0009 SY = SY+Y(I)
N J0010 SXX = SXX+X(I)*X(I)
N J0011 2 SYY = SYY+Y(I)*Y(I)
N J0012 TN = N
N J0013 SDX = SQRT((SXX-SX*SX/TN)/TN)
N J0014 SDY = SQRT((SYY-SY*SY/TN)/TN)
N J0015 NL1 = NL+1
N J0016 DO 3 K=1,NL1
N J0017 SXY = 0.
N J0018 NN = N-K+1
N J0019 DO 4 I=1,NN
N J0020 KK = I+K-1
N J0021 4 SXY = SXY+(X(I)-SX/TN)*(Y(KK)-SY/TN)
N J0022 3 CC(K) = (SXY/TN)/(SDX*SDY)
N J0023 RETURN
N J0024 END
    
```

USASI FORTRAN DIAGNOSTIC RESULTS FOR CRCORR

NO ERRORS

LI3,POIR

FLOW(GAL/MIN)	FILTERABLE RF TKN(MG/L)	LOADING(GM/DAY)	FILTERABLE EFF TKN(MG/L)
X	Y	Z	W
0.40	85.00	223.88	50.00
0.40	107.00	281.83	47.00
0.40	99.00	260.76	42.00
0.40	106.00	263.39	45.00
0.40	115.00	302.90	39.00
0.40	78.00	205.44	42.00
0.40	60.00	158.03	39.00
0.40	70.00	184.37	44.00
0.15	103.00	102.79	39.00
0.15	118.00	114.42	39.00
0.15	113.00	109.58	44.00
0.15	109.00	105.70	47.00
0.15	94.00	31.15	45.00
0.15	93.00	90.18	45.00
0.15	85.00	82.42	48.00
0.15	105.00	101.82	47.00
0.15	130.00	184.24	47.00
0.15	170.00	164.85	50.00
0.15	178.00	172.61	50.00
0.15	170.00	164.85	52.00
0.15	170.00	164.85	50.00
0.15	225.00	218.18	50.00
0.15	130.00	126.66	53.00
0.15	122.00	118.30	56.00
0.43	89.00	252.49	50.00
0.43	76.00	215.61	43.00
0.43	70.00	196.59	45.00
0.43	72.00	204.27	39.00
0.43	82.00	232.64	41.00
0.43	93.00	263.84	36.00
0.43	90.00	255.33	33.00
0.43	79.00	224.12	34.00
0.10	218.00	142.83	33.00
0.10	278.00	182.15	34.00
0.10	250.00	163.80	42.00
0.10	256.00	163.80	39.00
0.10	250.00	163.80	41.00
0.10	235.00	153.97	42.00
0.10	232.00	152.01	47.00
0.10	232.00	152.01	50.00

V WEIGHTS AT + AND - LAGS FOR NDIFF= 1

113	0.0000	19	0.0005
113	0.0007	18	0.0007
113	0.0022	16	0.0010
113	0.0059	15	0.0012
113	0.0111	14	0.0015
113	0.0221	14	0.0022
113	0.0443	13	0.0022
113	0.0886	12	0.0014
113	0.1772	11	0.0006
113	0.3544	10	0.0004
113	0.7088	9	0.0003
113	1.4176	8	0.0003
113	2.8352	7	0.0003
113	5.6704	6	0.0003
113	11.3408	5	0.0003
113	22.6816	4	0.0003
113	45.3632	3	0.0003
113	90.7264	2	0.0003
113	181.4528	1	0.0003

APPROX. 95 PER CENT CONF LIMIT ON IMPULSE RESPONSE = 0.023
1 714

FLOW (IGAL/MIN)	FILTERABLE		LOADING (GM/DAY)	FILTERABLE	
	RF	TKN (MG/L)		EFF	TKN (MG)
X	Y	Z	W		
0.40	34.40	90.61	9.50		
0.40	36.20	95.35	16.20		
0.40	33.70	88.76	21.00		
0.40	35.60	93.77	22.30		
0.40	35.80	94.29	23.30		
0.40	31.60	83.23	24.00		
0.40	31.30	82.44	24.40		
0.40	30.60	80.60	23.80		
0.15	18.70	18.13	23.00		
0.15	18.60	18.00	17.00		
0.15	16.40	15.90	10.10		
0.15	15.70	15.22	2.60		
0.15	14.10	13.67	0.90		
0.15	12.90	12.51	0.60		
0.15	12.20	11.83	2.00		
0.15	11.00	10.67	1.00		
0.15	28.80	27.93	1.10		
0.15	33.20	32.19	3.00		
0.15	34.30	33.26	2.30		
0.15	44.20	42.86	2.30		
0.15	49.20	47.71	2.90		
0.15	51.30	49.75	3.10		
0.15	32.50	31.52	3.70		
0.15	29.00	28.12	3.30		
0.43	18.10	17.35	2.70		
0.43	13.00	12.66	2.20		
0.43	11.40	11.10	1.80		
0.43	9.20	8.90	1.40		
0.43	10.70	10.36	1.30		
0.43	11.60	11.19	1.40		
0.43	11.80	11.48	1.40		
0.43	11.40	11.14	1.30		
0.10	21.10	20.82	1.20		
0.10	21.10	20.82	0.60		
0.10	19.40	19.11	0.70		
0.10	19.50	19.27	1.00		
0.10	8.00	7.84	1.40		
0.10	16.70	16.54	1.50		
0.10	14.80	14.70	1.50		
0.10	13.30	13.17	1.50		
0.43	35.20	34.78	1.50		
0.43	32.90	32.44	0.70		
0.43	37.90	37.33	1.60		
0.43	41.60	41.06	2.30		
0.43	47.10	46.53	2.60		
0.43	49.80	49.20	3.00		
0.43	47.20	46.65	3.20		
0.43	45.70	45.12	3.30		
0.43	51.00	50.38	3.50		
0.43	48.10	47.46	3.20		
0.43	44.00	43.35	3.20		

Table C-1 Cont'd

0.012	0.014	0.016	0.018	0.020	0.022	0.024	0.026	0.028	0.030	0.032	0.034	0.036	0.038	0.040	0.042	0.044	0.046	0.048	0.050	0.052	0.054	0.056	0.058	0.060	0.062	0.064	0.066	0.068	0.070	0.072	0.074	0.076	0.078	0.080	0.082	0.084	0.086	0.088	0.090	0.092	0.094	0.096	0.098	0.100	0.102	0.104	0.106	0.108	0.110	0.112	0.114	0.116	0.118	0.120	0.122	0.124	0.126	0.128	0.130	0.132	0.134	0.136	0.138	0.140	0.142	0.144	0.146	0.148	0.150	0.152	0.154	0.156	0.158	0.160	0.162	0.164	0.166	0.168	0.170	0.172	0.174	0.176	0.178	0.180	0.182	0.184	0.186	0.188	0.190	0.192	0.194	0.196	0.198	0.200
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0.000	0.001	0.002	0.003	0.004	0.005	0.006	0.007	0.008	0.009	0.010	0.011	0.012	0.013	0.014	0.015	0.016	0.017	0.018	0.019	0.020	0.021	0.022	0.023	0.024	0.025	0.026	0.027	0.028	0.029	0.030	0.031	0.032	0.033	0.034	0.035	0.036	0.037	0.038	0.039	0.040	0.041	0.042	0.043	0.044	0.045	0.046	0.047	0.048	0.049	0.050	0.051	0.052	0.053	0.054	0.055	0.056	0.057	0.058	0.059	0.060	0.061	0.062	0.063	0.064	0.065	0.066	0.067	0.068	0.069	0.070	0.071	0.072	0.073	0.074	0.075	0.076	0.077	0.078	0.079	0.080	0.081	0.082	0.083	0.084	0.085	0.086	0.087	0.088	0.089	0.090	0.091	0.092	0.093	0.094	0.095	0.096	0.097	0.098	0.099	0.100
-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------	-------

	V WEIGHTS AT + AND - LAGS	FOR NOIFF= J
-1	0.266	0.325
-2	0.012	0.091
-3	0.003	0.032
-4	0.011	0.053
-5	0.022	0.029
-6	0.037	0.006
-7	0.040	0.003
-8	0.041	0.008
-9	0.126	0.028
-10	0.008	0.006
-11	0.033	0.013
-12	0.041	0.015
-13	0.003	0.004
-14	0.004	0.001
-15	0.038	0.006
-16	0.001	0.011
-17	0.048	0.011
-18	0.002	0.015
-19	0.011	0.011
-20	0.050	0.043

APPROX. 95 PER CENT CONF LIMIT ON IMPULSE RESPONSE = 0.142

Table C-1 Cont'd

CROSS CORRELATIONS AT + AND - LAGS FOR NDIFF = 1
 CROSS-CORRELATIONS BETWEEN MANIPULATED VARIABLES + RESIDUALS X(T)*A(

-0	0.042	0	0.042
-1	0.049	1	0.038
-2	0.015	2	0.037
-3	0.032	3	0.033
-4	0.055	4	0.022
-5	0.059	5	0.048
-6	0.141	6	0.022
-7	0.243	7	0.063
-8	0.443	8	0.033
-9	0.082	9	0.173
-10	0.014	10	0.143
-11	0.020	11	0.143
-12	0.038	12	0.122
-13	0.037	13	0.043
-14	0.021	14	0.057
-15	0.040	15	0.049
-16	0.023	16	0.029
-17	0.039	17	0.074
-18	0.003	18	0.141
-19	0.005	19	0.051
-20	0.038	20	0.013

APPROX. 95 PERCENT CONF. LIMIT ON CROSS-CORRELATIONS = 0.182

STANDARD DEVIATIONS S(X) = 16.9733 S(A) = 0.3288E+01

CHI SQUARED STATISTIC = 79.49
 BASED ON (21 NO. OF DYNAMIC PARAMETERS)

	DEGREES OF FREEDOM									
1.00	-0.01	0.08	0.03	0.06	0.04	-0.04	-0.02	-0.35	-0.05	-0.01
-0.01	1.00	-0.01	0.08	0.03	0.00	0.04	-0.04	-0.02	-0.35	-0.01
0.08	-0.01	1.00	-0.01	0.08	0.03	0.00	0.04	-0.04	-0.35	-0.01
0.03	0.08	-1.00	1.00	-0.01	-0.01	0.03	0.00	0.04	-0.35	-0.01
0.06	0.03	0.08	-0.01	1.00	-0.01	0.00	0.03	0.00	0.04	-0.35
0.04	0.00	0.03	0.00	0.01	1.00	-0.01	-0.01	0.03	0.00	0.04
-0.04	-0.04	0.04	0.00	0.08	-1.00	0.01	-0.01	-0.08	0.00	0.03
-0.02	-0.04	0.04	0.00	0.03	0.08	-1.00	0.01	-0.08	0.00	0.03
-0.03	-0.02	-0.04	-0.04	0.00	0.03	0.00	-0.01	1.00	-0.01	-0.03
-0.05	-0.03	-0.02	-0.04	0.00	0.03	0.00	-0.01	-0.01	1.00	-0.03
-0.12	-0.05	-0.03	-0.02	-0.04	0.00	0.04	0.00	0.03	-0.01	1.00
-0.02	-0.12	-0.03	-0.05	-0.02	-0.04	0.00	0.04	0.00	0.03	-0.01
-0.04	-0.02	-0.03	-0.12	-0.05	-0.02	0.00	0.04	0.00	0.03	-0.01
-0.14	-0.04	-0.02	-0.02	-0.05	-0.02	-0.03	-0.04	-0.00	0.00	0.03
-0.09	-0.04	-0.04	-0.02	-0.02	-0.03	-0.04	-0.00	0.00	0.00	0.03
-0.04	-0.09	-0.14	-0.04	-0.02	-0.03	-0.04	-0.00	0.00	0.00	0.03
0.00	0.00	-0.04	-0.09	-0.14	-0.04	-0.02	-0.03	-0.04	-0.00	0.00
0.00	0.00	0.00	-0.04	-0.14	-0.04	-0.02	-0.03	-0.04	-0.00	0.00
-0.02	0.00	0.05	0.00	0.04	0.09	0.14	0.02	0.03	0.04	0.00
-0.05	0.02	0.05	0.00	0.04	0.09	0.14	0.02	0.03	0.04	0.00

NOTE: The final 9 columns of these matrices are provided on the next page.

D =	0.11456284E+00										
1.23	0.05	-0.05	-0.02	-0.01	-0.03	-0.03	0.01	0.06	0.48	0.10	0.01
-0.05	1.23	0.04	-0.05	-0.06	-0.06	-0.03	-0.01	-0.03	-0.01	0.47	0.01
-0.02	-0.04	1.23	0.04	0.05	0.06	0.03	0.01	0.03	0.01	-0.47	0.01
-0.01	-0.05	-0.04	1.23	0.05	0.06	0.03	0.01	0.03	0.01	-0.47	0.01
0.03	0.01	0.05	0.04	1.23	0.06	0.03	0.01	0.03	0.01	-0.47	0.01
0.06	0.03	0.06	0.03	0.01	1.23	0.04	0.01	0.03	0.01	-0.47	0.01
0.48	0.01	0.03	0.01	0.03	0.01	1.23	0.04	0.01	0.03	-0.47	0.01
0.10	0.01	0.03	0.01	0.03	0.01	0.03	1.23	0.04	0.01	-0.47	0.01
0.11	0.01	0.03	0.01	0.03	0.01	0.03	0.01	1.23	0.04	-0.47	0.01
-0.05	-0.10	0.48	0.07	-0.07	-0.03	-0.04	0.02	-0.03	0.00	0.12	1.33
0.05	-0.07	0.16	0.48	0.07	0.06	0.04	-0.04	-0.01	-0.10	0.00	-0.11
0.13	0.05	-0.08	0.08	0.09	0.09	0.03	0.00	0.00	-0.04	-0.08	-0.00
-0.10	-0.13	0.05	-0.08	0.09	0.09	0.04	0.00	0.00	-0.04	-0.08	-0.00
0.06	-0.10	-0.13	0.05	-0.07	-0.08	0.09	0.00	0.00	0.41	0.06	-0.03
0.19	0.06	-0.11	-0.13	0.07	0.08	0.09	0.00	0.00	0.41	0.06	-0.03
0.11	0.06	-0.11	-0.13	0.07	0.08	0.09	0.00	0.00	0.41	0.06	-0.03

	V WEIGHTS AT + AND - LAGS	FOR NOIFF = 1
-10	0.0003	0.0006
-11	0.0004	0.0003
-12	0.0008	0.0008
-13	0.0005	0.0005
-14	-0.0021	0.0037
-15	-0.0039	0.0008
-16	-0.0036	-0.0001
-17	-0.0054	-0.0010
-18	-0.0055	0.0015
-19	-0.0009	0.0007
-10	0.0011	0.0022
-11	-0.0004	0.0014
-12	-0.0009	0.0000
-13	-0.0023	0.0001
-14	-0.0021	0.0005
-15	-0.0015	0.0011
-16	-0.0001	0.0003
-17	-0.0004	0.0019
-18	-0.0001	0.0033
-19	-0.0006	0.0009

APPROX. 95 PER CENT CONF LIMIT ON IMPULSE RESPONSE = 0.039
 1 1 1

STOP

Table C-2

Main Program Used For Estimation of the Noise Model
With Output for Model A-1

```

PROGRAM MAIN
C
C
COMMON X(141),Y(141)
EXTERNAL MODELA,MODELB,MODEL C,MODEL D
DIMENSION SCRAT(1000),TH(8),A(14,14)
READ 3, EPS1, EPS2, FLAM, FNU, MIT, NPROB, NOB, NP
READ 1, ((A(I,J), I=1,7), J=1, NOB)
READ 2, ((A(I,J), I=8,14), J=1, NOB)

C
C
X DESIGNATES THE INFLUENT SERIES AND Y THE EFFLUENT SERIES
C
C
AX=0.0
AY=0.0
DO 5 I=1,NOB
X(I)=A(1,I)+A(7,I)*6.552
Y(I)=A(14,I)
AX=AX+X(I)
AY=AY+Y(I)
5 CONTINUE
PRINT 4
DO 9 I=1,NOB
9 PRINT 7,X(I),Y(I)

C
C
SUBTRACTING THE MEAN FROM THE SERIES
C
C
BX=AX/FLOAT(NOB)
BY=AY/FLOAT(NOB)
DO 6 J=1,NOB
X(J)=X(J)-BX
Y(J)=Y(J)-BY
6 CONTINUE
CALL DIFF(NOB)
NP=3
TH(1)=.083
TH(2)=-.0668
TH(3)=-.0655
CALL TSHAUS(NPROB,MODELA,NOB,NP,TH,EPS1,EPS2,MIT,FLAM,FNU,SCRAT)
NPROB=2
NP=2
TH(1)=.083
TH(2)=1.08
CALL TSHAUS(NPROB,MODELB,NOB,NP,TH,EPS1,EPS2,MIT,FLAM,FNU,SCRAT)
NPROB=3
NP=3
TH(1)=.083
TH(2)=-.0267
TH(3)=0.7309
CALL TSHAUS(NPROB,MODEL C,NOB,NP,TH,EPS1,EPS2,MIT,FLAM,FNU,SCRAT)
NPROB=4
NP=4
TH(1)=0.063
TH(2)=-.0458
TH(3)=.0617
TH(4)=0.569
CALL TSHAUS(NPROB,MODEL D,NOB,NP,TH,EPS1,EPS2,MIT,FLAM,FNU,SCRAT)
1 FORMAT(26X,F5.3,10X,F4.0,1X,F3.0,1X,F5.1)
2 FORMAT(26X,F5.3,21X,F3.0,1X,F3.0,1X,F5.1)
3 FORMAT(4F10.3,4I+)
4 FORMAT(1H1.19X,18H TKN LOAD - GH/DAY,15X,20H EFFLUENT TKN - PPH.
*//)
7 FORMAT(23X,E12.4,20X,E12.4)
STOP
END

```

USASI FORTRAN DIAGNOSTIC RESULTS FOR MAIN

NO ERRORS


```

LN 0001      SUBROUTINE DIFF(NO8)
LN 0002      COMMON X(141),Y(141)
LN 0003      NO8=NO8-1
LN 0004      DO 1 II=1,NO8
LN 0005      X(II)=X(II+1)-X(II)
LN 0006      1  Y(II)=Y(II+1)-Y(II)
LN 0007      RETURN
LN 0008      END

```

USASI FORTRAN DIAGNOSTIC RESULTS FOR DIFF

NO ERRORS

```

LN 0001      SUBROUTINE MODELA(NPROB,TH,A,NO8,NP)
LN 0002      COMMON X(141),Y(141)
LN 0003      DIMENSION A(1),TH(1)
LN 0004      A(1)=0.
LN 0005      A(2)=0.
LN 0006      A(3)=0.
LN 0007      DO 1 I=4,NO8
LN 0008      1  A(I)=Y(I)-TH(1)*X(I-1)+TH(2)*X(I-2)+TH(3)*X(I-3)
LN 0009      RETURN
LN 0010      END

```

USASI FORTRAN DIAGNOSTIC RESULTS FOR MODELA

NO ERRORS

```

LN 0001      SUBROUTINE MODELB(NPROB,TH,A,NO8,NP)
LN 0002      COMMON X(141),Y(141)
LN 0003      DIMENSION A(1),TH(1)
LN 0004      A(1)=0.0
LN 0005      DO 1 I=2,NO8
LN 0006      1  A(I) = Y(I)-TH(1)*X(I-1)-TH(2)*Y(I-1)+TH(2)*A(I-1)
LN 0007      RETURN
LN 0008      END

```

USASI FORTRAN DIAGNOSTIC RESULTS FOR MODELB

NO ERRORS

```

SUBROUTINE MODELC(NPROB,TH,A,NOB,NP)
COMMON X(1:1),Y(1:1)
DIMENSION A(1),TH(1)
A(1)=0.
A(2)=0.
DO 1 I=3,NOB
1  A(I)=Y(I)-TH(1)*X(I-1)+TH(2)*X(I-2)-TH(3)*Y(I-1)+TH(3)*A(I-
RETURN
END

```

USASI FORTRAN DIAGNOSTIC RESULTS FOR MODELC

NO ERRORS

```

SUBROUTINE MODELC0(NPROB,TH,A,NOB,NP)
COMMON X(1:1),Y(1:1)
DIMENSION A(1),TH(1)
A(1)=0.
A(2)=0.
A(3)=0.
DO 1 I=4,NOB
1  A(I)=Y(I)-TH(1)*X(I-1)+TH(2)*X(I-2)+TH(3)*X(I-3)-TH(4)*Y(I-1)+TH(
2)*A(I-1)
RETURN
END

```

USASI FORTRAN DIAGNOSTIC RESULTS FOR MODELC0

NO ERRORS

NON-LINEAR ESTIMATION, PROBLEM NUMBER 3

120 OBSERVATIONS, 3 PARAMETERS

504 SCRATCH REQUIRED

INITIAL PARAMETER VALUES

 $0.8300^1E-01 \quad -0.2670^2E-01 \quad 0.7390^3E+00$
INITIAL SUM OF SQUARES = $0.4295E+03$ DETERMINANT = $0.2354E+00$ ITERATION NO. 1
ANGLE IN SCALED COORD. = 40.58 DEGREES

TEST POINT PARAMETER VALUES

 $0.8000E-01 \quad -0.2987E-01 \quad 0.6059E+00$
TEST POINT SUM OF SQUARES = $0.3320E+03$

PARAMETER VALUES VIA REGRESSION

 $0.8000^1E-01 \quad -0.2987^2E-01 \quad 0.6059^3E+00$
LAMBDA = $0.100E-02$ SUM OF SQUARES AFTER REGRESSION
= 332.04DETERMINANT = $0.2563E+00$ ITERATION NO. 2
ANGLE IN SCALED COORD. = 32.50 DEGREES

TEST POINT PARAMETER VALUES

 $0.8296E-01 \quad -0.4295E-01 \quad 0.5121E+00$
TEST POINT SUM OF SQUARES = $0.3248E+03$

PARAMETER VALUES VIA REGRESSION

 $0.8296^1E-01 \quad -0.4295^2E-01 \quad 0.5121^3E+00$
LAMBDA = $0.100E-03$ SUM OF SQUARES AFTER REGRESSION
= 324.76DETERMINANT = $0.2889E+00$ ITERATION NO. 3
ANGLE IN SCALED COORD. = 48.88 DEGREES

TEST POINT PARAMETER VALUES

 $0.8356E-01 \quad -0.3998E-01 \quad 0.5328E+00$
TEST POINT SUM OF SQUARES = $0.3245E+03$

PARAMETER VALUES VIA REGRESSION

 $0.8356^1E-01 \quad -0.3998^2E-01 \quad 0.5328^3E+00$
LAMBDA = $0.100E-04$ SUM OF SQUARES AFTER REGRESSION
= 324.52DETERMINANT = $0.2791E+00$ ITERATION NO. 4
ANGLE IN SCALED COORD. = 36.32 DEGREES

TEST POINT SUM OF SQUARES = 0.3245E+03

PARAMETER VALUES VIA REGRESSION

0.8356E-01 -0.4114E-01 0.5252E+00

LAMBDA = 0.100E-05

SUM OF SQUARES AFTER REGRESSION
=324.50

DETERMINANT = 0.2825E+00 ITERATION NO. 5
ANGLE IN SCALED COORD. =34.45DEGREES

TEST POINT PARAMETER VALUES
0.8357E-01 -0.4074E-01 0.5278E+00

TEST POINT SUM OF SQUARES = 0.3245E+03

PARAMETER VALUES VIA REGRESSION

0.8357E-01 -0.4074E-01 0.5278E+00

LAMBDA = 0.100E-06

SUM OF SQUARES AFTER REGRESSION
=324.49

DETERMINANT = 0.2813E+00 ITERATION NO. 6
ANGLE IN SCALED COORD. =34.51DEGREES

TEST POINT PARAMETER VALUES
0.8357E-01 -0.4088E-01 0.5269E+00

TEST POINT SUM OF SQUARES = 0.3245E+03
DETERMINANT = 0.2813E+00 ANGLE IN SCALED COORD. =34.51DEGREES

TEST POINT PARAMETER VALUES
0.8357E-01 -0.4088E-01 0.5269E+00

TEST POINT SUM OF SQUARES = 0.3245E+03
DETERMINANT = 0.2813E+00 ANGLE IN SCALED COORD. =34.51DEGREES

TEST POINT PARAMETER VALUES
0.8357E-01 -0.4088E-01 0.5269E+00

TEST POINT SUM OF SQUARES = 0.3245E+03
DETERMINANT = 0.2813E+00 ANGLE IN SCALED COORD. =34.51DEGREES

TEST POINT PARAMETER VALUES
0.8357E-01 -0.4088E-01 0.5269E+00

TEST POINT SUM OF SQUARES = 0.3245E+03
DETERMINANT = 0.2815E+00 ANGLE IN SCALED COORD. =34.50DEGREES

TEST POINT PARAMETER VALUES
0.8357E-01 -0.4088E-01 0.5269E+00

TEST POINT SUM OF SQUARES = 0.3245E+03
DETERMINANT = 0.2832E+00 ANGLE IN SCALED COORD. =34.47DEGREES

TEST POINT PARAMETER VALUES
0.8357E-01 -0.4088E-01 0.5269E+00

TEST POINT SUM OF SQUARES = 0.3245E+03
DETERMINANT = 0.3008E+00 ANGLE IN SCALED COORD. =34.13DEGREES

TEST POINT PARAMETER VALUES
0.8357E-01 -0.4087E-01 0.5269E+00

TEST POINT SUM OF SQUARES = 0.3245E+03
DETERMINANT = 0.5042E+00 ANGLE IN SCALED COORD. =31.17DEGREES

TEST POINT PARAMETER VALUES

DETERMINANT = 0.6201E+01 ANGLE IN SCALED COORD. = 18.48 DEGREES

TEST POINT PARAMETER VALUES
0.8358E-01 -0.4075E-01 0.5276E+00

TEST POINT SUM OF SQUARES = 0.3245E+03

TEST POINT PARAMETER VALUES
0.8357E-01 -0.4074E-01 0.5277E+00

TEST POINT SUM OF SQUARES = 0.3245E+03

PARAMETER VALUES VIA REGRESSION

0.8357¹E-01 -0.4074²E-01 0.5277³E+00

LAMBDA = 0.100E+01

SUM OF SQUARES AFTER REGRESSION
= 324.49

ITERATION STOPS - RELATIVE CHANGE IN SUM OF SQUARES LESS THAN 0.1000E-06

FINAL RESIDUAL VALUES

0.0000E+00	0.0000E+00	-0.9757E+00	-0.1950E+00	-0.2529E+00	0.3109E+00
-0.4412E+01	0.7320E-01	0.7930E+00	0.2337E+01	-0.7069E+00	0.4851E+00
-0.1364E+01	-0.5930E+00	0.-111E+00	0.8544E+00	-0.1536E+01	-0.6637E+00
-0.3164E+00	-0.1425E+00	0.9713E+00	0.1664E+01	0.1229E+01	0.9301E+00
-0.1182E+01	0.2185E+01	0.1892E+01	-0.6668E+00	0.5185E-01	-0.4360E+00
-0.1639E+00	0.1289E+01	0.1337E+00	0.1371E-01	-0.1799E+01	0.6636E+00
-0.2004E+00	-0.6856E+00	-0.8866E+00	-0.9322E-01	-0.1648E+00	-0.1112E+00
-0.4866E+01	0.2516E+01	-0.2884E+01	-0.3403E+01	-0.1036E+01	0.1337E+00
0.1037E+01	0.1086E+01	0.3297E+00	0.9734E+00	0.1193E+00	-0.4179E+00
-0.2483E+01	-0.3068E+01	0.4384E+00	-0.1452E+01	0.8177E+00	0.1488E+00
-0.7263E+00	-0.2741E+01	-0.1382E+01	-0.6878E+00	0.1742E+01	0.4875E+00
0.1307E+01	-0.2997E+00	0.5413E+00	0.2967E+00	-0.1066E+01	0.8265E-00

NOTE: The final 4 columns of this matrix are given on page C-31

CORRELATION MATRIX

	1	2	3
1	1.0000		
2	0.3948	1.0000	
3	0.0068	0.7107	1.0000

NORMALIZING ELEMENTS

$0.5360E-02$	$0.7620E-02$	$0.3581E-01$
--------------	--------------	--------------

VARIANCE OF RESIDUALS = $0.2773E+01$, 117 DEGREES OF FREEDOM

INDIVIDUAL CONFIDENCE LIMITS FOR EACH PARAMETER (ON LINEAR HYPOTHESIS)

1	2	3
$0.1014E+00$	$-0.1536E-01$	$0.6469E+00$
$0.6572E-01$	$-0.6612E-01$	$0.4084E+00$

0.1335E+00	-0.2267E+00	-0.4021E+00	-0.1393E+01
0.7081E+00	-0.2389E+01	-0.1154E+01	-0.8547E+00
0.4192E+00	0.1039E+01	0.5996E+00	-0.1232E+00
0.1007E+01	0.3091E+00	0.3467E-01	0.1513E+00
-0.9074E+00	0.4044E+00	-0.1136E+00	-0.6164E+00
0.1771E+01	0.3865E+01	0.9081E+00	0.2623E+00
-0.1316E+01	0.6644E+00	-0.4926E+00	0.3082E+00
0.9299E+00	0.1714E+01	0.2477E+00	-0.9831E+00
-0.6843E+00	-0.1415E+01	0.8849E+00	0.1218E+01
0.1006E+01	-0.1571E+01	-0.5119E+00	0.6080E+01
0.4656E+01	0.3126E+01	0.7525E-01	0.1984E+00
0.1022E+01	0.7365E+00	0.1676E+01	0.1245E+01

AUTO AND PARTIAL CORRELATIONS OF THE RESIDUALS		
I	AUTO	PARTIAL
1	0.219	0.219
2	0.000	0.000
3	0.000	0.000
4	0.000	0.000
5	0.000	0.000
6	0.000	0.000
7	0.000	0.000
8	0.000	0.000
9	0.000	0.000
10	0.000	0.000
11	0.000	0.000
12	0.000	0.000
13	0.000	0.000
14	0.000	0.000
15	0.000	0.000
16	0.000	0.000
17	0.000	0.000
18	0.000	0.000
19	0.000	0.000
20	0.000	0.000
21	0.000	0.000
22	0.000	0.000
23	0.000	0.000
24	0.000	0.000
25	0.000	0.000
26	0.000	0.000
27	0.000	0.000
28	0.000	0.000
29	0.000	0.000
30	0.108	0.049

APPROX. 95 PERCENT CONF. LIMIT ON CORRELATIONS = 0.183
 CHI-SQUARED STATISTIC = 26.46
 BASED ON (30 - NO. OF STOCHASTIC PARAMETERS) DEGREES OF FREEDOM


```

LN 0007 CALL TSHAUS(NPROB,MODELCD,NOB,NP,TH,EPS1,EPS2,MIT,FLA1,FNU,SC
LN 0008 NZERO=1
LN 0009 IG=5*NP+1+NZERO
LN 0010 NOB=NO3-NZERO
LN 0011 CALL CROSS(X(NZERO),SCRAT(IG),NOB,20)
LN 0012 NOB=NO3+NZERO
LN 0013 NPROB=11
LN 0014 NP=5
LN 0015 TH(1)=.00253
LN 0016 TH(2)=.00254
LN 0017 TH(3)=.00256
LN 0018 TH(4)=.2
LN 0019 TH(5)=.25
LN 0020 CALL TSHAUS(NPROB,MODELAA,NOB,NP,TH,EPS1,EPS2,MIT,FLA1,FNU,SC
LN 0021 NZERO=5
LN 0022 IG=5*NP+1+NZERO
LN 0023 NOB=NO3-NZERO
LN 0024 CALL CROSS(X(NZERO),SCRAT(IG),NOB,20)
LN 0025 NOB=NO3+NZERO
LN 0026 NPROB=21
LN 0027 NP=2
LN 0028 TH(1)=.1905
LN 0029 TH(2)=1.06
LN 0030 CALL TSHAUS(NPROB,MODELBB,NOB,NP,TH,EPS1,EPS2,MIT,FLA1,FNU,SC
LN 0031 NZERO=2
LN 0032 IG=5*NP+1+NZERO
LN 0033 NOB=NO3-NZERO
LN 0034 CALL CROSS(X(NZERO),SCRAT(IG),NOB,20)
LN 0035 NOB=NO3+NZERO
LN 0036 1 FORMAT(25X,F5.3,19X,F4.0,1X,F3.0,1X,4F3.1)
LN 0037 2 FORMAT(25X,F5.3,21X,F3.0,1X,F3.0,1X,4F3.1)
LN 0038 3 FORMAT(4F10.3,4I4)
LN 0039 4 FORMAT(41.13X,134 TKY LOA) - G4/DAY,15X,204 EFFLUENT TKN - P
LN 0040 */)
LN 0041 7 FORMAT(23X,E12.4,20X,E12.4)
LN 0042 STOP
LN 0043 END

```

USASI FORTRAN DIAGNOSTIC RESULTS FOR MAIN

NO ERRORS

ANSI FORTRAN(2.3)/MASTER INTEGER WORD SIZE = 1 , * OPTION IS OFF , 0 OPT

```

LN 0001      SUBROUTINE DIFF(NOB)
LN 0002      COMMON X(141),Y(141)
LN 0003      NOB=NOB-1
LN 0004      DO 1 II=1,NOB
LN 0005      X(II)=X(II+1)-X(II)
LN 0006      1 Y(II)=Y(II+1)-Y(II)
LN 0007      RETURN
LN 0008      END

```

USASI FORTRAN DIAGNOSTIC RESULTS FOR DIFF

NO ERRORS

ANSI FORTRAN(2.3)/MASTER INTEGER WORD SIZE = 1 , * OPTION IS OFF , 0 OPT

```

LN 0001      SUBROUTINE MODELA(NPROB,TH,A,NOB,NP)
LN 0002      COMMON X(141),Y(141)
LN 0003      DIMENSION A(1),TH(1)
LN 0004      A(1)=0.
LN 0005      A(2)=0.
LN 0006      A(3)=0.
LN 0007      A(4)=0.
LN 0008      DO 1 I=5,NOB
LN 0009      1 A(I)=Y(I)-TH(1)*X(I-1)+TH(2)*X(I-2)+TH(3)*X(I-3)
LN 0010      2-TH(4)*Y(I-1)
LN 0011      3 +TH(1)*TH(4)*X(I-2)-TH(2)*TH(4)*X(I-3)-TH(3)*TH(4)*X(I-4)
LN 0012      RETURN
LN 0013      END

```

USASI FORTRAN DIAGNOSTIC RESULTS FOR MODELA

NO ERRORS

ANSI FORTRAN(2.3)/MASTER INTEGER WORD SIZE = 1 , * OPTION IS OFF , 0 OPT

```

LN 0001      SUBROUTINE MODELB(NPROB,TH,A,NOB,NP)
LN 0002      COMMON X(141),Y(141)
LN 0003      DIMENSION A(1),TH(1)
LN 0004      A(1)=0.0
LN 0005      A(2)=0.
LN 0006      DO 1 I=3,NOB
LN 0007      1 A(I) = Y(I)-TH(1)*X(I-1)-TH(2)*Y(I-1)+TH(2)*A(I-1)
LN 0008      2 -TH(3)*Y(I-1)+TH(3)*TH(2)*Y(I-2)
LN 0009      3 +TH(1)*TH(3)*X(I-2)
LN 0010      RETURN
LN 0011      END

```

USASI FORTRAN DIAGNOSTIC RESULTS FOR MODELB

NO ERRORS

RTRAN(2,3)/MASTER INTEGER WORD SIZE = 1 , * OPTION IS OFF , 0 OPTION IS

```

01      SUBROUTINE MODELCD(NPROB,TH,A,NOB,NP)
02      COMMON X(141),Y(141)
03      DIMENSION A(1),TH(1)
04      A(1)=C.
05      A(2)=C.
06      A(3)=C.
07      A(4)=C.
08      DO 1 I=5,NOB
09      1  A(I)=Y(I)-TH(1)*X(I-1)+TH(2)*X(I-2)+TH(3)*X(I-3)-TH(4)*Y(I-1)+TH(4
10      2)*A(I-1)
11      3 -TH(5)*Y(I-1)+TH(5)*TH(4)*Y(I-2)
12      4 +TH(1)*TH(5)*X(I-2)-TH(2)*TH(5)*X(I-3)-TH(3)*TH(5)*X(I-4)
13      RETURN
14      END

```

USASI FORTRAN DIAGNOSTIC RESULTS FOR MODELCD

NO ERRORS

RTRAN(2,3)/MASTER INTEGER WORD SIZE = 1 , * OPTION IS OFF , 0 OPTION IS

```

1      SUBROUTINE MODELCD(NPROB,TH,A,NOB,NP)
2      COMMON X(141),Y(141)
3      DIMENSION A(1),TH(1)
4      A(1)=C.
5      A(2)=C.
6      A(3)=C.
7      DO 1 I=4,NOB
8      1  A(I)=Y(I)-TH(1)*X(I-1)+TH(2)*X(I-2)-TH(3)*Y(I-1)+TH(3)*A(I-1)
9      2 -TH(4)*Y(I-1)+TH(4)*TH(3)*Y(I-2)
10     3 +TH(1)*TH(4)*X(I-2)-TH(2)*TH(4)*X(I-3)
11     RETURN
12     END

```

USASI FORTRAN DIAGNOSTIC RESULTS FOR MODELCD

NO ERRORS

FORTRAN(2.3)/MASTER INTEGER WORD SIZE = 1 , * OPTION IS OFF , 0 OPTION IS

```

0001      SUBROUTINE MODELAA(NPROB,TH,A,NOB,NP)
0002      COMMON X(141),Y(141)
0003      DIMENSION A(1),TH(1)
0004      A(1)=0.
0005      A(2)=0.
0006      A(3)=0.
0007      A(4)=0.
0008      A(5)=0.
0009      DO 1 I=6,NOB
0010      1  A(I)=Y(I)-TH(1)*X(I-1)+TH(2)*X(I-2)+TH(3)*X(I-3)-TH(4)*Y(I-1)-TH
0011      *Y(I-2)
0012      2  +TH(1)*TH(4)*X(I-2)-TH(2)*TH(4)*X(I-3)-TH(3)*TH(4)*X(I-4)
0013      3  +TH(1)*TH(5)*X(I-3)-TH(2)*TH(5)*X(I-4)-TH(3)*TH(5)*X(I-5)
0014      RETURN
0015      END

```

USASI FORTRAN DIAGNOSTIC RESULTS FOR MODELAA

NO ERRORS

FORTRAN(2.3)/MASTER INTEGER WORD SIZE = 1 , * OPTION IS OFF , 0 OPTION IS

```

0001      SUBROUTINE MODELBB(NPROB,TH,A,NOB,NP)
0002      COMMON X(141),Y(141)
0003      DIMENSION A(1),TH(1)
0004      A(1)=0.
0005      A(2)=0.
0006      DO 1 I=3,NOB
0007      1  A(I) = Y(I)-TH(1)*X(I-1)-TH(2)*Y(I-1)+TH(2)*A(I-1)
0008      2  +A(I-1)-TH(2)*A(I-2)
0009      RETURN
0010      END

```

USASI FORTRAN DIAGNOSTIC RESULTS FOR MODELBB

NO ERRORS

00.6295	00.7600
00.6283	00.1030
00.6183	00.1330
00.5763	00.1410
00.5931	00.1510
00.5763	00.1520
00.5400	00.1470
00.4812	00.1476
00.1590	00.1260
00.2405	00.1010
00.3394	00.8900
00.2589	00.6300
00.3384	00.2930
00.3675	00.3000
00.2143	00.2700
00.1745	00.2700
00.4364	00.2800
00.4868	00.5100
00.5539	00.5700
00.6183	00.7100
00.4393	00.1700
00.3693	00.1600
00.3497	00.1180
00.4945	00.9200
00.5508	00.1210
00.5479	00.1770
00.5422	00.2280
00.5488	00.2610
00.5498	00.2630
00.5421	00.2560
00.5337	00.2790
00.5242	00.2750
00.4945	00.2790
00.5337	00.2550
00.5993	00.2670
00.5973	00.2395
00.5838	00.2317
00.5822	00.3310

120 OBSERVATIONS, 4 PARAMETERS 636 SCRATCH REQUIRED

INITIAL PARAMETER VALUES

$0.8357E-01$ $-0.4074E-01$ $0.5277E+00$ $0.2190E+00$

INITIAL SUM OF SQUARES = $0.3076E+03$

ITERATION NO. 1
DETERMINANT = $0.3975E+00$ ANGLE IN SCALED COORD. = 18.40 DEGREES

TEST POINT PARAMETER VALUES
 $0.8298E-01$ $-0.3341E-01$ $0.5684E+00$ $0.2235E+00$

TEST POINT SUM OF SQUARES = $0.3068E+03$

PARAMETER VALUES VIA REGRESSION

$0.8298E-01$ $-0.3341E-01$ $0.5684E+00$ $0.2235E+00$

LAMBDA = $0.100E-02$ SUM OF SQUARES
After Regression = 306.7

ITERATION NO. 2
DETERMINANT = $0.3725E+00$ ANGLE IN SCALED COORD. = 38.75 DEGREES

TEST POINT PARAMETER VALUES
 $0.8316E-01$ $-0.3529E-01$ $0.5535E+00$ $0.2365E+00$

TEST POINT SUM OF SQUARES = $0.3067E+03$

PARAMETER VALUES VIA REGRESSION

$0.8316E-01$ $-0.3529E-01$ $0.5535E+00$ $0.2365E+00$

LAMBDA = $0.100E-03$ SUM OF SQUARES
After Regression = 306.6

ITERATION NO. 3
DETERMINANT = $0.3873E+00$ ANGLE IN SCALED COORD. = 32.57 DEGREES

TEST POINT PARAMETER VALUES
 $0.8310E-01$ $-0.3424E-01$ $0.5610E+00$ $0.2327E+00$

TEST POINT SUM OF SQUARES = $0.3067E+03$

PARAMETER VALUES VIA REGRESSION

$0.8310E-01$ $-0.3424E-01$ $0.5610E+00$ $0.2327E+00$

LAMBDA = $0.100E-04$ SUM OF SQUARES
After Regression = 306.65

ITERATION NO. 4
DETERMINANT = $0.3811E+00$ ANGLE IN SCALED COORD. = 35.82 DEGREES

TEST POINT PARAMETER VALUES
 $0.8314E-01$ $-0.3471E-01$ $0.5574E+00$ $0.2348E+00$

TEST POINT SUM OF SQUARES = 0.3067E+03

PARAMETER VALUES VIA REGRESSION

0.8314E-01 -0.3471E-01 0.5574E+00 0.2348E+00

LAMBDA = 0.100E-05

SUM OF SQUARE
After Regression = 306.65

ITERATION NO. 5
DETERMINANT = 0.3841E+00 ANGLE IN SCALED COORD. = 34.96 DEGREES

TEST POINT PARAMETER VALUES
0.8312E-01 -0.3449E-01 0.5591E+00 0.2338E+00

TEST POINT SUM OF SQUARES = 0.3066E+03

PARAMETER VALUES VIA REGRESSION

0.8312E-01 -0.3449E-01 0.5591E+00 0.2338E+00

LAMBDA = 0.100E-06

SUM OF SQUARE
After Regression = 306.65

ITERATION NO. 6
DETERMINANT = 0.3827E+00 ANGLE IN SCALED COORD. = 35.27 DEGREES

TEST POINT PARAMETER VALUES
0.8313E-01 -0.3460E-01 0.5583E+00 0.2343E+00

TEST POINT SUM OF SQUARES = 0.3066E+03
DETERMINANT = 0.3827E+00 ANGLE IN SCALED COORD. = 35.27 DEGREES

TEST POINT PARAMETER VALUES
0.8313E-01 -0.3460E-01 0.5583E+00 0.2343E+00

TEST POINT SUM OF SQUARES = 0.3066E+03
DETERMINANT = 0.3827E+00 ANGLE IN SCALED COORD. = 35.27 DEGREES

TEST POINT PARAMETER VALUES
0.8313E-01 -0.3460E-01 0.5583E+00 0.2343E+00

TEST POINT SUM OF SQUARES = 0.3066E+03
DETERMINANT = 0.3827E+00 ANGLE IN SCALED COORD. = 35.27 DEGREES

TEST POINT PARAMETER VALUES
0.8313E-01 -0.3460E-01 0.5583E+00 0.2343E+00

TEST POINT SUM OF SQUARES = 0.3066E+03
DETERMINANT = 0.3830E+00 ANGLE IN SCALED COORD. = 35.26 DEGREES

TEST POINT PARAMETER VALUES
0.8313E-01 -0.3460E-01 0.5583E+00 0.2343E+00

TEST POINT SUM OF SQUARES = 0.3066E+03
DETERMINANT = 0.3853E+00 ANGLE IN SCALED COORD. = 35.23 DEGREES

TEST POINT PARAMETER VALUES
0.8313E-01 -0.3459E-01 0.5583E+00 0.2343E+00

TEST POINT SUM OF SQUARES = 0.3066E+03
DETERMINANT = 0.4096E+00 ANGLE IN SCALED COORD. = 34.95 DEGREES

TEST POINT PARAMETER VALUES
0.8313E-01 -0.3459E-01 0.5583E+00 0.2343E+00

TEST POINT SUM OF SQUARES = 0.3066E+03
DETERMINANT = 0.6960E+00 ANGLE IN SCALED COORD. = 31.52 DEGREES

TEST POINT PARAMETER VALUES

DETERMINANT = 0.1315E+02 ANGLE IN SCALED COORD. = 16.41DEGREES

TEST POINT PARAMETER VALUES
0.8313E-01 -0.3450E-01 0.5589E+00 0.2340E+00

TEST POINT SUM OF SQUARES = 0.3066E+03

TEST POINT PARAMETER VALUES
0.8312E-01 -0.3450E-01 0.5590E+00 0.2339E+00

TEST POINT SUM OF SQUARES = 0.3066E+03

PARAMETER VALUES VIA REGRESSION

0.8312¹E-01 -0.3450²E-01 0.5590³E+00 0.2339⁴E+00

LAMBDA = 0.100E+01

SUM OF SQUARES
After Regression = 306.65

ITERATION STOPS - RELATIVE CHANGE IN SUM OF SQUARES LESS THAN 0.1000E-0

FINAL RESIDUAL VALUES

0.0000E+00	0.0000E+00	0.0000E+00	0.4975E+00	0.9057E-01	0.9914E+0
-0.3983E+01	0.1206E+01	0.8634E+00	0.2213E+01	-0.1209E+01	0.6822E+0
-0.1151E+01	-0.2882E+00	0.5131E+00	0.6554E+00	-0.1737E+01	-0.2273E+0
-0.2649E+00	-0.6238E-01	0.9922E+00	0.1387E+01	0.8607E+00	0.6677E+0
-0.1173E+01	0.2804E+01	0.1242E+01	-0.1180E+01	0.1311E+00	-0.4974E+0
-0.9157E-01	0.1276E+01	-0.1701E+00	0.1063E-02	-0.1755E+01	0.1043E+0
-0.1510E+00	-0.5668E+00	-0.6809E+00	0.1522E+00	-0.9219E-01	-0.7992E+0
-0.4973E+01	0.3514E+01	-0.3615E+01	-0.2877E+01	-0.2672E+00	0.1612E+0
0.1323E+01	0.9181E+00	0.5514E-01	0.8709E+00	-0.1454E+00	-0.4494E+0
-0.2673E+01	-0.2404E+01	0.1152E+01	-0.1503E+01	0.1167E+01	0.1232E+0
-0.2640E+01	-0.2685E+01	-0.7779E+00	-0.3533E+00	0.1979E+01	0.4288E+0
0.1254E+01	-0.6135E+00	0.6060E+00	0.1667E+00	-0.1144E+01	0.3357E+0

NOTE: The final 4 columns of this matrix are on the next page.

CORRELATION MATRIX

	1	2	3	4
1	1.0000			
2	0.1944	1.0000		
3	-0.1021	0.7028	1.0000	
4	-0.1447	0.0054	0.0115	1.0000

NORMALIZING ELEMENTS

1	2	3	4
0.5346E-02	0.7461E-02	0.4199E-01	0.5560E-01

VARIANCE OF RESIDUALS = 0.2644E+01, 116 DEGREES OF FREEDOM

INDIVIDUAL CONFIDENCE LIMITS FOR EACH PARAMETER (ON LINEAR HYPOTHESIS)

0.1005E+00	-0.1024E-01	0.6955E+00	0.4147E+00
0.6574E-01	-0.5876E-01	0.4225E+00	0.5312E-01

AUTO AND PARTIAL CORRELATIONS OF THE RESIDUALS

I	AUTO	PARTIAL
1	0.018	0.018
2	-0.080	-0.080
3	-0.050	-0.048
4	-0.029	-0.034
5	-0.067	-0.075
6	0.066	0.061
7	0.119	0.124
8	0.033	0.035
9	0.112	0.095
10	0.075	0.085
11	0.099	0.099
12	0.021	0.029
13	-0.101	-0.112
14	0.006	0.003
15	0.136	0.115
16	0.031	0.011
17	0.059	0.023
18	0.098	0.047
19	0.023	0.031
20	0.025	0.054
21	0.042	0.042
22	0.033	0.037
23	0.055	0.110
24	0.092	0.112
25	0.017	0.030
26	0.117	0.126
27	0.076	0.047
28	0.140	0.147
29	0.073	0.102
30	0.155	0.092

APPROX. 95 PERCENT CONF. LIMIT ON CORRELATIONS = 0.183

CHI-SQUARED STATISTIC = 22.73
 BASED ON (30 - NO. OF STOCHASTIC PARAMETERS) DEGREES OF FREEDOM

CROSS-CORRELATIONS BETWEEN MANIPULATED VARIABLES AND RESIDUALS

Variable	Correlation	Variable	Correlation
1	0.037	1	0.027
2	0.038	2	0.020
3	0.033	3	0.047
4	0.045	4	0.047
5	0.037	5	0.030
6	0.176	6	0.170
7	0.014	7	0.022
8	0.008	8	0.012
9	0.048	9	0.033
10	0.058	10	0.056
11	0.115	11	0.077
12	0.022	12	0.049
13	0.011	13	0.070
14	0.036	14	0.011
15	0.141	15	0.044
16	0.024	16	0.004
17	0.024	17	0.004
18	0.024	18	0.004
19	0.024	19	0.004
20	0.024	20	0.004
21	0.024	21	0.004
22	0.024	22	0.004
23	0.024	23	0.004
24	0.024	24	0.004
25	0.024	25	0.004
26	0.024	26	0.004
27	0.024	27	0.004
28	0.024	28	0.004
29	0.024	29	0.004
30	0.024	30	0.004

APPROX. 95 PERCENT CONF. LIMIT ON CROSS-CORRELATIONS = 0.185

STANDARD DEVIATIONS S(X) = 0.1723E+02 S(A) = 0.1614E+01

CHI-SQUARED STATISTIC = 21.33 BASED ON 21 DEGREES OF FREEDOM

Table C-4
 Subroutine TSHAUS

C-45

```

SUBROUTINE TSHAUS(NPROB,MODEL,NOB,NP,TH,EPS1,EPS2,MIT,FLAM,FNU,
1 SCRAT)
C RESIDUALS ARE RETURNED IN SCRAT(IG) WHERE IG=5*NP+1
  DIMENSION SCRAT(1)
  IA=1
  IB=IA+NP
  IC=IB+NP
  ID=IC+NP
  IE=ID+NP
  IG = IE+NP
  IH=IG+NOB
  II = IH + NP * NOB
  IJ = IH
  CALL HAUSTS(NPROB,MODEL,NOB, NP,TH, EPS1,EPS2,MIT
1,FLAM,FNU, SCRAT(IA), SCRAT(IB), SCRAT(IC), SCRAT(ID),
2 SCRAT(IE), SCRAT(IG), SCRAT(IH), SCRAT(II),SCRAT(IJ))
  CALL IDENT(SCRAT(IG),NOB,20,2)
  RETURN
  END
SUBROUTINE HAUSTS(NPRBO, MODEL, NBO, NQ,TH, EP1S,EP2S,
1MIT,FLAM,FNU, Q,P,E,PHI,TB, R,A,D,DELZ)
C FORTRAN II VERSION
C ADAPTED FOR THE CDC 6400 (J. F. MAGGREGOR 9/72)
C DIMENSION TH(NQ), R(NBO)
C DIMENSION Q(NQ), P(NQ), E(NQ), PHI(NQ), TB(NQ)
C DIMENSION A(NQ,NQ), D(NQ,NQ), DELZ(NBO,NQ)
  DIMENSION TH(1), Q(1), P(1), E(1),
1 PHI(1), TB(1), R(1), A(1), D(1), DELZ(1)
  DIMENSION AC(31),PP(31)
  ACOS(X) = ATAN(SQRT(1.0/X**2 - 1.0))
  NP = NQ
  NPROB = NPRBO
  NOB = NBO
  EPS1 = EP1S
  EPS2 = EP2S
  NPSQ = NP * NP
  NSCRAC = 5*NP+NPSQ + NOB+NP*NOB
  PRINT 1000, NPROB, NOB, NP, NSCRAC
  PRINT 1001
  CALL GASS60(1, NP, TH, TEMP, TMEP)
  IF(MINO(NP-1,50-NP,NOB-NP,MIT-1,999-MIT))99,15,15
15 IF(FNU-1.0)99, 99, 16
16 CONTINUE
  DO 19 I=1,NP
  IF( ABS(TH(I)) ) 99,99,19
19 CONTINUE
  GA = FLAM
  NIT = 1
  IF(EPS1) 5,70,70
5 EPS1 = 0
70 SSQ = 0
  CALL MODEL(NPROB, TH, R, NOB, NP)
  DO 90 I = 1, NOB
90 SSQ=SSQ+R(I)*R(I)
  PRINT 1003, SSQ

```

```

100 GA = GA / FNU
    INTCNT = 0
    PRINT 1004, NIT
101 JS = 1 - NCB
    DO 130 J=1, NP
    TEMP = TH(J)
    P(J) = 0.01*TH(J)
    TH(J) = TH(J) + P(J)
    Q(J) = 0
    JS = JS + NOB
    CALL MODEL(NPROB, TH, DELZ(JS), NOB, NP)
    IJ = JS - 1
    DO 120 I = 1, NOB
    IJ = IJ + 1
    DELZ(IJ) = R(I) - DELZ(IJ)
120 Q(J) = Q(J) + DELZ(IJ) * R(I)
    Q(J) = Q(J) / P(J)
C                                     Q=XT*R (STEEPEST DESCENT)
130 TH(J) = TEMP
131 DO 150 I = 1, NP
    DO 151 J=1, I
    SUM = 0
    KJ = NOB*(J-1)
    KI = NOB*(I-1)
    DO 160 K = 1, NOB
    KI = KI + 1
    KJ = KJ + 1
160 SUM = SUM + DELZ(KI) * DELZ(KJ)
    TEMP = SUM / (P(I)*P(J))
    JI = J + NP*(I-1)
    D(JI) = TEMP
    IJ = I + NP*(J-1)
151 D(IJ) = TEMP
150 E(I) = SQRT(D(JI))
666 CONTINUE
    DO 153 I = 1, NP
    IJ = I - NP
    DO 153 J=1, I
    IJ = IJ + NP
    A(IJ) = D(IJ) / (E(I)*E(J))
    JI = J + NP*(I-1)
153 A(JI) = A(IJ)
C                                     A = SCALED MOMENT MATRIX
    II = - NP
    DO 155 I=1, NP
    P(I) = Q(I) / E(I)
    PHI(I) = P(I)
    II = NP + 1 + II
155 A(II) = A(II) + GA
C
    I=1
    CALL MATIN(A, NP, P, I, DET)
C                                     P/E = CORRECTION VECTOR
    STEP=1.0
    SUM1=0.
    SUM2=0.
    SUM3=0.
    DO 231 I=1, NP
    SUM1 = P(I)*PHI(I) + SUM1

```

```

SUM2=P(I)*P(I)+SUM2
SUM3= PHI(I) * PHI(I) + SUM3
231 PHI(I) = P(I)
    TEMP = SUM1/SQRT(SUM2*SUM3)
    TEMP = AMIN1(TEMP, 1.0)
    TEMP = 57.295*ACOS(TEMP)
    PRINT 1041, DET, TEMP
170 DO 220 I = 1, NP
    P(I) = PHI(I) *STEP / E(I)
    TB(I) = TH(I) + P(I)
220 CONTINUE
    PRINT 7000
7000 FORMAT(30H0TEST POINT PARAMETER VALUES      )
    PRINT 2006, (TB(I), I = 1, NP)
    SUMB=0
    CALL MODEL(NPROB, TB, R, NOB, NP)
    DO 230 I=1,NOB
230  SUMB=SUMB+R(I)*R(I)
    PRINT 1043, SUMB
    IF(SUMB - (1.0+EPS1)*SSQ) 662, 662, 663
663  IF(-AMIN1(TEMP-30.0, GA)) 665, 665, 664
665  STEP=STEP/2.0
    INTCNT = INTCNT + 1
    IF(INTCNT - 36) 170, 2700, 2700
664  GA=GA*FNU
    INTCNT = INTCNT + 1
    IF(INTCNT - 36) 666, 2700, 2700
662  PRINT 1007
    DO 669 I=1,NP
669  TH(I)=TB(I)
    CALL GASS60(1, NP, TH, TEMP, TEMP)
    PRINT 1040, GA, SUMB
    IF-(EPS2) 229,229,225
229  IF (EPS1) 270,270,265
225  DO 240 I = 1, NP
    IF(ABS(P(I))/(1.E-20+ABS(TH(I)))-EPS2) 240, 240, 241
241  IF (EPS1) 270,270,265
240  CONTINUE
    PRINT 1009, EPS2
    GO TO 280
265  IF(ABS(SUMB - SSQ) - EPS1*SSQ) 266, 266, 270
266  PRINT 1010, EPS1
    GO TO 280
270  SSQ=SUMB
    NIT=NIT+1
    IF(NIT - MIT) 100, 100, 280
2700 PRINT 2710
2710 FORMAT(/115H0**** THE SUM OF SQUARES CANNOT BE REDUCED TO THE SUM
10F SQUARES AT THE END OF THE LAST ITERATION - ITERATING STOPS /)
C
C
C
280  PRINT 1011
    PRINT 2001, (R(I), I = 1, NOB)
    SSQ=SUMB
    IDF=NOB-NP
    PRINT 1015
    I=0
    CALL MATIN(D, NP, P, I, DET)

```



```

DO 7692 I=1, NP
II = I + NP*(I-1)
7692 E(I) = SQRT(D(II))
DO 340 I=1, NP
JI = I + NP*(I-1) - 1
IJ = I + NP*(I-2)
DO 340 J = I, NP
JI = JI + 1
A(JI) = D(JI) / (E(I)*E(J))
IJ = IJ + NP
340 A(IJ) = A(JI)
CALL GASS60(3, NP, TEMP, TEMP, A)
PRINT 1016
CALL GASS60(1, NP, E, TEMP, TEMP)
IF(IDF) 341, 410, 341
341 SDEV = SSQ / IDF
PRINT 1014, SDEV, IDF
SDEV = SQRT(SDEV)
DO 391 I=1, NP
P(I) = TH(I) + 2.0 * E(I) * SDEV
391 TB(I) = TH(I) - 2.0 * E(I) * SDEV
PRINT 1039
CALL GASS60(2, NP, TB, P, TEMP)
CALL ACORR(R, AC, SDF, NOB, 30)
CALL PARTAL(AC, PP, 30)
PRINT 59
59 FORMAT(11H1, 47H AUTO AND PARTIAL CORRELATIONS OF THE RESIDUALS, //
19X, 1HI, 14X, 4HAUTO, 14X, 7HPARTIAL //)
DO 58 I=1, 30
58 PRINT 57, I, AC(I), PP(I)
57 FORMAT (8X, I2, 13X, F5.3, 13X, F6.3)
CL = 2.0 / SQRT(FLCAT(NOB))
PRINT 54, CL
54 FORMAT(//50H APPROX. 95 PERCENT CONF. LIMIT ON CORRELATIONS = ,F5.
13)
CHI = 0.
DO 56 I=1, 30
56 CHI = CHI + AC(I) * AC(I)
CHI = CHI * FLOAT(NOB)
PRINT 55, CHI
55 FORMAT(/25H CHI-SQUARED STATISTIC = ,F6.2/65H BASED ON (30 - NO. 0
IF STOCHASTIC PARAMETERS) DEGREES OF FREEDOM /)
410 CONTINUE
RETURN
99 PRINT 1034
GO TO 410
10000 FORMAT(38H1NON-LINEAR ESTIMATION, PROBLEM NUMBER I3, // I5,
1 14H OBSERVATIONS, I5, 11H PARAMETERS I14, 17H SCRATCH REQUIRED)
1001 FORMAT(/25H0INITIAL PARAMETER VALUES )
1003 FORMAT(/25H0INITIAL SUM OF SQUARES = E12.4)
1004 FORMAT(/////45X, 13HITERATION NO. I4)
1007 FORMAT(/32H0PARAMETER VALUES VIA REGRESSION )
10090 FORMAT(/62H0ITERATION STOPS - RELATIVE CHANGE IN EACH PARAMETER LE
1SS THAN E12.4)
10100 FORMAT(/62H0ITERATION STOPS - RELATIVE CHANGE IN SUM OF SQUARES LE
1SS THAN E12.4)
1011 FORMAT(22H1FINAL RESIDUAL VALUES )
1012 FORMAT(/////10H0RESIDUALS )
1014 FORMAT(/24H0VARIANCE OF RESIDUALS = ,E12.4, 1H, I4,

```

```

123H DEGREES OF FREEDOM )
1015 FORMAT(////19H0CORRELATION MATRIX )
1016 FORMAT(////21H0NORMALIZING ELEMENTS )
1034 FORMAT(/16H0PARAMETER ERROR )
10390FORMAT(/71H0INDIVIDUAL CONFIDENCE LIMITS FOR EACH PARAMETER (ON LI
  INEAR HYPOTHESIS) )
10400FORMAT(/9H0LAMBDA =E10.3,40X,33HSUM OF SQUARES AFTER REGRESSION =
  1E15.7)
1041 FORMAT(14H DETERMINANT = E12.4, 6X, 25H ANGLE IN SCALED COORD. =
  1 F5.2, 8HDEGREES )
1043 FORMAT(28H0TEST POINT SUM OF SQUARES = E12.4)
2001 FORMAT(/10E12.4)
2006 FORMAT(10E12.4)
END
SUBROUTINE MATIN(A, NVAR, B, NB, DET)
  DIMENSION A(NVAR, 1), B(NVAR, 1)
  COMMON/GASPAR/DUMIES(7), PIVOTM
  PIVOTM = A(1,1)
  DET = 1.0
  DO 550 ICOL = 1, NVAR
    PIVOT = A(ICOL, ICOL)
    PIVOTM = AMIN1(PIVOT, PIVOTM)
    DET = PIVOT * DET
C
C   DIVIDE PIVCT ROW BY PIVOT ELEMENT
C
    A(ICOL, ICOL) = 1.0

    PIVOT = AMAX1(PIVOT, 1.E-20)
    PIVOT = A(ICOL, ICOL)/PIVOT
    DO 350 L=1,NVAR
350   A(ICOL, L) = A(ICOL, L)*PIVOT
    IF(NB .EQ. 0) GO TO 371
    DO 370 L=1,NB
370   B(ICOL, L) = B(ICOL, L)*PIVOT
C
C   REDUCE NON-PIVOT ROWS
C
371   DO 550 L1=1,NVAR
    IF(L1 .EQ. ICOL) GO TO 550
    T = A(L1, ICOL)
    A(L1, ICOL) = 0.
    DO 450 L=1,NVAR
450   A(L1, L) = A(L1, L) - A(ICOL, L)*T
    IF(NB .EQ. 0) GO TO 550
    DO 500 L=1,NB
500   B(L1, L) = B(L1, L) - B(ICOL, L)*T
550   CONTINUE
    RETURN
  END
SUBROUTINE GASS60(ITYPE, NQ, A, B, C)
  DIMENSION A(NQ),B(NQ),C(NQ,NQ)
  NP = NQ
  NR = NP/10
  LOW = 1
  LUP = 10
10   IF( NR )15,20,30
15   RETURN
20   LUP=NP

```



```

IF (LOW .GT. LUP) RETURN
30 PRINT 500, (J, J=LOW, LUP)
GO TO (40, 60, 80), ITYPE
40 PRINT 600, (A(J), J=LOW, LUP)
GO TO 100
60 PRINT 600, (B(J), J=LOW, LUP)
GO TO 40
80 DO 90 I=LOW, LUP
90 PRINT 720, I, (C(J, I), J=LOW, I)
LOW2=LUP+1
IF (LOW2 .GT. NP) GO TO 100
DO 95 I=LOW2, NP
95 PRINT 720, I, (C(J, I), J=LOW, LUP)
100 LOW = LOW + 10
LUP = LUP + 10
NR = NR - 1
GO TO 10
500 FORMAT(/I8, 9I12)
600 FORMAT(10E12.4)
720 FORMAT(1H0, I3, 1X, F7.4, 9F12.4)
1 CONTINUE
RETURN
END
SUBROUTINE CROSS(X, A, NOB, NL)
DIMENSION X(NOB), A(NOB)
DIMENSION CC1(41), CC2(41)
CALL CRCORR(X, A, CC2, SDX, SDA, NOB, NL)
CALL CRCORR(A, X, CC1, SDA, SDX, NOB, NL)
PRINT 6
6 FORMAT (1H1, 10X, 78H CROSS-CORRELATIONS BETWEEN MANIPULATED VARIABLE
1ES AND RESIDUALS X(T)*A(T+K), //)
NL1 = NL+1
DO 7 K=1, NL1
KK = K-1
K1 = -KK
7 PRINT 8, K1, CC1(K), KK, CC2(K)
8 FORMAT (5X, I3, 5X, F6.3, 10X, I3, 5X, F6.3)
CL = 2.0/SQRT(FLOAT(NOB))
PRINT 12, CL
12 FORMAT(/56H APPROX. 95 PERCENT CONF. LIMIT ON CROSS-CORRELATIONS
1= ,F6.3)
PRINT 9, SDX, SDA
9 FORMAT (/28H STANDARD DEVIATIONS S(X) =,E12.4, 5X, 6HS(A) =,E12.4
1)
Q = 0.
DO 10 J=1, 11
10 Q = Q+CC1(J)*CC1(J)+CC2(J)*CC2(J)
Q = (Q-CC1(1)*CC1(1))*FLOAT(NOB)
NDF = 21
PRINT 11, Q, NDF
11 FORMAT (/25H CHI-SQUARED STATISTIC = ,F6.2, 10H BASED ON ,I2, 20H DE
1GREES OF FREEDOM /)
RETURN
END
SUBROUTINE CRCORR(X, Y, CC, SDX, SDY, N, NL)
DIMENSION X(N), Y(N), CC(1)
SX = 0.
SY = 0.
SXX = 0.

```



```

SYY = 0.
DO 2 I=1,N
SX = SX+X(I)
SY = SY+Y(I)
SXX = SXX+X(I)*X(I)
2 SYY = SYY+Y(I)*Y(I)
TN = N
SDX = SQRT((SXX-SX*SX/TN)/TN)
SDY = SQRT((SYY-SY*SY/TN)/TN)
NL1 = NL+1
DO 3 K=1,NL1
SXY = 0.
NN = N-K+1
DO 4 I=1,NN
KK = I+K-1
4 SXY = SXY+(X(I)-SX/TN)*(Y(KK)-SY/TN)
3 CC(K) = (SXY/TN)/(SDX*SDY)
RETURN
END
SUBROUTINE IDENT(Z,NOB,NL,NDIFF)
DIMENSION Z(1),AC(60),PP(60)
DIMENSION W(400)
NDATA = NOB
ND = 0
NZ=NL+1
DO 10 I=1,NOB
10 W(I)=Z(I)
PRINT 30
30 FORMAT(1H1,10X,49H AUTO AND PARTIAL CORRELATIONS OF ORIGINAL SERIE
1S,/)
GO TO 20
41 PRINT 31
31 FORMAT(1H1,10X,61H AUTO AND PARTIAL CORRELATIONS OF FIRST DIFFEREN
1CES OF SERIES,/)
GO TO 20
42 PRINT 32
32 FORMAT(1H1,10X,62H AUTO AND PARTIAL CORRELATIONS OF SECOND DIFFERE
1NCES OF SERIES,/)
20 PRINT 33
33 FORMAT(9X,1HI,14X,4HAUTO,14X,7HPARTIAL,/)
CALL ACORR(W,AC,SOZ,NOATA,NL)
113 FORMAT(1H1,4HTEST)
CALL PARTAL(AC,PP,NL)
DO 58 I=1,NL
58 PRINT 57, I,AC(I),PP(I)
57 FORMAT (8X,I2,13X,F6.3,13X,F6.3)
PRINT 59, AC(NZ)
59 FORMAT(//22H MEAN OF THE SERIES = ,E12.5)
CL = 2.0/SQRT(FLCAT(NDATA))
PRINT 54,CL
54 FORMAT(//50H APPROX. 95 PERCENT CONF. LIMIT ON CORRELATIONS = ,F5.
13)
PRINT 60, SOZ
60 FORMAT(//31H STANDARD DEVIATION OF SERIES =,E14.7)
CHI = 0.
DO 56 I=1,NL
56 CHI = CHI+AC(I)*AC(I)
CHI = CHI*FLOAT(NDATA)
PRINT 55, CHI,NL

```

```

55 FORMAT(/25H CHI-SQUARED STATISTIC = ,F6.2,10H BASED ON ,I2,19H DEG
1RESS OF FREEDOM,/)
  NDATA = NDATA-1
  ND = ND+1
  IF(ND.GT.NDIFF) GO TO 100
  DO 40 I=1,NDATA
40 W(I)=W(I+1)-W(I)
  GO TO (41,42), ND
100 RETURN
  END
  SUBROUTINE ACORR(Z,AC,SDZ,N,NL)
  DIMENSION Z(1),AC(1)
  NL1 = NL+1
  TN = N
  SZ = 0.
  DO 13 I=1,N
13 SZ = SZ+Z(I)
  ZBAR = SZ/TN
  DO 10 JJ=1,NL1
  J = JJ-1
  SZZ = 0.
  NN = N-J
  DO 11 I=1,NN
  K=I+J
11 SZZ=SZZ+(Z(I)-ZBAR)*(Z(K)-ZBAR)
10 AC(JJ) = SZZ/TN
  SDZ = SQRT(AC(1))
  VZ = AC(1)
  DO 12 J=1,NL
12 AC(J) = AC(J+1)/VZ
  RETURN
  END
  SUBROUTINE PARTAL(R,PAUTO,M)
  DIMENSION R(1),PAUTO(1),PHAT(40),PHATN(40)
  PAUTO(1) = R(1)
  PHAT(1) = R(1)*(1.-R(2))/(1.-R(1)**2)
  PHAT(2) = (R(2)-R(1)**2)/(1.-R(1)**2)
  PAUTO(2) = PHAT(2)
  DO 4 I=3,M
  L = I-1
  FNUM = 0.
  DENOM = 0.
  DO 1 J=1,L
  K = I-J
  FNUM = PHAT(J)*R(K)+FNUM
1 DENOM = DENOM+PHAT(J)*R(J)
  PHATN(I) = (R(I)-FNUM)/(1.-DENOM)
  PAUTO(I) = PHATN(I)
  DO 2 J=1,L
  K = I-J
2 PHATN(J) = PHAT(J)-PHATN(I)*PHAT(K)
  DO 3 J=1,I
3 PHAT(J) = PHATN(J)
4 CONTINUE
  RETURN
  END

```


Forecast Program Used To Predict Effluent Response

```

PROGRAM TST (INPUT,OUTPUT,TAPES=INPUT,TAPE6=OUTPUT)
DIMENSION TITLX(2),TITLY(2),DATA(1000),A(14,141)
DIMENSION C1(141),C2(141),B1(141),B2(141)
DIMENSION X(141),Y(141),FS(141)
REAL N(141)
DIMENSION TH(10),TIME(141)
COMMON/AAB/NTY,XORG
COMMON/AAA/XMIN,XMAX,V,NTX,XSCALE,NPLTS
DATA NS/07,NF/141/,YORG/0.00/,NTOT/02/,NTX/12/,NOB/141/
NTOT=3
XSCALE=10.
NTX=13
NOB=121
NPT=0
CALL INIT(NTOT)
READ (5,10) ((A(I,J),I=1,7),J=1,NOB)
READ (5,10) ((A(I,J),I=8,14),J=1,NOB)
READ (5,12) YSCALE,NTY,TITLX(1),TITLX(2),TITLY(1),TITLY(2)
IFLAG=0
NPT=NPT+1
DO 15 I=1,NOB
Y(I)=A(14,I)
X(I)=A(7,I)*A(1,I)*6.552
TIME(I)=I-1
15 CONTINUE
NOB=NOB-6
CALL PLDTA(IFLAG,NPT,NS,NOB,YSCALE,YORG,TITLX,TITLY,Y(5),TIME(5))
PRINT(6,30)
PRINT(6,31)
IFLAG=1
NPT=NPT+1
FS(1)=0.
FS(2)=0.
FS(3)=0.
FS(4)=0.
FS(5)=0.
N(5)=0.
TH(5)=0.2203
TH(4)=0.3994
TH(3)=-0.02815
TH(2)=-0.046
TH(1)=0.0811
16 CONTINUE
NOB=NOB+6
DO 83 I=6,NOB
FS(I)=(TH(4)+TH(5))*Y(I-1)-TH(4)*TH(5)*Y(I-2)+TH(1)*X(I-1)-TH(2)*
2X(I-2)-TH(3)*X(I-3)-TH(5)*(TH(1)*X(I-2)-TH(2)*X(I-3)-TH(3)*X(I-4))
3-TH(4)*N(I-1)
4+Y(I-1)-TH(4)*Y(I-2)-TH(5)*Y(I-2)+TH(4)*TH(5)*Y(I-3)
5-TH(1)*X(I-2)+TH(1)*TH(5)*X(I-3)+TH(3)*X(I-4)+TH(2)*X(I-3)-TH(2)*
6TH(5)*X(I-4)-TH(3)*TH(5)*X(I-5)
N(I)=Y(I)-FS(I)
83 CONTINUE
10 FORMAT(26X,F5.3,19X,F4.0,1X,F3.0,1X,4F5.1)
11 FORMAT(26X,F5.3,21X,F3.0,1X,F3.0,1X,4F5.1)
12 FORMAT(F5.1,I5,4A10)
30 FORMAT(1H1,48H EFFLUENT TKN INFLUENT TKN LOAD FORECAST TKN/)
31 FORMAT(52H MG/L GM/DAY MG/L ///)
32 FORMAT(F10.2,3X,F10.2,1X,F10.2,4X,F10.2)
33 FORMAT(1H1,15H AVERAGE VALUES,/,F10.2,3X,F10.2,1X,F10.2,4X,F10.2)
34 FORMAT(3F15.2)
37 FORMAT(1H1,15H AVERAGE VALUES,/,F10.2,5X,F10.2)
DO 35 I=1,NOB
35 TIME(I)=I-1
NOB=NOB-6
CALL PLDTA(IFLAG,NPT,NS,NOB,YSCALE,YORG,TITLX,TITLY,FS(5),TIME(5))
NPT=NPT+1
NOB=NOB+6
DO 36 I=1,NOB
36 TIME(I)=I-1
IFLAG=0
READ (5,12) YSCALE,NTY,TITLX(1),TITLX(2),TITLY(1),TITLY(2)
CALL PLDTA(IFLAG,NPT,NS,NOB,YSCALE,YORG,TITLX,TITLY,X,TIME)
STOP
END

```



```

SUBROUTINE INIT(NTOT)

```

```

COMMON/AAB/NTY,XORG

```

```

COMMON/AAA/XMIN,XMAX,V,NTX,XSCALE,NPLTS

```

```

THIS SUBROUTINE INITIALIZES THE PLOT ROUTINE

```

```

A MAXIMUM OF 1000 POINTS MAY BE PLOTTED PER CALL TO PLDTA
THE CALL TO THIS SUBROUTINE MUST BE THE FIRST EXECUTABLE STATEMENT
IN THE MAIN PROGRAM
AS OF OCTOBER 20 1976 THE CONTROL CARDS NECESSARY TO PLOT ON
THE BENSEN-LEHNER ARE

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```

ACCN,MT1.

```

```

ATTACH,PLOTLIB,ID=GPAK.

```

```

OTHER CONTROL CARDS

```

```

FTN.

```

```

LDSET(LIB=PLOTLIB)

```

```

LGO.

```

```

NIX - NUMBER OF TIC MARKS ON X AXIS

```

```

EVERY TIC MARK IS 10 MM FROM THE NEXT AND IS ANNOTATED

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```

XSCALE IS THE INCREMENT IN THE X DIRECTION

```

```

IE THE TIC MARKS ARE / 10 20 ETC IF XSCALE S 10.

```

```

NPLTS - NUMBER OF PLOTS DESIRED INCLUDED ANY PLOTS TO BE SUPERIMP-

```

```

USED
INITIALIZE PLOT ROUTINE TO CENTIMETERS

```

```

XORG=0.0

```

```

XL=XORG+NIX*XSCALE

```

```

XMAX=1.1*XL

```

```

V=XORG-10*XSCALE

```

```

XMIN=V

```

```

NPLTS=NTOT

```

```

RETURN

```

```

END

```

```

SUBROUTINE PLDTA(IFLAG,NPT,NS,NOB,YSCALE,YORG,TITLX,TITLY,DATA,TIM
*E)
DIMENSION TITLX(2),TITLY(2),FMT(5)
COMMON/AAB/NTY,XORG
COMMON/AAA/XMIN,XMAX,V,NTX,XSCALE,NPLTS
DIMENSION Z(1000),TIME(1000),DATA(1000),DIST(1000),SLOPE(1000)
DATA FMT/$(F5.2)$,$$(F5.2)$,$$(F4.1)$,$$(F4.1)$,$$(F5.0)$#/

C
C   NPT - NUMBER OF CURRENT DATA SET
C   IFLAG - IF NON ZERO THEN CURRENT PLOT SUPERIMPOSED ON LAST GRID
C   NS - THE FIRST ABSCISSA VALUE IE 0 OR 10 OR 15
C   NOB - NUMBER OF DATA POINTS TO BE PLOTTED
C   YSCALE - THE INCREMENT PER TIC MARK ALONG THE ORDINATE AXIS
C   YORG - ORIGIN OF THE PLOT IN DATA UNITS. THEREFORE THE VALUES ALONG
C   THE ORDINATE WILL BE      YORG      YORG+YSCALE      YORG+2*YSCALE . .
C   YORG+5*YSCALE
C   TITLX - A 20 CHARACTER TITLE TO BE PRINTED ON THE ABSCISSA
C   TITLY - A 20 CHARACTER TITLE TO BE PRINTED ON THE ORDINATE
C   DATA - A VECTOR CONTAINING THE ORDINATE VALUES TO BE PLOTTED
C   THE FIRST ELEMENT CORRESPONDS TO THE NS TH DATA POINT
C   NTY - NO. OF TICS IN Y DIRECTION
C
WRITE(6,6001)NPT,NOB,YORG,YSCALE
6001  FORMAT(1H1,///,20X,$PLOT NUMBER$,I2,///,20X,$NUMBER OBS $I4,
1/,20X,$ORIGIN OF ORDINATE AXIS$,F10.6/,20X,$INCR ORDINATE AXIS$,
2F10.6)
WRITE(6,6002)
6002  FORMAT(1H0,20X,$DATA FOR PLOTTING$)
WRITE(6,6003)(DATA(L),L=1,NOB)
6003  FORMAT(1H ,7E10.4)
C   IF(NPT.NE.1)GO TO 20
C   INITIALIZE PLOT BOUNDARIES ON FIRST PLOT ONLY
CALL PLOT(0.,0.,-20)
CALL PLOT(0.,0.,3)
GO TO 30
20  CONTINUE
IF(IFLAG.NE.0)GO TO 10
C   INCREMENT NUMBER OF INDIVIDUAL GRIDS
CALL PLOT(DXL+10.,0.,-3)
30  CONTINUE
XNTY=NTY
W=YORG-(XNTY-1.)*YSCALE/2.
YMIN=W
YL=YORG+YSCALE*(XNTY-1.)
YMAX=YL*1.1
CALL PLTIN(XSCALE,YSCALE,V,W,XMIN,XMAX,YMIN,YMAX)
C   PLACE GRID ON X AND Y AXIS
C   DETERMINE DATA ORIGIN IN PLOTTER UNITS
CALL UNITTO(XORG,YORG,DX,DY)
CALL PLOT(DX,DY,3)
C   DETERMINE END POINTS OF AXIS IN PLOTTER UNITS
DXL=DX+NTX
DYL=DY+NTY
CALL PLOT(DXL,DY,2)
C   PLACE TIC MARKS ALONG AXIS
Y=DY-.1

```

```

DO 39 I=1,NTX
X=DX+I
CALL MATH(X,Y,.1,90.,2)
C 39 CONTINUE
DRAW Y AXIS
CALL PLOT(DX,DY,3)
CALL PLOT(DX,DYL,2)
C PLACE Y AXIS AT RIGHT HAND EDGE OF X AXIS
CALL PLOT(DXL,DY,3)
CALL PLOT(DXL,DYL,2)
C PLACE LINE ACROSS THE TOP OF GRAPH
CALL PLOT(DX,DYL,3)
CALL PLOT(DXL,DYL,2)
C INSERT TIC MARK AT RIGHT HAND EDGE
XX=DXL
DO 49 I=1,NTY
Y=DY+I
Y=Y-0.05
CALL MATH(XX,Y,.1,0.,2)
C 49 CONTINUE
PLACE TIC MARKS AT LEFT HAND EDGE
X=DX-.1
DO 50 I=1,NTY
Y=DY+I
Y=Y-0.05
CALL MATH(X,Y,.1,0.,2)
C 50 CONTINUE
INSERT SCALE ON X AXIS
PLACE / ON AXIS
Y=DY-.5
CALL SYMBOL(DX,Y,.20,1H0,90.,1)
C INSERT NO ALONG AXIS
Y=DY-1.
DO 59 I=2,NTX,2
X=DX+I
NUM=XSCALE*I
CALL INUMBR(X,Y,.2,NUM,0.,4H(I4))
C 59 CONTINUE
X=DX-1.5
PUT NO ALONG Y AXIS
NTY=NTY+1
DO 69 I=1,NTY
Y=DY+I-1
YVAL=YORG+(I-1)*YSCALE
CALL NUMBER(X,Y,.2,YVAL,0.,6H(F6.1))
C 69 CONTINUE
INSERT TITLE ON X AXIS
X=DX+0.3*NTX
Y=DY-1.5
CALL SYMBOL(X,Y,.25,TITLX,0.,20)
X=DX-1.8
Y=DY+0.5
CALL SYMBOL(X,Y,.25,TITLY,90.,20)
C 10 CONTINUE
PLOT DATA WITHIN GRID
DO 9 J=1,NOB
YD=DATA(J)

```



```

      TD=TIME(J)
C-----CONSTRAIN PEN MOVEMENT-----
      IF(YD.GT.YL)YD=YL
      IF(YD.LT.YORG)YD=YORG
C-----CONVERT TO PLOTTER UNITS-----
      CALL UNITTO(TD,YD,TP,YP)
      TIME(J)=TP
      Z(J)=YP
      9 CONTINUE
C-----PLOT DATA-----
      IF(IFLAG.NE.0)GO TO 78
      DO 597 ML=1,NOB
      ZZ1=TIME(ML)
      ZZ2=Z(ML)
      597 CALL GRAF(ZZ1,ZZ2,.15,1)
      GO TO 79
      78 IF(IFLAG.EQ.1) CALL MPLOT(TIME,Z,NOB)
      IF(IFLAG.EQ.2) CALL GRAF(TIME,Z,.15,1)
C-----CHECK TO SEE IF MAXIMUM NUMBER OF PLOTS HAS BEEN REACHED-----
      79 IF(NPLTS.NE.NPT)RETURN
      CALL PLOT(DXL+20.,0.,3)
      CALL PLOT(X,Y,999)
      RETURN
      END

```

```

SUBROUTINE NUMBER(X,Y,HEIGHT,ANUM,THETA,FMT)
DIMENSION BCD(1)
ENCODE(10,FMT,BCD)ANUM
CALL SYMBOL(X,Y,HEIGHT,BCD,THETA,10)
RETURN
END

```

```

SUBROUTINE INUMBR(X,Y,HEIGHT, NUM,THETA,FMT)
DIMENSION BCD(1)
ENCODE(10,FMT,BCD)NUM
CALL SYMBOL(X,Y,HEIGHT,BCD,THETA,10)
RETURN
END

```

APPENDIX DREACTOR RETENTION TIMES & MIXING CHARACTERISTICS

Dye studies were performed on the 0.5 metre RBC and the 2.0 metre RBC for two reasons:

1. The tracer studies provided an indication of peak retention times and RBC response to influent spikes. This then provided the basic information necessary for choosing discrete time intervals for the dynamic experiments.
2. To identify the mixing characteristics of each RBC, which previously had been shown to approximate two CSTR's in series for a 0.5 metre RBC.

Dye studies were performed with a Turner Model III flowmeter. Samples (200 ml) were obtained at discrete time intervals so as not to miss the peak. Fresh water was used to feed the RBC's except in the case of 2.0 m which was being acclimatized at the same time with raw sewage. The 0.5 metre RBCs did not have a biological growth on them at the time the dye studies were conducted. Calibration curves were determined using dye solutions of known concentration. To initiate each study a slug of dye of known concentration was added to the RBC inlet. The RBC was stopped for approximately 10 seconds for this purpose.

The results of the studies and further details of each study are included in Tables D1, D2, D3, D4 and D5. A computer listing of the program used is presented in Table D6. Flow conditions for both the carbon oxidation and carbon oxidation plus modes were investigated.

A description of the flow models used to develop the computer program is given below.

Tanks in Series Model Theory

The final effluent of a system of j equal sized CSTR's in series can be found from the following equation:

$$\frac{C}{C_0} = \frac{j^j \theta^{j-1} e^{-j\theta}}{(j-1)!} \quad \dots(D-1)$$

where: C = effluent tracer concentration
 θ = dimensionless time
 j = number of tanks
 C_0 = the quantity of tracer added divided by the volume of the entire system.

This applies only to a pulse input of tracer. In this type of system, as j approaches large values (say > 15), the flow regime approximates plug flow whereas, when j is equal to 1, the flow is completely mixed. If the time at which the peak dye concentration occurs is known, the above equation can be solved for j by taking the derivative and equating the result to zero. Theta peak is determined by dividing the peak time by the residence time. The final form of the equation is:

$$j = \frac{1}{1 - \theta} \quad \dots (D-2)$$

Dispersion Model Theory

The dispersion model is developed in such a way that it

assumes plug flow for a given reactor system with the inclusion of a term which describes the degree of molecular dispersion or deviation from the ideal. The general equation of this model is:

$$D \frac{\partial^2 C}{\partial x^2} - u \frac{\partial C}{\partial x} - \frac{\partial C}{\partial t} = 0 \quad \dots(D-3)$$

where: u = mean displacement velocity
 C = concentration
 $\frac{\partial C}{\partial x}$ = concentration gradient
 $\frac{\partial C}{\partial t}$ = reaction term
 D = turbulence expression

The solution of this equation for a tracer pulse input to a closed vessel given by Mjjachi (1953) is quoted by Timpany (1):

$$\frac{C}{C_0} = 2 \sum_{n=1}^{\infty} \frac{U_n (U \sin U_n + \cos U_n)}{(U^2 + 2U + U_n^2)} \text{EXP} \frac{U - (U^2 + U_n^2) \theta}{2U} \quad \dots(D-4)$$

where: $U_n = \text{COT}^{-1} \left(\frac{U_n}{U} - \frac{U}{U_n} \right) / 2$
 $U = \frac{UL}{2D}$
 L = tank length

The value U_n is best calculated by trial and error using an iterative approach. Also, the summation in equation 4 is taken to some reasonably large but finite value for practical purposes.

Instead of determining a value for D by the normal variance technique suggested by Levenspiel (1967), a correlation between peak time and D/UL developed by Timpany (1966) was used. Proper use of the variance method for D/UL calculation generally requires concentration data to be entered for at least seven detention times which is rarely practical. Since the concentration data approached zero after two detentions times data for about three detention times were used.

Tracer Response Analysis, 0.5 Ml, At 0.82 L/min.

TRACER RESPONSE ANALYSIS

Rotating Biological Contactor
HYDRAULIC CHARACTERIZATION

TEST METHOD USING A PULSE INPUT OF RODAMINE WT DYE
REACTOR OPERATION AND TEST CONDITIONS

VOLUME OF REACTOR	=	141.80 LITRES
HYDRAULIC LOADING	=	0.32 LITRES/MIN
THEORETICAL DET	=	173.99 MIN
DYE INJECTION	=	0.0030 LITRES
CONC OF DYE ADDED	=	0.238E+07 PPR
DYE / TANK VOLUME	=	50.35 PPR

TEST RESULTS AND CALCULATED VALUES

DYE PEAK TIME	=	85.00 MIN
PEAK/THEOR DET	=	0.489

PEAK/MEAN DYE RES	=	0.596 MIN
MEAN DYE RESIDENCE	=	142.50 MIN
PER DYE RECOVERY	=	94.314
FR. STAGNANT ZONE	=	0.181

CSTR S IN SERIES USING THEORETICAL RES.	=	1.96
CSTR S IN SERIES USING MEAN DYE RES.	=	2.48
D/UL VALUE USING THEORETICAL RESIDENCE	=	0.3336E+00
D/UL VALUE USING MEAN DYE RESIDENCE	=	0.2282E+00

EXPERIMENTAL RESULTS C/CO VERSUS THETA

THETA	C/CO
0.043	0.216
0.078	0.279
0.120	0.318
0.172	0.347
0.216	0.371
0.259	0.395
0.345	0.447
0.338	0.477
0.431	0.517
0.474	0.537
0.517	0.552
0.560	0.566
0.603	0.572
0.647	0.577
0.690	0.577
0.733	0.572
0.776	0.566
0.819	0.552
0.862	0.537
0.905	0.517
0.948	0.497
0.991	0.477
1.035	0.447
1.078	0.413
1.121	0.385
1.164	0.361
1.207	0.337
1.250	0.318
1.293	0.299
1.437	0.279
1.581	0.247
1.724	0.216
1.868	0.185
2.012	0.154
2.155	0.123
2.299	0.092
2.443	0.061

CALCULATED C/CO VERSUS THETA VALUES
FOR CSTR IN SERIES MODEL

THEORETICAL DETENTION		ACTUAL DETENTION	
THETA	C/CO	THETA	C/CO
0.000	0.000	0.000	0.000
0.005	0.000	0.000	0.000
0.010	0.000	0.000	0.000
0.015	0.000	0.000	0.000
0.020	0.000	0.000	0.000
0.025	0.000	0.000	0.000
0.030	0.000	0.000	0.000
0.035	0.000	0.000	0.000
0.040	0.000	0.000	0.000
0.045	0.000	0.000	0.000
0.050	0.000	0.000	0.000
0.055	0.000	0.000	0.000
0.060	0.000	0.000	0.000
0.065	0.000	0.000	0.000
0.070	0.000	0.000	0.000
0.075	0.000	0.000	0.000
0.080	0.000	0.000	0.000
0.085	0.000	0.000	0.000
0.090	0.000	0.000	0.000
0.095	0.000	0.000	0.000
1.000	0.000	1.000	0.000
1.005	0.000	1.000	0.000
1.010	0.000	1.000	0.000
1.015	0.000	1.000	0.000
1.020	0.000	1.000	0.000
1.025	0.000	1.000	0.000
1.030	0.000	1.000	0.000
1.035	0.000	1.000	0.000
1.040	0.000	1.000	0.000
1.045	0.000	1.000	0.000
1.050	0.000	1.000	0.000
1.055	0.000	1.000	0.000
1.060	0.000	1.000	0.000
1.065	0.000	1.000	0.000
1.070	0.000	1.000	0.000
1.075	0.000	1.000	0.000
1.080	0.000	1.000	0.000
1.085	0.000	1.000	0.000
1.090	0.000	1.000	0.000
1.095	0.000	1.000	0.000
2.000	0.000	2.000	0.000
2.005	0.000	2.000	0.000
2.010	0.000	2.000	0.000
2.015	0.000	2.000	0.000
2.020	0.000	2.000	0.000
2.025	0.000	2.000	0.000
2.030	0.000	2.000	0.000
2.035	0.000	2.000	0.000
2.040	0.000	2.000	0.000
2.045	0.000	2.000	0.000
2.050	0.000	2.000	0.000
2.055	0.000	2.000	0.000
2.060	0.000	2.000	0.000
2.065	0.000	2.000	0.000
2.070	0.000	2.000	0.000
2.075	0.000	2.000	0.000
2.080	0.000	2.000	0.000
2.085	0.000	2.000	0.000
2.090	0.000	2.000	0.000
2.095	0.000	2.000	0.000
3.000	0.000	3.000	0.000
3.005	0.000	3.000	0.000
3.010	0.000	3.000	0.000
3.015	0.000	3.000	0.000
3.020	0.000	3.000	0.000
3.025	0.000	3.000	0.000
3.030	0.000	3.000	0.000
3.035	0.000	3.000	0.000
3.040	0.000	3.000	0.000
3.045	0.000	3.000	0.000
3.050	0.000	3.000	0.000
3.055	0.000	3.000	0.000
3.060	0.000	3.000	0.000
3.065	0.000	3.000	0.000
3.070	0.000	3.000	0.000
3.075	0.000	3.000	0.000
3.080	0.000	3.000	0.000
3.085	0.000	3.000	0.000
3.090	0.000	3.000	0.000
3.095	0.000	3.000	0.000
4.000	0.000	4.000	0.000
4.005	0.000	4.000	0.000
4.010	0.000	4.000	0.000
4.015	0.000	4.000	0.000
4.020	0.000	4.000	0.000
4.025	0.000	4.000	0.000
4.030	0.000	4.000	0.000
4.035	0.000	4.000	0.000
4.040	0.000	4.000	0.000
4.045	0.000	4.000	0.000
4.050	0.000	4.000	0.000
4.055	0.000	4.000	0.000
4.060	0.000	4.000	0.000
4.065	0.000	4.000	0.000
4.070	0.000	4.000	0.000
4.075	0.000	4.000	0.000
4.080	0.000	4.000	0.000
4.085	0.000	4.000	0.000
4.090	0.000	4.000	0.000
4.095	0.000	4.000	0.000

CALCULATED C/CO VERSUS THETA VALUES
FOR DISPERSION MODEL

THEORETICAL DETENTION		ACTUAL DETENTION	
0.1000	0.0233	0.1000	0.001
0.2000	0.0333	0.2000	0.112
0.3000	0.0417	0.3000	0.412
0.4000	0.0484	0.4000	0.749
0.5000	0.0537	0.5000	0.917
0.6000	0.0587	0.6000	0.952
0.7000	0.0637	0.7000	0.929
0.8000	0.0683	0.8000	0.851
0.9000	0.0723	0.9000	0.664
1.0000	0.0758	1.0000	0.572
1.1000	0.0789	1.1000	0.488
1.2000	0.0815	1.2000	0.413
1.3000	0.0837	1.3000	0.349
1.4000	0.0855	1.4000	0.293
1.5000	0.0869	1.5000	0.245
1.6000	0.0879	1.6000	0.205
1.7000	0.0885	1.7000	0.173
1.8000	0.0888	1.8000	0.144
1.9000	0.0889	1.9000	0.120
2.0000	0.0888	2.0000	0.101
2.1000	0.0885	2.1000	0.084
2.2000	0.0880	2.2000	
2.3000	0.0873	2.3000	0.070
2.4000	0.0864	2.4000	0.058
2.5000	0.0853	2.5000	0.049
2.6000	0.0840	2.6000	0.041
2.7000	0.0825	2.7000	0.034
2.8000	0.0808	2.8000	0.028
2.9000	0.0789	2.9000	0.024
3.0000	0.0769	3.0000	0.020

Tracer Response Analysis, 0.5 M2, At 2.33 L/min.

TRACER RESPONSE ANALYSIS

Rotating Biological Contactor
HYDRAULIC CHARACTERIZATION

 TEST METHOD USING A PULSE INPUT OF RHODAMINE WT DYE
 REACTOR OPERATION AND TEST CONDITIONS

VOLUME OF REACTOR	=	131.4 LITRES
HYDRAULIC LOADING	=	2.33 LITRES/MIN
THEORETICAL DET	=	56.39 MIN
DYE INJECTION	=	0.0030 LITRES
CONC OF DYE ADDED	=	4.238E+07 PPB
DYE / TANK VOLUME	=	54.3+PPB

TEST RESULTS AND CALCULATED VALUES

DYE PEAK TIME	=	39.00 MIN
PEAK/THEOR DET	=	0.692
PEAK/MEAN DYE RES	=	0.591 MIN
MEAN DYE RESIDENCE	=	66.00 MIN
PER DYE RECOVERY	=	80.247%
FR. STAGNANT ZONE	=	-0.170

CSTR S IN SERIES USING THEORETICAL RES.	=	3.24
CSTR S IN SERIES USING MEAN DYE RES.	=	2.44
DZUL VALUE USING THEORETICAL RESIDENCE	=	0.1444E+00
DZUL VALUE USING MEAN DYE RESIDENCE	=	0.2344E+00

CALCULATED C/QO VERSUS THETA VALUES
FOR DISPERSION MODEL

THEORETICAL DETENTION		ACTUAL DETENTION	
0.100	0.000	0.100	0.001
0.200	0.013	0.200	0.022
0.300	0.134	0.300	0.457
0.400	0.522	0.400	0.728
0.500	0.821	0.500	0.919
0.600	0.995	0.600	0.952
0.700	1.044	0.700	0.922
0.800	1.004	0.800	0.845
0.900	0.914	0.900	0.733
1.000	0.791	1.000	0.637
1.100	0.683	1.100	0.566
1.200	0.573	1.200	0.493
1.300	0.473	1.300	0.410
1.400	0.337	1.400	0.347
1.500	0.315	1.500	0.292
1.600	0.254	1.600	0.246
1.700	0.204	1.700	0.206
1.800	0.163	1.800	0.173
1.900	0.131	1.900	0.145
2.000	0.104	2.000	0.121
2.100	0.083	2.100	0.101
2.200	0.066	2.200	0.085
2.300	0.052	2.300	0.071
2.400	0.041	2.400	0.059
2.500	0.033	2.500	0.050
2.600	0.026	2.600	0.041
2.700	0.021	2.700	0.035
2.800	0.016	2.800	0.029
2.900	0.013	2.900	0.024
3.000	0.010	3.000	0.020

Tracer Response Analysis, 0.5 M2, At 3.46 L/min,

TRACER RESPONSE ANALYSIS

Rotating Biological Contactor
HYDRAULIC CHARACTERIZATIONTEST METHOD USING A PULSE INPUT OF RODAMINE WT DYE
REACTOR OPERATION AND TEST CONDITIONS

VOLUME OF REACTOR	=	133.50 LITRES
HYDRAULIC LOADING	=	3.46 LITRES/MIN
THEORETICAL DET	=	38.58 MIN
DYE INJECTION	=	0.0030 LITRES
CONC OF DYE ADDED	=	0.233E+07 PPM
DYE / TANK VOLUME	=	23.48 PPM

TEST RESULTS AND CALCULATED VALUES

DYE PEAK TIME	=	22.00 MIN
PEAK/THEOR DET	=	0.570

PEAK/MEAN DYE RES	=	0.629 MIN
MEAN DYE RESIDENCE	=	35.90 MIN
PER DYE RECOVERY	=	71.957%
FR. STAGNANT ZONE	=	0.033

CSTP S IN SERIES USING THEORETICAL RES.	=	2.33
CSTP S IN SERIES USING MEAN DYE RES.	=	2.69
D/UL VALUE USING THEORETICAL RESIDENCE	=	0.2590E+00
D/UL VALUE USING MEAN DYE RESIDENCE	=	0.1356E+00

EXPERIMENTAL RESULTS C/CO VERSUS THETA

THETA	C/CO
0.078	0.000
0.156	0.000
0.233	0.000
0.311	0.000
0.389	0.000
0.467	0.000
0.544	0.000
0.622	0.000
0.700	0.000
0.778	0.000
0.855	0.000
0.933	0.000
1.011	0.000
1.089	0.000
1.166	0.000
1.244	0.000
1.322	0.000
1.400	0.000
1.477	0.000
1.555	0.000
1.633	0.000
1.711	0.000
1.788	0.000
1.866	0.000
1.944	0.000
2.022	0.000
2.100	0.000
2.177	0.000
2.255	0.000
2.333	0.000
2.411	0.000
2.489	0.000
2.567	0.000
2.644	0.000
2.722	0.000
2.800	0.000
2.878	0.000
2.955	0.000
3.033	0.000
3.111	0.000
3.189	0.000
3.267	0.000
3.344	0.000
3.422	0.000
3.500	0.000
3.578	0.000
3.655	0.000
3.733	0.000
3.811	0.000
3.889	0.000
3.967	0.000
4.044	0.000
4.122	0.000
4.200	0.000
4.278	0.000
4.355	0.000
4.433	0.000
4.511	0.000
4.589	0.000
4.667	0.000
4.744	0.000
4.822	0.000
4.900	0.000
4.978	0.000
5.055	0.000
5.133	0.000
5.211	0.000
5.289	0.000
5.367	0.000
5.444	0.000
5.522	0.000
5.600	0.000
5.678	0.000
5.755	0.000
5.833	0.000
5.911	0.000
5.989	0.000
6.067	0.000
6.144	0.000
6.222	0.000
6.300	0.000
6.378	0.000
6.455	0.000
6.533	0.000
6.611	0.000
6.689	0.000
6.767	0.000
6.844	0.000
6.922	0.000
7.000	0.000
7.078	0.000
7.155	0.000
7.233	0.000
7.311	0.000
7.389	0.000
7.467	0.000
7.544	0.000
7.622	0.000
7.700	0.000
7.778	0.000
7.855	0.000
7.933	0.000
8.011	0.000
8.089	0.000
8.167	0.000
8.244	0.000
8.322	0.000
8.400	0.000
8.478	0.000
8.555	0.000
8.633	0.000
8.711	0.000
8.789	0.000
8.867	0.000
8.944	0.000
9.022	0.000
9.100	0.000
9.178	0.000
9.255	0.000
9.333	0.000
9.411	0.000
9.489	0.000
9.567	0.000
9.644	0.000
9.722	0.000
9.800	0.000
9.878	0.000
9.955	0.000
10.033	0.000
10.111	0.000
10.189	0.000
10.267	0.000
10.344	0.000
10.422	0.000
10.500	0.000
10.578	0.000
10.655	0.000
10.733	0.000
10.811	0.000
10.889	0.000
10.967	0.000
11.044	0.000
11.122	0.000
11.200	0.000
11.278	0.000
11.355	0.000
11.433	0.000
11.511	0.000
11.589	0.000
11.667	0.000
11.744	0.000
11.822	0.000
11.900	0.000
11.978	0.000
12.055	0.000
12.133	0.000
12.211	0.000
12.289	0.000
12.367	0.000
12.444	0.000
12.522	0.000
12.600	0.000
12.678	0.000
12.755	0.000
12.833	0.000
12.911	0.000
12.989	0.000
13.067	0.000
13.144	0.000
13.222	0.000
13.300	0.000
13.378	0.000
13.455	0.000
13.533	0.000
13.611	0.000
13.689	0.000
13.767	0.000
13.844	0.000
13.922	0.000
14.000	0.000
14.078	0.000
14.155	0.000
14.233	0.000
14.311	0.000
14.389	0.000
14.467	0.000
14.544	0.000
14.622	0.000
14.700	0.000
14.778	0.000
14.855	0.000
14.933	0.000
15.011	0.000
15.089	0.000
15.167	0.000
15.244	0.000
15.322	0.000
15.400	0.000
15.478	0.000
15.555	0.000
15.633	0.000
15.711	0.000
15.789	0.000
15.867	0.000
15.944	0.000
16.022	0.000
16.100	0.000
16.178	0.000
16.255	0.000
16.333	0.000
16.411	0.000
16.489	0.000
16.567	0.000
16.644	0.000
16.722	0.000
16.800	0.000
16.878	0.000
16.955	0.000
17.033	0.000
17.111	0.000
17.189	0.000
17.267	0.000
17.344	0.000
17.422	0.000
17.500	0.000
17.578	0.000
17.655	0.000
17.733	0.000
17.811	0.000
17.889	0.000
17.967	0.000
18.044	0.000
18.122	0.000
18.200	0.000
18.278	0.000
18.355	0.000
18.433	0.000
18.511	0.000
18.589	0.000
18.667	0.000
18.744	0.000
18.822	0.000
18.900	0.000
18.978	0.000
19.055	0.000
19.133	0.000
19.211	0.000
19.289	0.000
19.367	0.000
19.444	0.000
19.522	0.000
19.600	0.000
19.678	0.000
19.755	0.000
19.833	0.000
19.911	0.000
19.989	0.000
20.067	0.000
20.144	0.000
20.222	0.000
20.300	0.000
20.378	0.000
20.455	0.000
20.533	0.000
20.611	0.000
20.689	0.000
20.767	0.000
20.844	0.000
20.922	0.000
21.000	0.000
21.078	0.000
21.155	0.000
21.233	0.000
21.311	0.000
21.389	0.000
21.467	0.000
21.544	0.000
21.622	0.000
21.700	0.000
21.778	0.000
21.855	0.000
21.933	0.000
22.011	0.000
22.089	0.000
22.167	0.000
22.244	0.000
22.322	0.000
22.400	0.000
22.478	0.000
22.555	0.000
22.633	0.000
22.711	0.000
22.789	0.000
22.867	0.000
22.944	0.000
23.022	0.000
23.100	0.000
23.178	0.000
23.255	0.000
23.333	0.000
23.411	0.000
23.489	0.000
23.567	0.000
23.644	0.000
23.722	0.000
23.800	0.000
23.878	0.000
23.955	0.000
24.033	0.000
24.111	0.000
24.189	0.000
24.267	0.000
24.344	0.000
24.422	0.000
24.500	0.000
24.578	0.000
24.655	0.000
24.733	0.000
24.811	0.000
24.889	0.000
24.967	0.000
25.044	0.000
25.122	0.000
25.200	0.000
25.278	0.000
25.355	0.000
25.433	0.000
25.511	0.000
25.589	0.000
25.667	0.000
25.744	0.000
25.822	0.000
25.900	0.000
25.978	0.000
26.055	0.000
26.133	0.000
26.211	0.000
26.289	0.000
26.367	0.000
26.444	0.000
26.522	0.000
26.600	0.000
26.678	0.000
26.755	0.000
26.833	0.000
26.911	0.000
26.989	0.000
27.067	0.000
27.144	0.000
27.222	0.000
27.300	0.000
27.378	0.000
27.455	0.000
27.533	0.000
27.611	0.000
27.689	0.000
27.767	0.000
27.844	0.000
27.922	0.000
28.000	0.000
28.078	0.000
28.155	0.000
28.233	0.000
28.311	0.000
28.389	0.000
28.467	0.000
28.544	0.000
28.622	0.000
28.700	0.000
28.778	0.000
28.855	0.000
28.933	0.000
29.011	0.000
29.089	0.000
29.167	0.000
29.244	0.000
29.322	0.000
29.400	0.000
29.478	0.000
29.555	0.000
29.633	0.000
29.711	0.000
29.789	0.000
29.867	0.000
29.944	0.000
30.022	0.000
30.100	0.000
30.178	0.000
30.255	0.000
30.333	0.000
30.411	0.000
30.489	0.000
30.567	0.000
30.644	0.000
30.722	0.000
30.800	0.000
30.878	0.000
30.955	0.000
31.033	0.000
31.111	0.000
31.189	0.000
31.267	0.000
31.344	0.000
31.422	0.000
31.500	0.000
31.578	0.000
31.655	0.000
31.733	0.000
31.811	0.000
31.889	0.000

CALCULATED C/CO VERSUS THETA VALUES
FOR DISPERSION MODEL

THEORETICAL DETENTION		ACTUAL DETENTION	
0.100	0.002	0.100	0.000
0.200	0.015	0.200	0.007
0.300	0.035	0.300	0.023
0.400	0.073	0.400	0.068
0.500	0.125	0.500	0.135
0.600	0.194	0.600	0.229
0.700	0.285	0.700	0.359
0.800	0.396	0.800	0.522
0.900	0.516	0.900	0.724
1.000	0.632	1.000	0.900
1.100	0.740	1.100	1.050
1.200	0.838	1.200	1.181
1.300	0.929	1.300	1.293
1.400	1.013	1.400	1.387
1.500	1.090	1.500	1.462
1.600	1.161	1.600	1.521
1.700	1.225	1.700	1.567
1.800	1.283	1.800	1.601
1.900	1.335	1.900	1.625
2.000	1.381	2.000	1.640
2.100	1.421	2.100	1.647
2.200	1.456	2.200	1.646
2.300	1.485	2.300	1.638
2.400	1.509	2.400	1.623
2.500	1.528	2.500	1.602
2.600	1.542	2.600	1.576
2.700	1.552	2.700	1.545
2.800	1.558	2.800	1.510
2.900	1.560	2.900	1.471
3.000	1.559	3.000	1.429

Tracer Response Analysis, 2.0 M, At 37.2 L/min.

TRACER RESPONSE ANALYSIS

ROTATING BIOLOGICAL CONTACTOR
HYDRAULIC CHARACTERIZATION

TEST METHOD USING A PULSE INPUT OF RODAMINE WT DYE

REACTOR OPERATION AND TEST CONDITIONS

VOLUME OF REACTOR	=	3468.40 LITRES
HYDRAULIC LOADING	=	37.20 LITRES/MIN
THEORETICAL DET	=	93.24 MIN
DYE INJECTION	=	0.0013 LITRES
CONC OF DYE ADDED	=	0.238E+03 PPB
DYE / TANK VOLUME	=	85.77 PPB

TEST RESULTS AND CALCULATED VALUES

DYE PEAK TIME	=	74.00 MIN
PEAK/THEOR DET	=	0.794

PEAK/MEAN DYE RES	=	0.725 MIN
MEAN DYE RESIDENCE	=	102.00 MIN
PER DYE RECOVERY	=	91.973%
FR. STAGNANT ZONE	=	-0.094

CSTR S IN SERIES USING THEORETICAL RES.	=	4.85
CSTR S IN SERIES USING MEAN DYE RES.	=	3.64
D/UL VALUE USING THEORETICAL RESIDENCE	=	0.8935E-01
D/UL VALUE USING MEAN DYE RESIDENCE	=	0.1227E+00

EXPERIMENTAL RESULTS C/CO VERSUS THETA

THETA	C/CO
0.064	0.000
0.129	0.000
0.193	0.002
0.257	0.015
0.322	0.048
0.386	0.148
0.450	0.347
0.515	0.497
0.579	0.553
0.644	0.615
0.708	0.651
0.772	0.673
0.837	0.675
0.901	0.663
0.965	0.646
1.030	0.625
1.094	0.607
1.158	0.575
1.223	0.546
1.287	0.513
1.351	0.484
1.416	0.457
1.480	0.416
1.544	0.385
1.609	0.356
1.673	0.327
1.738	0.300
1.802	0.273
1.866	0.248
1.931	0.228
2.015	0.199
2.100	0.174
2.184	0.154
2.269	0.130
2.353	0.122

CALCULATED C/CO VERSUS THETA VALUES
FOR CSTR IN SERIES MODEL

THEORETICAL DETENTION		ACTUAL DETENTION	
THETA	C/CO	THETA	C/CO
0.000	0.000	0.000	0.000
0.050	0.991	0.050	0.994
0.100	0.988	0.100	0.989
0.150	0.981	0.150	0.983
0.200	0.977	0.200	0.979
0.250	0.976	0.250	0.978
0.300	0.973	0.300	0.975
0.350	0.969	0.350	0.971
0.400	0.965	0.400	0.967
0.450	0.963	0.450	0.964
0.500	0.962	0.500	0.962
0.550	0.961	0.550	0.961
0.600	0.960	0.600	0.960
0.650	0.959	0.650	0.959
0.700	0.958	0.700	0.958
0.750	0.957	0.750	0.957
0.800	0.956	0.800	0.956
0.850	0.955	0.850	0.955
0.900	0.954	0.900	0.954
0.950	0.953	0.950	0.953
1.000	0.952	1.000	0.952
1.100	0.951	1.100	0.951
1.200	0.950	1.200	0.950
1.300	0.949	1.300	0.949
1.400	0.948	1.400	0.948
1.500	0.947	1.500	0.947
1.600	0.946	1.600	0.946
1.700	0.945	1.700	0.945
1.800	0.944	1.800	0.944
1.900	0.943	1.900	0.943
2.000	0.942	2.000	0.942
2.100	0.941	2.100	0.941
2.200	0.940	2.200	0.940
2.300	0.939	2.300	0.939
2.400	0.938	2.400	0.938
2.500	0.937	2.500	0.937
2.600	0.936	2.600	0.936
2.700	0.935	2.700	0.935
2.800	0.934	2.800	0.934
2.900	0.933	2.900	0.933
3.000	0.932	3.000	0.932
3.100	0.931	3.100	0.931
3.200	0.930	3.200	0.930
3.300	0.929	3.300	0.929
3.400	0.928	3.400	0.928
3.500	0.927	3.500	0.927
3.600	0.926	3.600	0.926
3.700	0.925	3.700	0.925
3.800	0.924	3.800	0.924
3.900	0.923	3.900	0.923
4.000	0.922	4.000	0.922
4.100	0.921	4.100	0.921
4.200	0.920	4.200	0.920

CALCULATED C/CO VERSUS THETA VALUES
FOR DISPERSION MODEL

THEORETICAL DETENTION		ACTUAL DETENTION	
0.100	0.030	0.100	0.000
0.200	0.001	0.200	0.007
0.300	0.041	0.300	0.127
0.400	0.249	0.400	0.427
0.500	0.536	0.500	0.739
0.600	0.931	0.600	0.987
0.700	1.133	0.700	1.085
0.800	1.185	0.800	1.063
0.900	1.123	0.900	0.978
1.000	0.994	1.000	0.859
1.100	0.839	1.100	0.731
1.200	0.634	1.200	0.607
1.300	0.543	1.300	0.496
1.400	0.422	1.400	0.400
1.500	0.324	1.500	0.320
1.600	0.245	1.600	0.254
1.700	0.184	1.700	0.200
1.800	0.137	1.800	0.157
1.900	0.101	1.900	0.123
2.000	0.074	2.000	0.095
2.100	0.055	2.100	0.074
2.200	0.040	2.200	0.058
2.300	0.029	2.300	0.045
2.400	0.021	2.400	0.035
2.500	0.015	2.500	0.027
2.600	0.011	2.600	0.021
2.700	0.008	2.700	0.016
2.800	0.006	2.800	0.012
2.900	0.004	2.900	0.009
3.000	0.003	3.000	0.007

Tracer Response Analysis, 2.0 M, At 100.6 L/min.

TRACER RESPONSE ANALYSIS

ROTATING BIOLOGICAL CONTACTOR
HYDRAULIC CHARACTERIZATION~~TEST METHOD USING A PULSE INPUT OF RODAMINE WT DYE~~

REACTOR OPERATION AND TEST CONDITIONS

VOLUME OF REACTOR	=	35.5.60 LITRES
HYDRAULIC LOADING	=	100.56 LITRES/MIN
THEORETICAL DET	=	35.26 MIN
DYE INJECTION	=	0.0013 LITRES
CONC OF DYE ADDED	=	0.238E+03 PPB
DYE / TANK VOLUME	=	83.91 PPB

TEST RESULTS AND CALCULATED VALUES

DYE PEAK TIME	=	30.50 MIN
PEAK/THEOR DET	=	0.865

PEAK/MEAN DYE RES	=	0.752 MIN
MEAN DYE RESIDENCE	=	40.00 MIN
PER DYE RECOVERY	=	91.351%
FR. STAGNANT ZONE	=	-0.134

CSTR S IN SERIES USING THEORETICAL RES.	=	7.41
CSTR S IN SERIES USING MEAN DYE RES.	=	4.21
D/UL VALUE USING THEORETICAL RESIDENCE	=	0.6257E-01
D/UL VALUE USING MEAN DYE RESIDENCE	=	0.1326E+00

EXPERIMENTAL RESULTS C/CO VERSUS THETA

THETA	C/CO
0.085	0.000
0.170	0.010
0.255	0.061
0.340	0.151
0.425	0.334
0.511	0.454
0.596	0.546
0.681	0.622
0.766	0.664
0.851	0.689
0.936	0.687
1.021	0.668
1.106	0.643
1.191	0.601
1.276	0.553
1.361	0.495
1.446	0.440
1.532	0.394
1.617	0.360
1.702	0.313
1.787	0.277
1.872	0.245
1.957	0.224
2.042	0.196
2.127	0.173
2.212	0.152
2.297	0.131
2.382	0.113
2.467	0.095
2.553	0.080
2.636	0.063
2.721	0.050
2.803	0.044
2.887	0.035

CALCULATED C/CO VERSUS THETA VALUES
FOR CSTR IN SERIES MODFL

THEORETICAL DETENTION		ACTUAL DETENTION	
THETA	C/CO	THETA	C/CO
0.0000	0.0000	0.0000	0.0000
0.0050	0.0000	0.0050	0.0000
0.0100	0.0011	0.0100	0.0029
0.0150	0.0025	0.0150	0.0073
0.0200	0.0049	0.0200	0.0153
0.0250	0.0092	0.0250	0.0247
0.0300	0.0131	0.0300	0.0351
0.0350	0.0185	0.0350	0.0451
0.0400	0.0247	0.0400	0.0551
0.0450	0.0310	0.0450	0.0643
0.0500	0.0374	0.0500	0.0722
0.0550	0.0440	0.0550	0.0787
0.0600	0.0500	0.0600	0.0836
0.0650	0.0562	0.0650	0.0870
0.0700	0.0622	0.0700	0.0890
0.0750	0.0688	0.0750	0.0896
0.0800	0.0759	0.0800	0.0890
0.0850	0.0824	0.0850	0.0874
0.0900	0.0886	0.0900	0.0850
0.0950	0.0943	0.0950	0.0813
1.0000	0.0994	1.0000	0.0761
1.1000	0.0768	1.1000	0.0677
1.2000	0.0478	1.2000	0.0433
1.3000	0.0262	1.3000	0.0230
1.4000	0.0131	1.4000	0.0118
1.5000	0.0061	1.5000	0.0058
2.0000	0.0027	2.0000	0.0030
2.2000	0.0011	2.2000	0.0014
2.4000	0.0004	2.4000	0.0007
2.6000	0.0002	2.6000	0.0003
3.0000	0.0001	3.0000	0.0001
3.2000	0.0000	3.2000	0.0000
3.4000	0.0000	3.4000	0.0000
3.6000	0.0000	3.6000	0.0000
3.8000	0.0000	3.8000	0.0000
4.0000	0.0000	4.0000	0.0000
4.2000	0.0000	4.2000	0.0000

CALCULATED C/CO VERSUS THETA VALUES
FOR DISPERSION MODEL

THEORETICAL DETENTION		ACTUAL DETENTION	
0.100	0.000	0.100	0.000
0.200	0.000	0.200	0.002
0.300	0.007	0.300	0.072
0.400	0.102	0.400	0.325
0.500	0.393	0.500	0.676
0.600	0.804	0.600	0.964
0.700	1.147	0.700	1.112
0.800	1.313	0.800	1.129
0.900	1.299	0.900	1.034
1.000	1.164	1.000	0.929
1.100	0.972	1.100	0.787
1.200	0.769	1.200	0.648
1.300	0.586	1.300	0.522
1.400	0.433	1.400	0.413
1.500	0.312	1.500	0.323
1.600	0.221	1.600	0.250
1.700	0.154	1.700	0.193
1.800	0.106	1.800	0.147
1.900	0.072	1.900	0.112
2.000	0.049	2.000	0.089
2.100	0.033	2.100	0.064
2.200	0.022	2.200	0.048
2.300	0.015	2.300	0.036
2.400	0.010	2.400	0.027
2.500	0.006	2.500	0.020
2.600	0.004	2.600	0.015
2.700	0.003	2.700	0.011
2.800	0.002	2.800	0.008
2.900	0.001	2.900	0.006
3.000	0.001	3.000	0.005

```

C THE TWO CHIEF REFERENCES USED FOR THIS PROGRAMME ARE
C
C 1. LEVENSPIEL , CHEMICAL REACTION ENGINEERING , CHAPTER 9
C
C 2. TIMPANY , VARIATION IN AXIAL MIXING IN AN AERATION
C TANK. MASTERS THESIS, DEPT. OF CHEM ENG., MCMASTER
C UNIVERSITY , 1966.
C
C THE D/UL VALUE FOR THE DISPERSION MODEL IS SOLVED BY USING THE
C CORRELATIONS OF PEAK TIME VERSUS D/UL DEVELOPED BY TIMPANY. (PP 31 -
C
C THE CSTR IN SERIES MODEL IS SOLVED BY TAKING THE DERIVATIVE OF
C EQUATION 9-35 IN LEVENSPIEL - EQUATING THE RESULT TO ZERO AND
C SOLVING FOR THE NUMBER OF EQUAL TANKS IN SERIES, J , IN TERMS
C OF THETA. THETA IS FOUND BY DIVIDING THE PEAK DYE TIME BY THE
C THEORETICAL RESIDENCE TIME.
C
C THE C/CO VALUES FOR THE DISPERSION MODEL ARE SOLVED BY ITERATION
C USING EQUATION 8 IN CHAPTER 2 OF TIMPANY.
C
C THE C/CO VALUES FOR THE CSTR IN SERIES MODEL ARE SOLVED USING
C EQUATION 9-35 IN LEVENSPIEL FOR VARIOUS VALUES OF THETA.
C
C
C DIMENSION C(500),CUL(500),TANKS(5),AW(5),DULP(5),THETA(2,100)
C DIMENSION CCO(2,100),U(500),AMU(5,500),COCO(5,500),CCI(100)
C DIMENSION TTB(100),TBAR(2),RATIO(500),BETA(500),ETA(500)
C DIMENSION BLUE(100)
C DIMENSION CE(200)
C
C VOLT=TANK VOLUME IN LITRES VFLR=FLOW IN LITRES/MIN
C TPEAK=PEAK TIME IN MINUTES DYIN=AMOUNT OF DYE IN LITRES
C DYCON=CONC OF DYE IN PPB DT=MINUTES BETWEEN DATA PTS
C N=NUMBER OF DATA PTS
C C(I)=CONC OF DYE IN EFFLUENT PPB
C
C DO 545 IJKJ=1,3
C READ 1,VOLT,VFLR,TPEAK,DYIN,DYCON,DT
C PRINT 1,VOLT,VFLR,TPEAK,DYIN,DYCON,DT
C READ 2,N
C PRINT 2,N
C READ 3,(C(I),I=1,N)
C PRINT 3,(C(I),I=1,N)
C
C PERCENT DYE RECOVERY
C
C AMT= C(1)*DT*VFLR*10.**(-6)
C CUL(1) = AMT
C DO 100 I=2,N
C AMT=.5*(C(I)+C(I-1))*DT*VFLR*10.**(-6)
C LUL = I-1
C CUL(I) = CUL(LUL)+AMT
100 CONTINUE
C DYE = DYCON*DYIN*10.**(-6)
C PER = CUL(N)/DYE*100.
C
C CALCULATION OF MEAN RESIDENCE TIME OF THE TOTAL DYE RETRIEVED
C CALCULATION OF PERCENT STAGNANT ZONE
C

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TBAR(1) = VOLT/VFLR
ANT = 1.
I = 1
ZONE = CUL(N)/2.
201 IF(CUL(I).GT.ZONE) GO TO 200
ANT = ANT + 1.
I = I + 1
GO TO 201
200 TBAR(2) = ANT*DT
DEAD = (TBAR(1)-TBAR(2))/TBAR(1)
C
C CALCULATION NUMBER OF TANKS IN SERIES
C
TP1 = TPEAK/TBAR(1)
TP2 = TPEAK/TBAR(2)
TANKS(1) = 1./((TP1*(1./TP1-1.))
TANKS(2) = 1./((TP2*(1./TP2-1.))
C
C TRUNCATE TO NEAREST WHOLE NUMBER OF TANKS
C
AW(1) = TANKS(1)
AA = 1.5
203 IF(AW(1).LT.AA) GO TO 202
AA = AA + 1.
GO TO 203
202 AW(1) = AA - .5
AW(2) = TANKS(2)
AA = 1.5
205 IF(AW(2).LT.AA) GO TO 204
AA = AA + 1.
GO TO 205
204 AW(2) = AA - .5
C
C CALCULATION OF TAMPANY'S PEAK TIME D/UL VAULES
C
IF((TP1.GT.0.03).AND.(TP1.LT.0.3)) GO TO 206
IF((TP1.GT.0.3).AND.(TP1.LT.0.8)) GO TO 207
GO TO 208
206 DULP(1) = .2*(TP1**(-1.34))
GO TO 209
207 DULP(1) = 4.027*(10.**(-2.09*TP1))
GO TO 209
208 PRINT 300
IF(TP1.LE.0.03) GO TO 206
GO TO 207
209 CONTINUE
IF((TP2.GT.0.03).AND.(TP2.LE.0.3)) GO TO 210
IF((TP2.GT.0.3).AND.(TP2.LE.0.8)) GO TO 211
GO TO 213
210 DULP(2) = .2*(TP2**(-1.34))
GO TO 214
211 DULP(2) = 4.027*(10.**(-2.09*TP2))
GO TO 214
213 PRINT 300
IF(TP2.LE.0.03) GO TO 210
GO TO 211
214 CONTINUE
C
C CALCULATION OF C/CO VS THETA VALUES FOR GSTR MODELS

```



```

C   DERIVATIVE AT PEAK DYE CONC METHOD USED
C
DO 101 I=1,2
X=AW(I)**AW(I)
XX=1.
BB=1.
216 IF(AW(I).EQ.BB) GO TO 215
XX=XX*(AW(I)-BB)
BB=BB+1.
GO TO 216
215 FACT = XX
THETA(I,1) = 0.
DO 102 J=2,21
THETA(I,J) = THETA(I,J-1) + .05
102 CCO(I,J) = X/XX*THETA(I,J)**(AW(I)-1.)*EXP(-AW(I)*THETA(I,J))
DO 103 J= 22,37
THETA(I,J) = THETA(I,J-1) + .2
103 CCO(I,J) = X/XX*THETA(I,J)**(AW(I)-1.)*EXP(-AW(I)*THETA(I,J))
101 CONTINUE

```

```

C
C   CLACULATION OF ACTUAL C/CO VALUES FROM EXPERIMENTAL DATA
C

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```

CNOT = DYIN*DYCON/VOLT
MM = 0
DRAG = 0.
NN = 0
DO 104 I = 1,30
NN = NN + 3
MM = MM + 1
RATIO(MM) = C(NN)/CNOT
DRAG = DRAG + 3*DT
BETA(MM) = DRAG/TBAR(1)
104 ETA(MM) = DRAG/TBAR(2)
IM = (N- 90)/10 + 29
DO 105 I = 31, IM
MM = MM + 1
DRAG = DRAG + 10.*DT
NN = NN + 10
RATIO(MM) = C(NN)/CNOT
BETA(MM) = DRAG/TBAR(1)
ETA(MM) = DRAG/CNOT
105 CONTINUE

```

```

C
C   CALCULATION OF C/CO VALUES VS THETA FOR D/UL METHOD
C

```

```

C
M = 1
40 I=1
AMU(M,I)=1.4
U(M)=.5/DULP(M)
45 AMU(M,I)=AMU(M,I)-.001
FR = COS(AMU(M,I))/SIN(AMU(M,I))
FR = FR - AMU(M,I)*DULP(M) + .25/(AMU(M,I)*DULP(M))
IF(FR)45,45,50
50 AMU(M,I) = AMU(M,I) + .00001
FR = COS(AMU(M,I))/SIN(AMU(M,I))
FR = FR - AMU(M,I)*DULP(M) + .25/(AMU(M,I)*DULP(M))
IF(FR)55,50,50
55 AMU(M,I) = AMU(M,I) - .0000001

```

```

FR = COS(AMU(M,I))/SIN(AMU(M,I))
FR = FP - AMU(M,I)*DULP(M) + .25/(AMU(M,I)*DULP(M))
IF (FR)55,55,60
60 I= I + 1
AMU(M,I) = AMU(M,I-1)+3.1417
IF (I.LE.50) GO TO 45
M = M + 1
IF (M.LE.2) GO TO 40
DO 80 M=1,2
999 ZETA = 0.0
DO 70 K=1,30
ZETA = ZETA + .1
COCO(M,K) = 0.0
DO 65 I=1,50
A=2.0*AMU(M,I)*(U(M)*SIN(AMU(M,I)) + AMU(M,I)*COS(AMU(M,I)))
B=EXP(U(M)-((U(M)**2 + AMU(M,I)**2)/(2.0*U(M)))*ZETA)
D=U(M)**2 + 2.0*U(M) + AMU(M,I)**2
998 CE(I) = A*B/D
COCO(M,K) = COCO(M,K) + CE(I)
65 CONTINUE
67 CONTINUE
70 CONTINUE
80 CONTINUE
C
C PRINT INSTRUCTIONS AND DATA PRESENTATION FORMAT
C
PRINT 700
700 FORMAT(1H1,42X,24HTRACER RESPONSE ANALYSIS///)
PRINT 701
701 FORMAT(40X,29H FLUIDIZED BED)
PRINT 702
702 FORMAT(42X,26HHYDRAULIC CHARACTERIZATION///)
PRINT 703
703 FORMAT(30X,7TEST METHOD USING A PULSE INPUT OF RODAMINE WT DYE#,/)
PRINT 704
704 FORMAT(23X,37HREACTOR OPERATION AND TEST CONDITIONS///)
PRINT 705,VOLT
705 FORMAT(31X,20HVOLUME OF REACTOR = ,F7.2,6HLITRES)
PRINT 706,VFLR
706 FORMAT(31X,20HHYDRAULIC LOADING = ,F7.2,10HLITRES/MIN)
PRINT 707,TBAR(1)
707 FORMAT(31X,20HTHEORETICAL DET = ,F7.2,3HMIN)
PRINT 708,DYIN
708 FORMAT(31X,20HDYE INJECTION = ,F7.4,6HLITRES)
PRINT 709,DYCON
709 FORMAT(31X,20HCONC OF DYE ADDED = ,E10.3,3HPPB)
PRINT 710,CNOT
710 FORMAT(31X,20HDYE / TANK VOLUME = ,F7.2,3HPPB,///)
PRINT 711
711 FORMAT(23X,34HTEST RESULTS AND CALCULATED VALUES,///)
PRINT 712,IPEAK
712 FORMAT(31X,20HDYE PEAK TIME = ,F7.2,3HMIN)
PRINT 713,IP1
713 FORMAT(31X,20HPEAK/THEOR DET = ,F7.3/)
PRINT 714,IP2
714 FORMAT(31X,20HPEAK/MEAN DYE RES = ,F7.3,3HMIN)
PRINT 715,TBAR(2)
715 FORMAT(31X,7MEAN DYE RESIDENCE =#,F7.2,#MIN#)
PRINT 716,PER

```



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716 FORMAT(31X,20HPEP DYE RECOVERY = ,F7.3,1H*)
PRINT 717,DEAD
717 FORMAT(31X,20HFR. STAGNANT ZONE = ,F7.3,/)
PRINT 718,TANKS(1)
718 FORMAT(31X,42HCSTR S IN SERIES USING THEORETICAL RES. = ,F7.2)
PRINT 719,TANKS(2)
719 FORMAT(31X,42HCSTR S IN SERIES USING MEAN DYE RES. = ,F7.2)
PRINT 720,CULP(1)
720 FORMAT(31X,42HD/UL VALUE USING THEORETICAL RESIDENCE = ,E11.4)
PRINT 721,DULP(2)
721 FORMAT(31X,42HD/UL VALUE USING MEAN DYE RESIDENCE = ,E11.4)
PRINT 722
722 FORMAT(31X,38HEXPERIMENTAL RESULTS C/CO VERSUS THETA,/////)
PRINT 723
723 FORMAT(14X,5HTHETA,15X,4HC/CO,////)
PRINT 724,(BETA(I),RATIO(I),I=1,MM)
724 FORMAT(15X,F5.3,15X,F5.3)
PRINT 725
725 FORMAT(31X,35HCALCULATED C/CO VERSUS THETA VALUES)
PRINT 726
726 FORMAT(37X,24HFOR CSTR IN SERIES MODEL,/////)
PRINT 727
727 FORMAT(12X,#THEORETICAL DETENSION          ACTUAL DETENSION#,//)
PRINT 728
728 FORMAT(15X,5HTHETA,6X,4HC/CO,15X,5HTHETA,6X,4HC/CO,////)
PRINT 729,(THETA(1,J),CCO(1,J),THETA(2,J),CCO(2,J),J=1,37)
729 FORMAT(15X,F5.3,6X,F5.3,15X,F5.3,6X,F5.3)
PRINT 725
PRINT 730
730 FORMAT(26X,20HFOR DISPERSION MODEL,/////)
PRINT 727
BLUE(1)= .1
DO-731 K=2,30
BLUE(K)= BLUE(K-1) + .1
731 CONTINUE
PRINT 729,(BLUE(K),COCO(1,K),BLUE(K),COCO(2,K),K=1,30)
1 FORMAT(4F10.4,E10.2,F10.4)
2 FORMAT(I10)
3 FORMAT(2X,5F10.3)
300 FORMAT(10X,44HPEAK TIME OUTSIDE LIMIT FOR D/UL CALCULATION)
545 CONTINUE
STOP
END
FINIS
$X,LGO

```


APPENDIX E
ABBREVIATIONS AND SYMBOLS

Abbreviations and symbols appearing in this report are defined within the text, with the exception of those used in Appendix A, "Data Listing". These are defined below:

Amb. AIR	- ambient air
ALK	- alkalinity
DO	- dissolved oxygen
DW	- flow of dilution water in L/m
EFC 0807	- effluent composite sample from 0800 hours to 0700 hours
EFG 1200	- effluent grab sample taken at 1200 hours
INT. AIR	- internal air i.e. air within the hood enclosing the RBC
RF	- flow of raw feed in L/m
RFC 0807	- raw feed composite sample taken between 0800 hours and 0700 hours
RFG 1200	- raw feed grab sample taken at 1200 hours
STG	- RBC stage
SS	- suspended solids
VSS	- volatile suspended solids
30 min settle	- 30 minute settling test in 1 litre graduated cylinder